TECHNISCHE UNIVERSITÄT DRESDEN

BioDIP Biopolis Dresden Imaging Platform

PoL Physics of Life TU Dresden

BioImage Analysis and Data Processing Workshop 2025
Charles University, Prague

Bio-Image Analysis with napari Plugins

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Re-using material from:

Robert Haase (Scads.AI Leipzig); Maleeha Hassan (Helmholtz AI Dresden); Johannes Soltwedel (PoL – TU Dresden);

DRESDEN concept

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Morning part:

- Installation and introduction to **napari**
- Loading images from OMERO with **napari-omero**
- Segmentation with Machine Learning using **micro-sam**
- Segmentation with Machine Learning using **napari-apoc**

Afternoon part:

- Feature Extraction and Multichannel Analysis with **napari-skimage-regionprops**
- Object Classification with Machine Learning using **napari-apoc** and **napari-clusters-plotter**
- Scientific Plotting with **seaborn**

Installation

- napari as a Python Library
- napari as a Bundle App
- Installing Plugins

Installing napari and napari plugins

For this course, an environment with containing napari and a few plugins should already be installed in the local computers (napari-intro-env or devbio-napari). They should work for most exercises, but we should create a new one for the latest napari-clusters-plotter changes. The instructions are provided here:

— https://biapol.github.io/BioImage-Analysis-and-Data-Processing-Workshop-2025/intro.html

The following slides are meant to clarify what these instructions mean. We will create the course environment together soon after.

Installing napari as a Python package

napari is a Python library

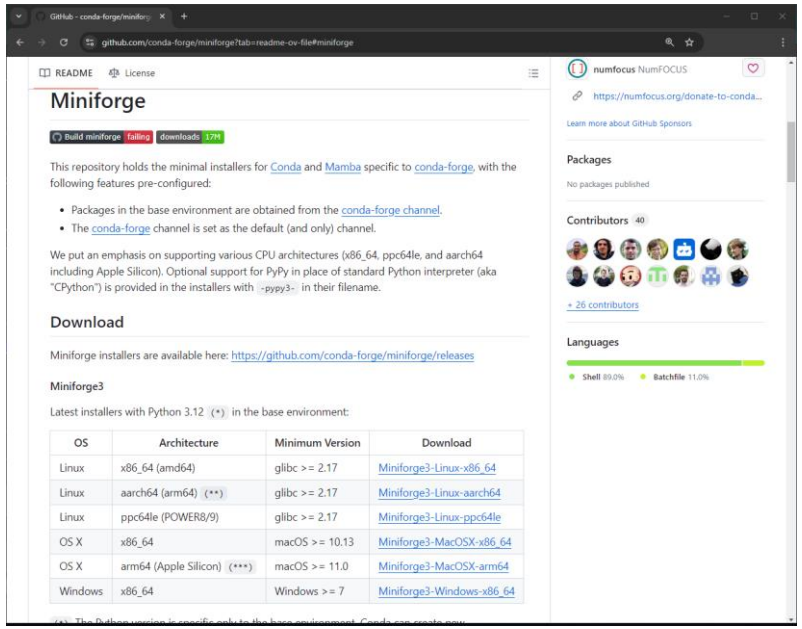
The recommended way to install napari is as a Python package

https://napari.org/stable/tutorials/fundamentals/installation.html#install-as-python-package-recommended

Your computer needs Python to run a Python package

— How do I install Python?

Install Miniforge!



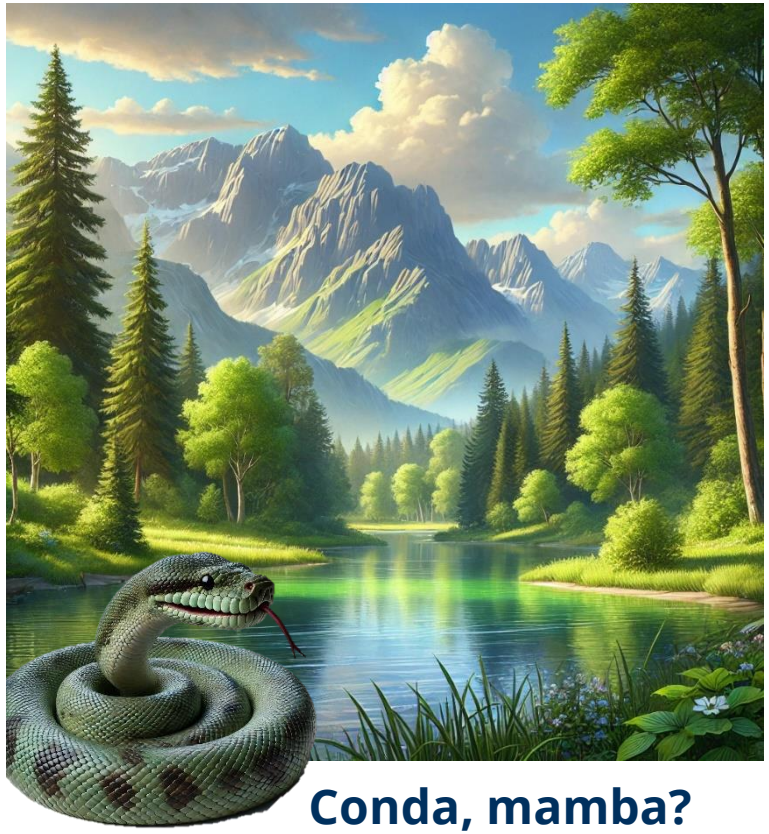
https://github.com/conda-forge/miniforge?tab=readme-ov-file#miniforge

- Miniforge is a minimal installer for `conda` and `mamba` (efficient conda in C++)
- `conda` is an environment and a package manager
- `conda` can create Virtual Enviroments and install Python (and other) packages, including napari

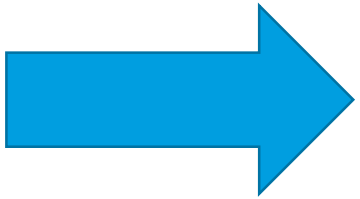
https://hackmd.io/@talley/SJB_lObBi#Terms

Installing napari as a Python package

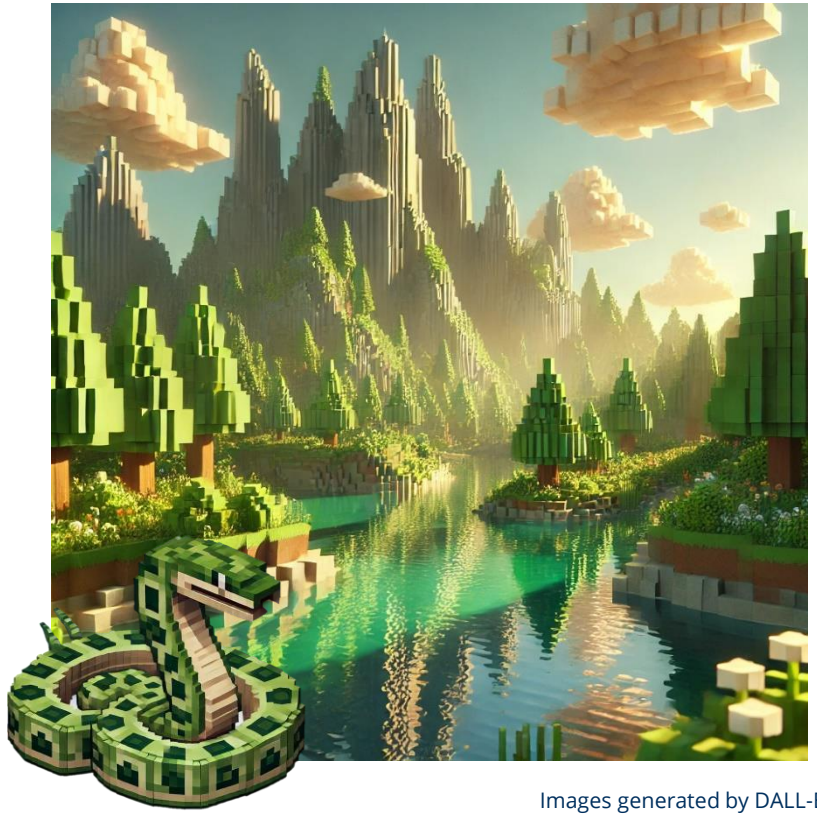
Wait, **environments ?**



Conda, mamba?



Yes, virtual environments!



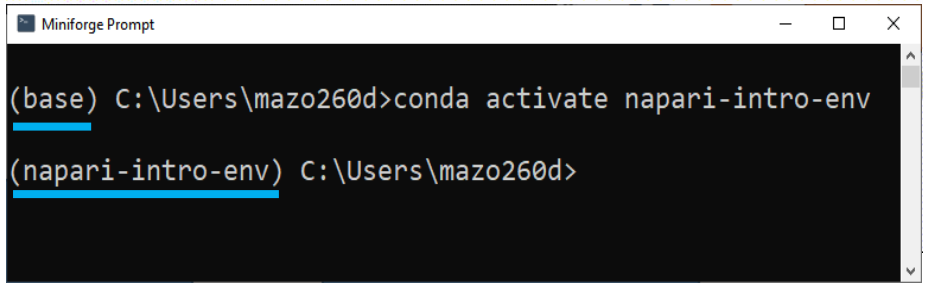
Images generated by DALL-E, developed by OpenAI.

Smiley icons from Flaticon.com

Installing napari as a Python package

Install it as a Python package in a Virtual Environment

— A virtual environment is an isolated collection of packages, settings, and an associated python interpreter



Commands:

```
mamba create -y -n napari-intro-env -c conda-forge python=3.10
mamba activate napari-intro-env
mamba install -c conda-forge napari pyqt
```

name channel package (version)

Some packages can only be installed with `pip`

- `conda` checks for version compatibility before installing
- `pip` tries to install, then lets you know if something went wrong
- `pip` cannot create environments

Installing napari

Napari can also be downloaded and installed like other software

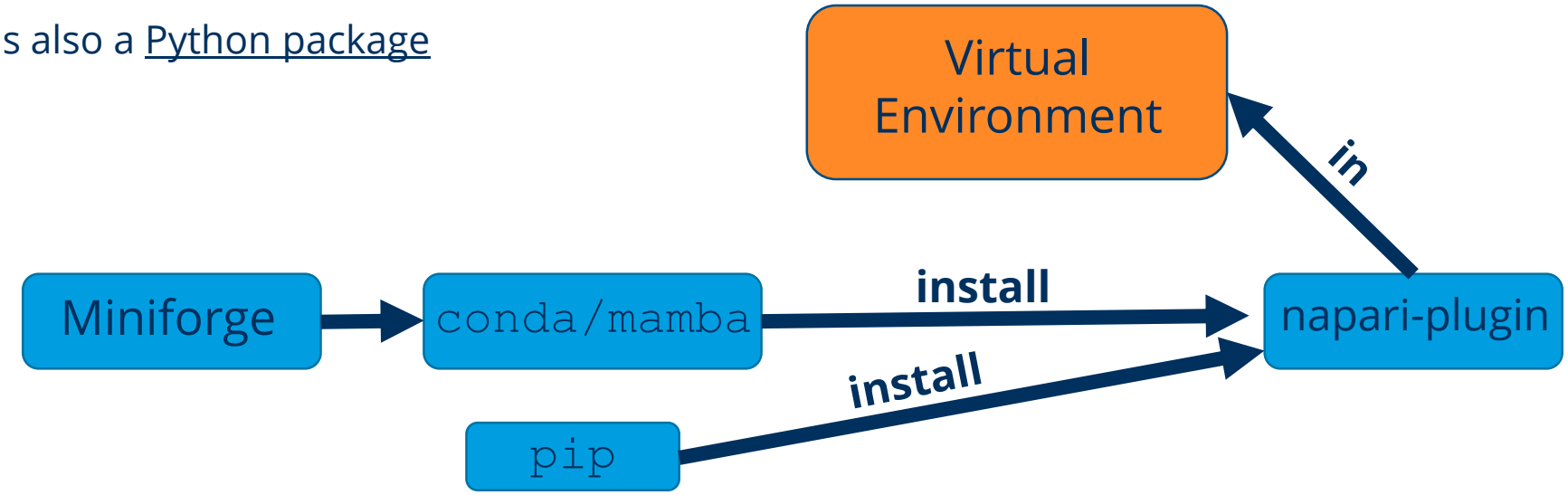
- https://napari.org/stable/tutorials/fundamentals/installation_bundle_conda.html#how-to-install-napari-as-a-bundled-app

This is more convenient, however it may be difficult to manage different plugin versions mid-long term

- In case of a dependency version mismatch, it may be necessary to uninstall and re-install the software and all the plugins again

Installing a plugin

A napari plugin is also a Python package



Commands:

mamba activate napari-intro-env

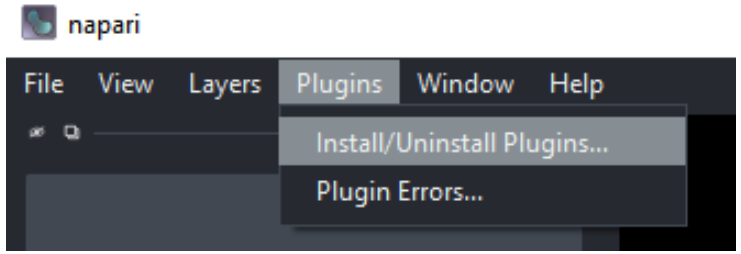
mamba install napari-plugin **OR** **pip** install napari-plugin

OR

conda install napari-plugin

Always check plugin installation instructions!

Via Plugins Menu:



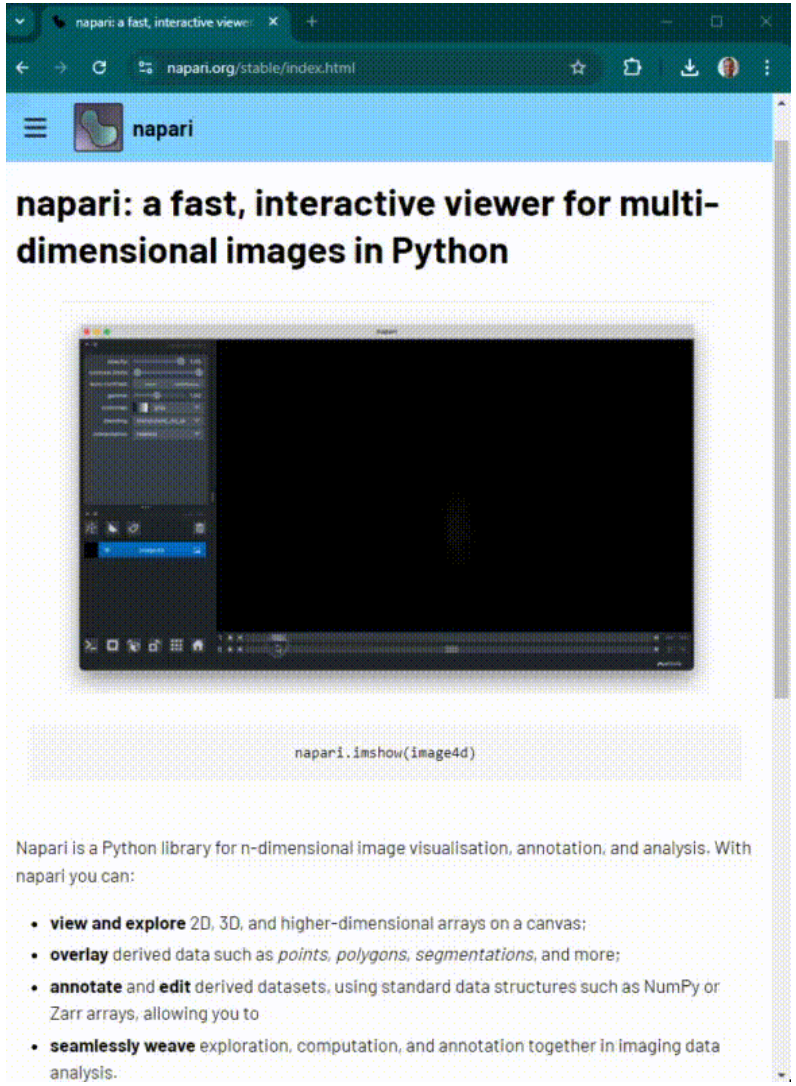
Introduction to napari

- napari Viewer
- Layer Types
- napari Plugins

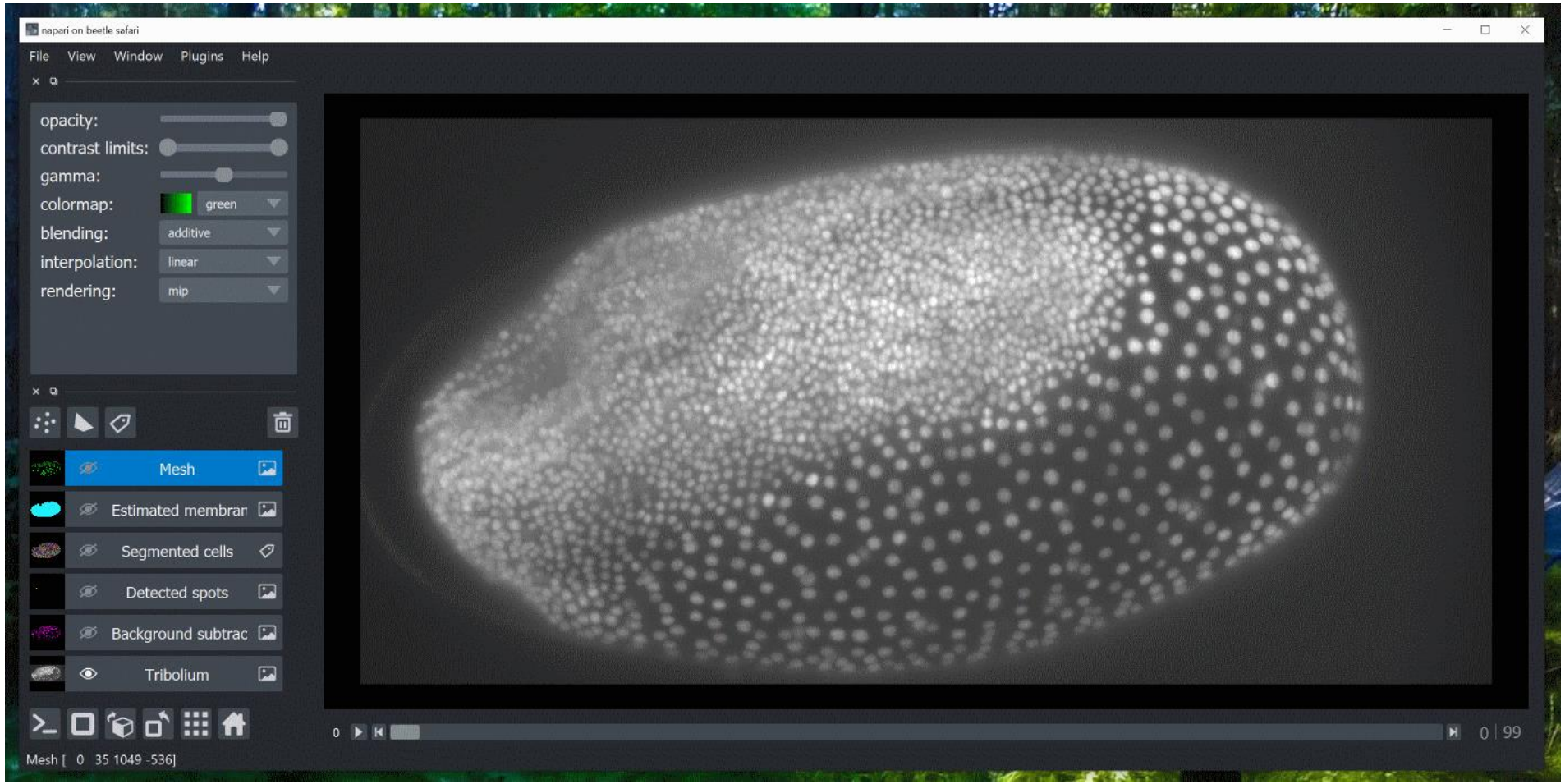
Napari: 3D viewer for Python

Multi-dimensional image viewer in python

https://napari.org/



Napari: 3D viewer for Python



https://napari.org/

Image data source: Daniela Vorkel, Myers lab, MPI-CBG/CSBD

Napari user interface

Menus

View configuration / tools

```
layer.opacity = 0.5
```

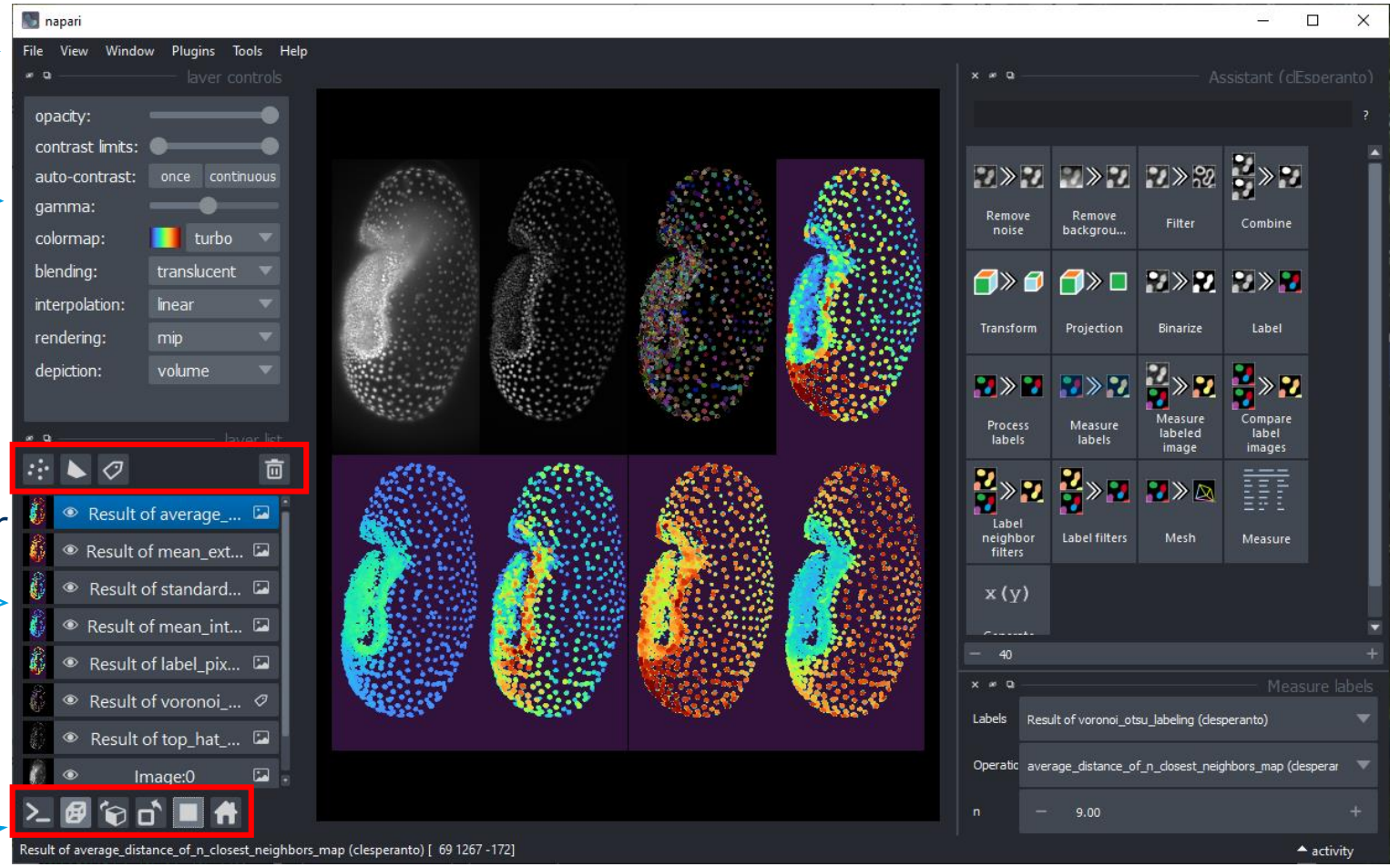
Add empty layers

Delete layer

Layers

```
layer.visible = False
```

Viewer controls



Dock widgets (custom plugins)

Open Python Console

2D/3D view

Change axes order

Transpose visible axes

Grid mode

Reset view

https://napari.org/tutorials/fundamentals/viewer.html

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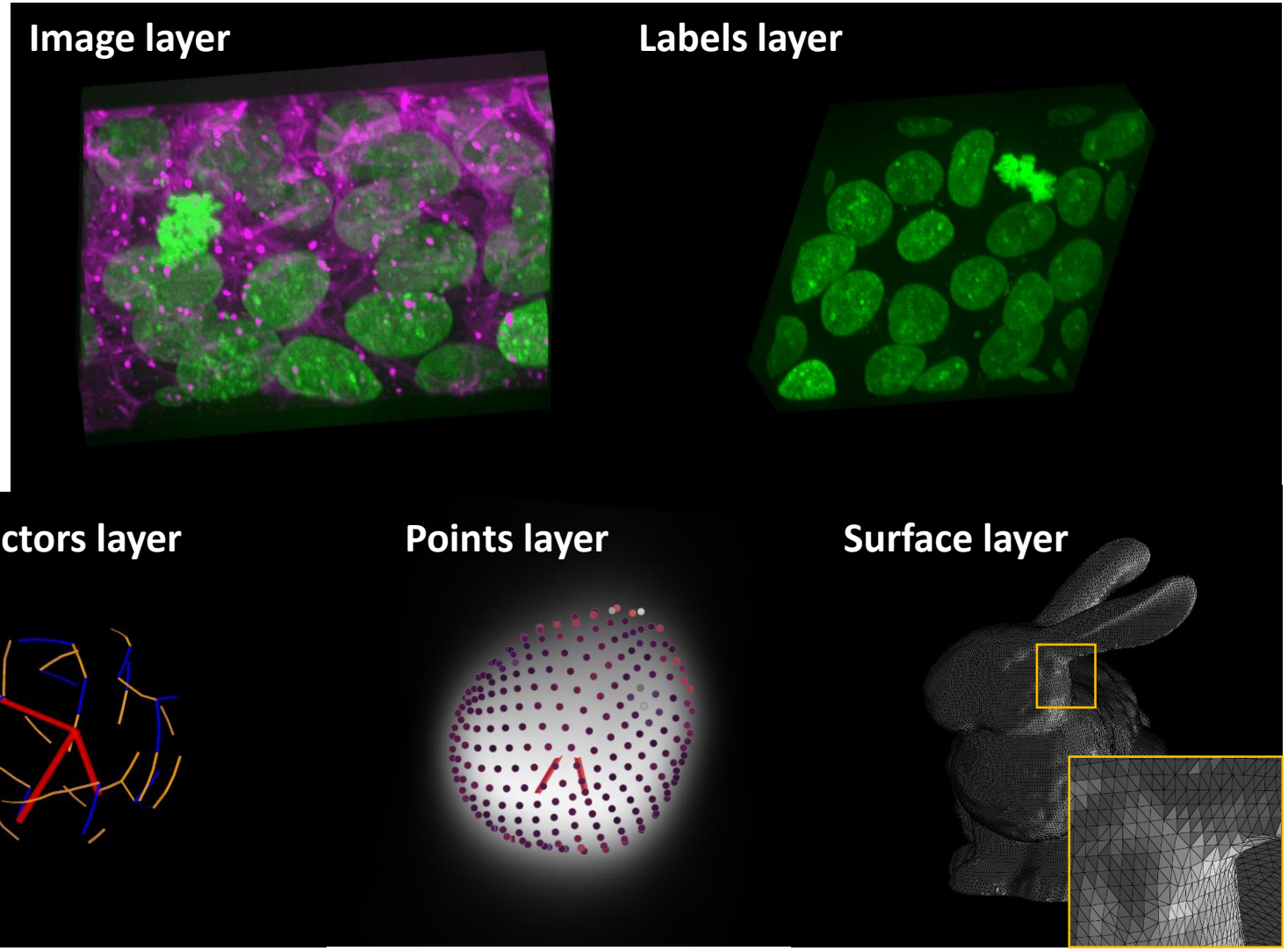
Layer types

- **Image layer:** Can be n-dimensional grayscale data (e.g., [CTZYX])
- **Labels layer:** Similar to image layer, but contains only integer numbers (e.g., 0,1,2,3,…)
- **Points layer:** List of coordinates in space
- **Vectors layer**: Direction from point A to point B
- **Surface layer:** Mesh (Vertices, Faces, Values)
- **Tracks layer:** Follow objects through time
- **Shapes layer**: Draw paths or regular shapes



https://github.com/quantumjot/arboretum licensed under MIT license

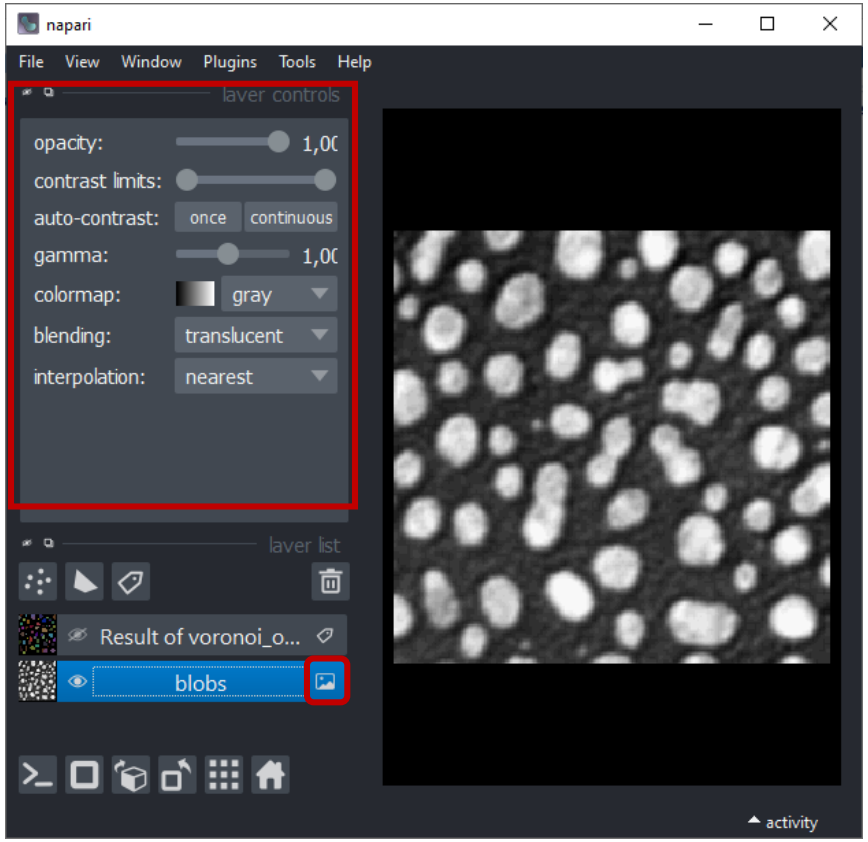
https://github.com/campaslab/napari-stress

https://gitlab.kitware.com/vtk/vtk

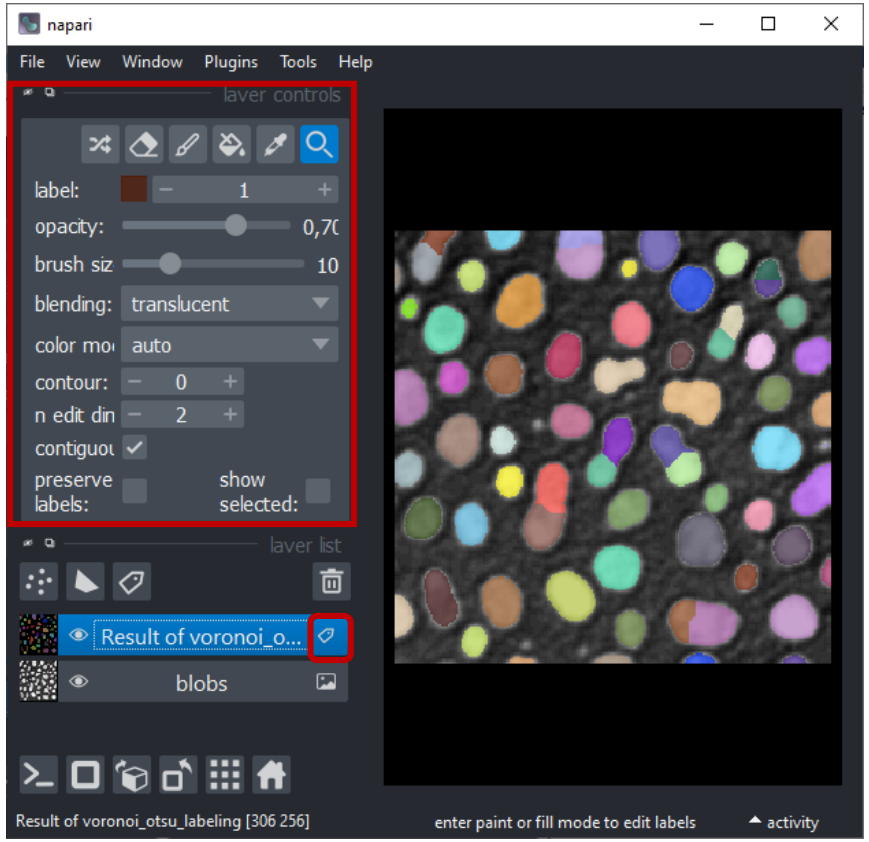
Layer types

Different layers have different tools and options

Image Layer



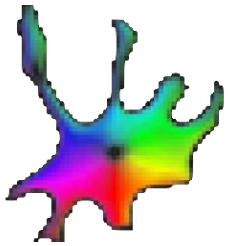
Labels Layer



Python image & data analysis tools are powerful!

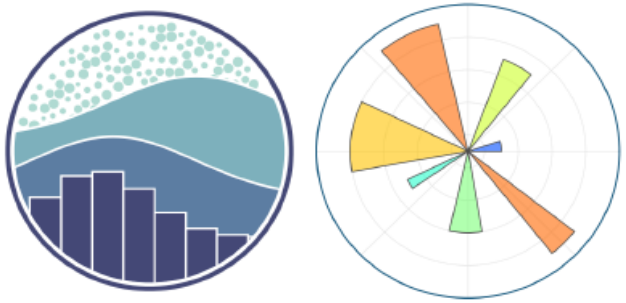
Python scientific image computing toolbox
https://github.com/scikit-image

Python machine-learning toolbox
https://github.com/scikit-learn



Cell segmentation
https://github.com/MouseLand/cellpose

Object detection with Stardist
https://github.com/stardist



Data visualization & exploration
https://github.com/matplotlib
https://github.com/seaborn

cle._

GPU-accelerated image processing
https://github.com/clEsperanto

Napari

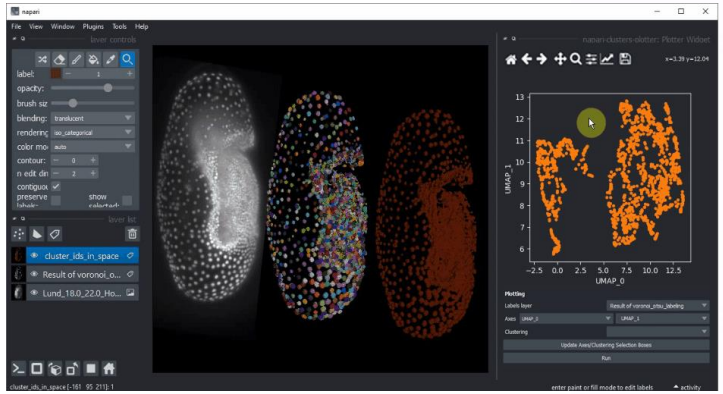
Existing functionality can be turned into plugins!

→ Interactivity

→ Automatic GUI generation

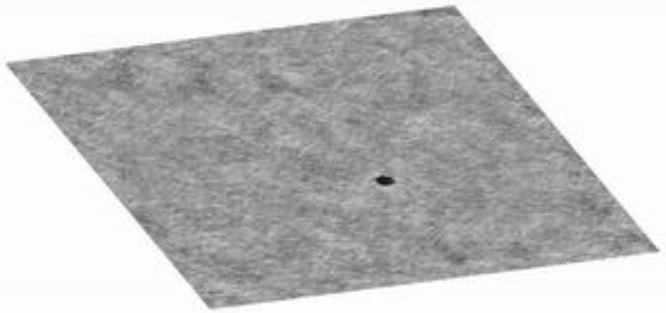
napari plugins

clusters-plotter



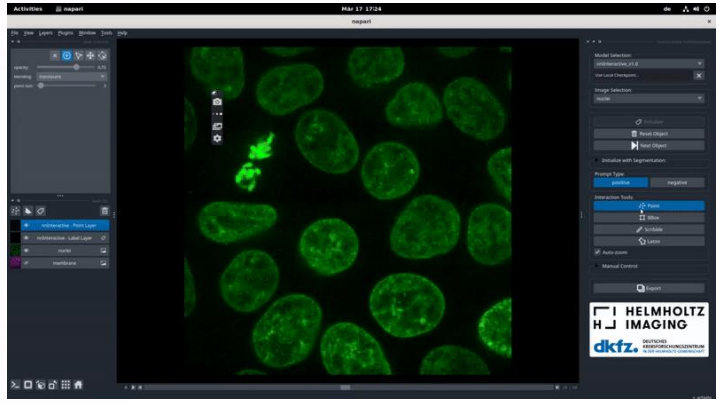
https://github.com/BiAPoL/napari-clusters-plotter

animation



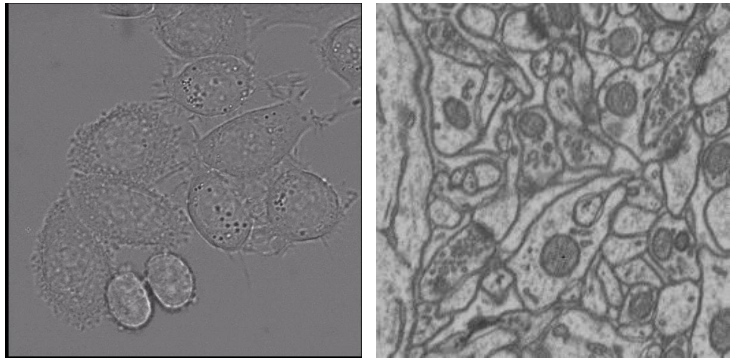
https://github.com/napari/napari-animation

segmentation



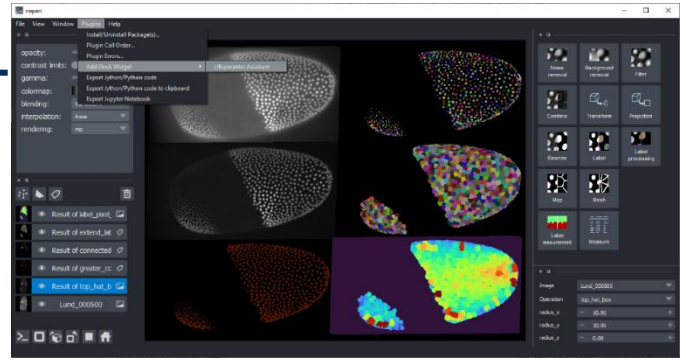
https://github.com/MIC-DKFZ/napari-nninteractive

micro-sam



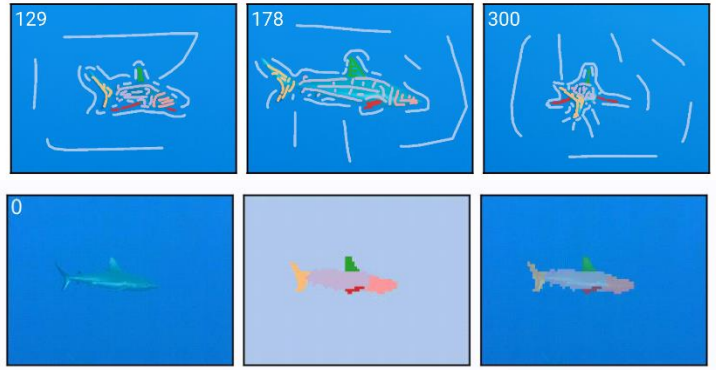
https://github.com/computational-cell-analytics/micro-sam

devbio-napari



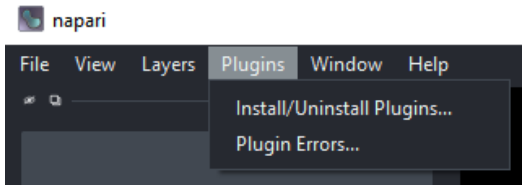
https://github.com/haesleinhuepf/devbio-napari

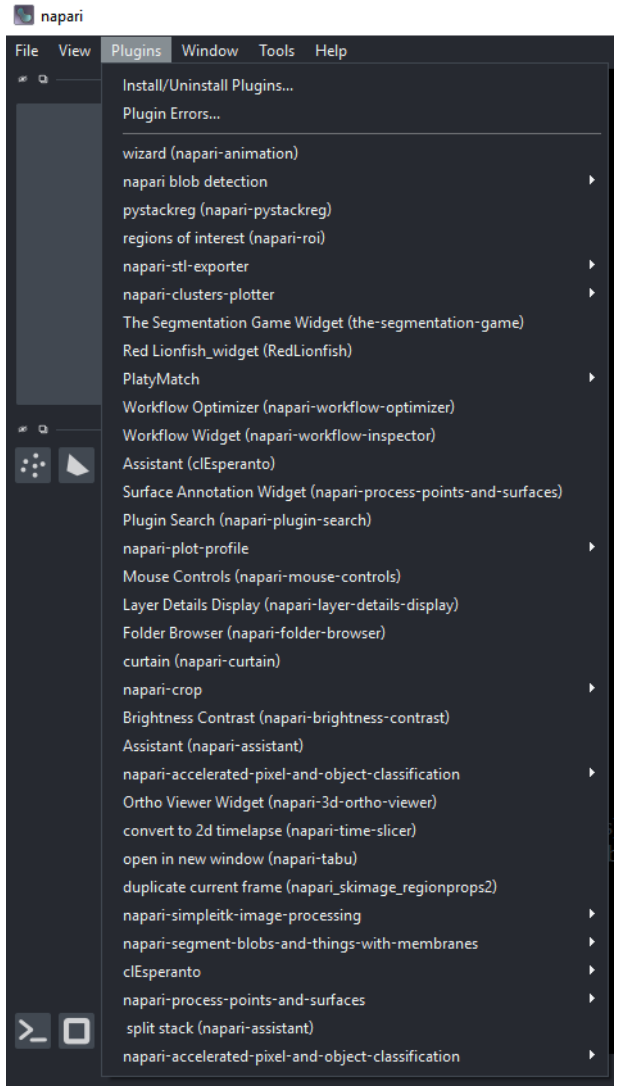
conv-paint

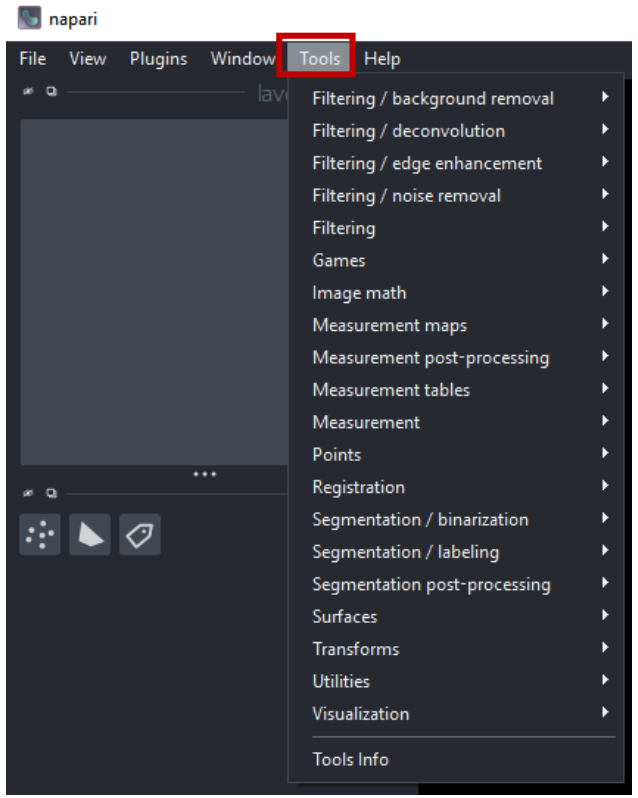


https://github.com/guiwitz/napari-convpaint

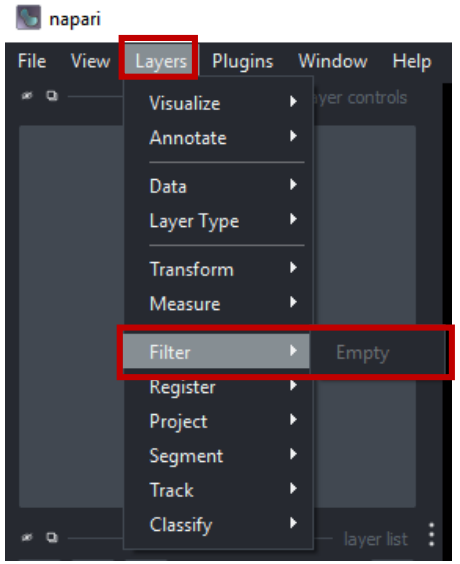
Plugins and Layers Menus







devbio-napari plugin bundle



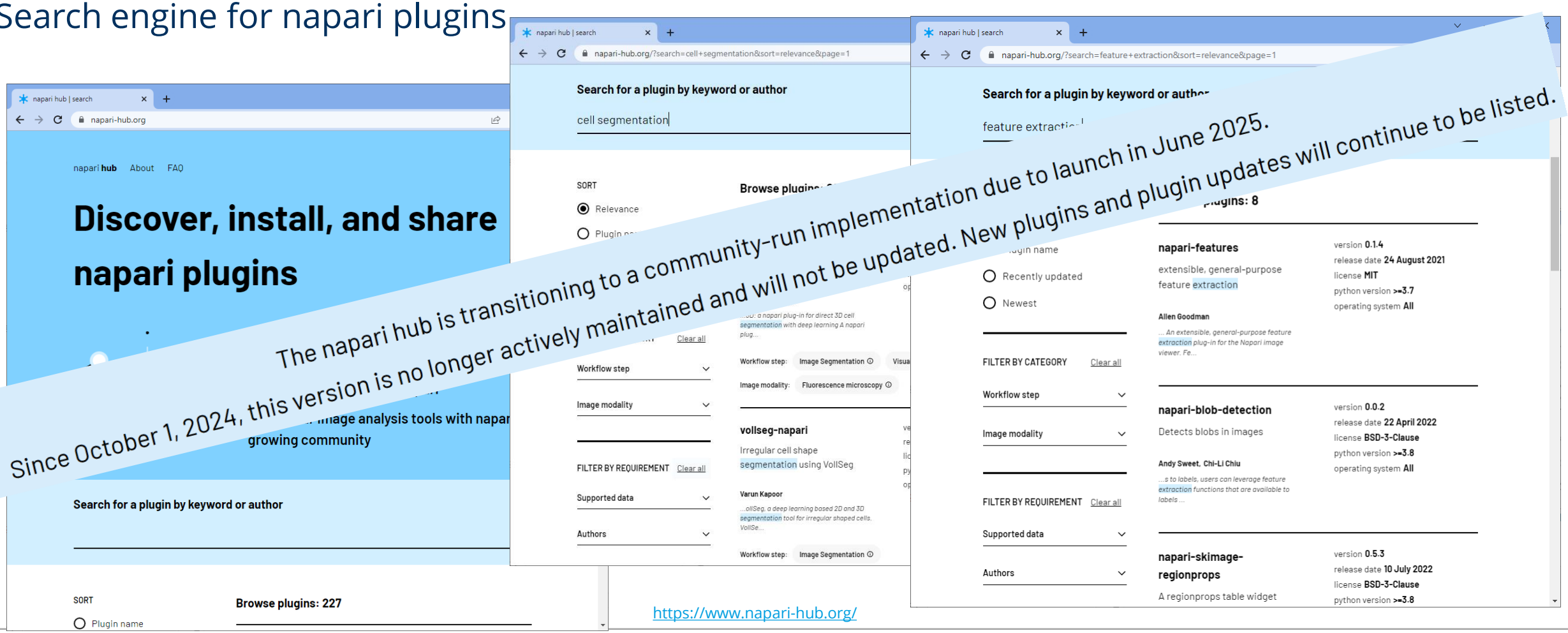
napari >= 0.5.0

Layers sub-menu items depend on plugin developers populating them

The Napari Hub

The plugin you are looking for may be near you!

Search engine for napari plugins



https://www.napari-hub.org/

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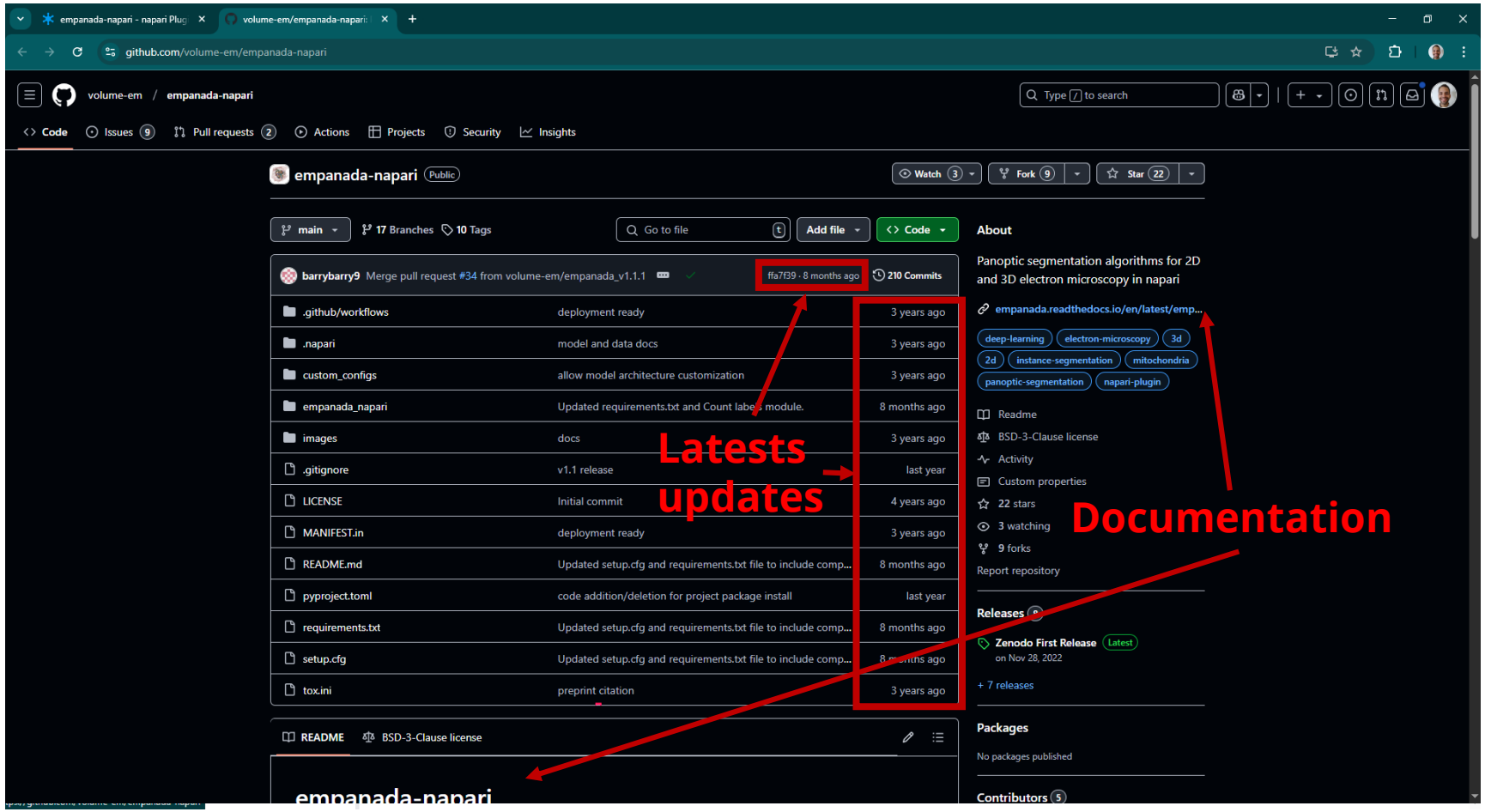
Inspecting plugin usage and maintenance - GitHub



Exercise: Create a conda environment for this course

Please follow the instructions in the link below to create a local environment:

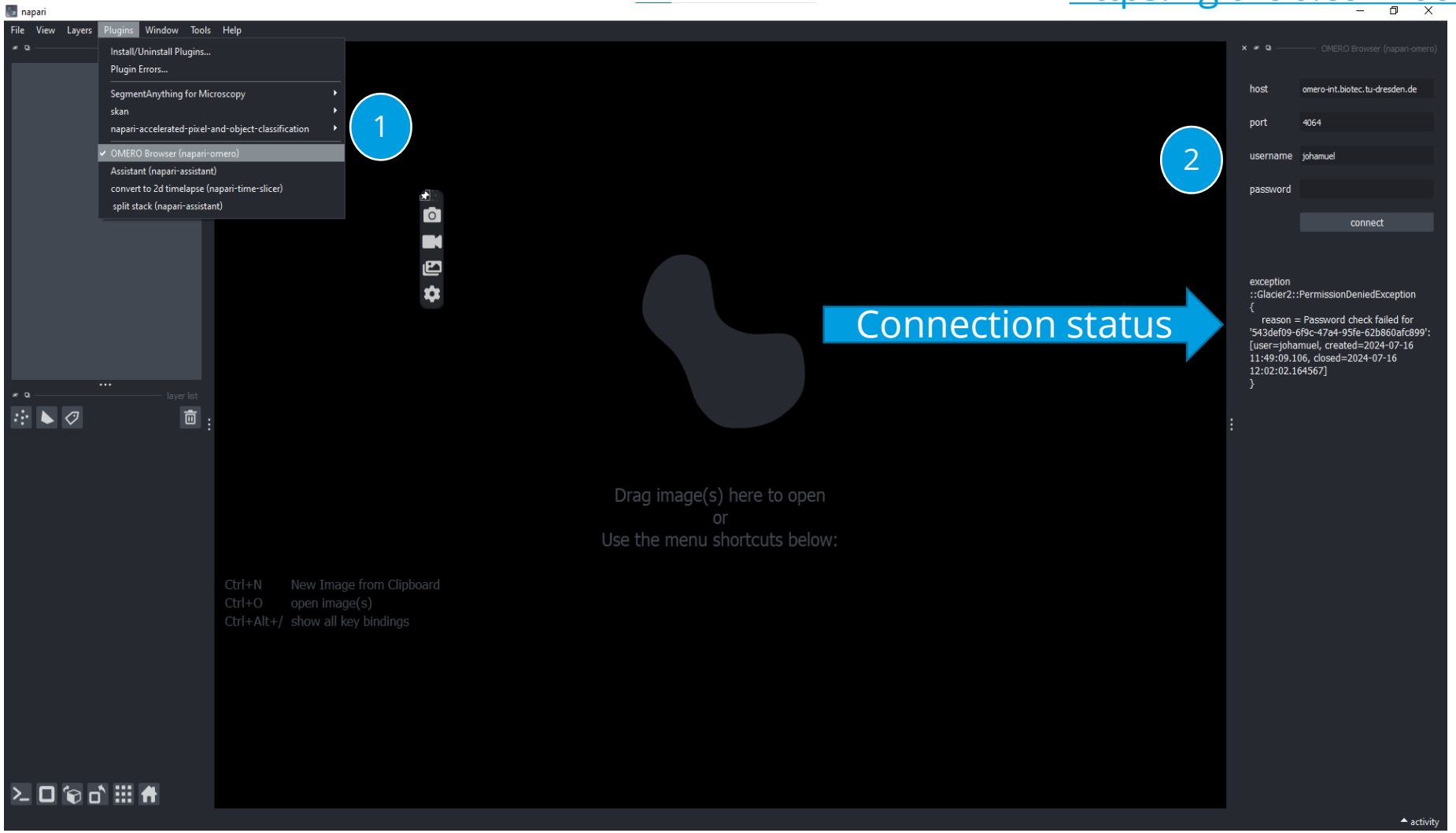
https://biapol.github.io/BioImage-Analysis-and-Data-Processing-Workshop-2025/intro.html

Napari – OMERO integration

- Demonstration: napari-OMERO plugin

Napari-OMERO usage

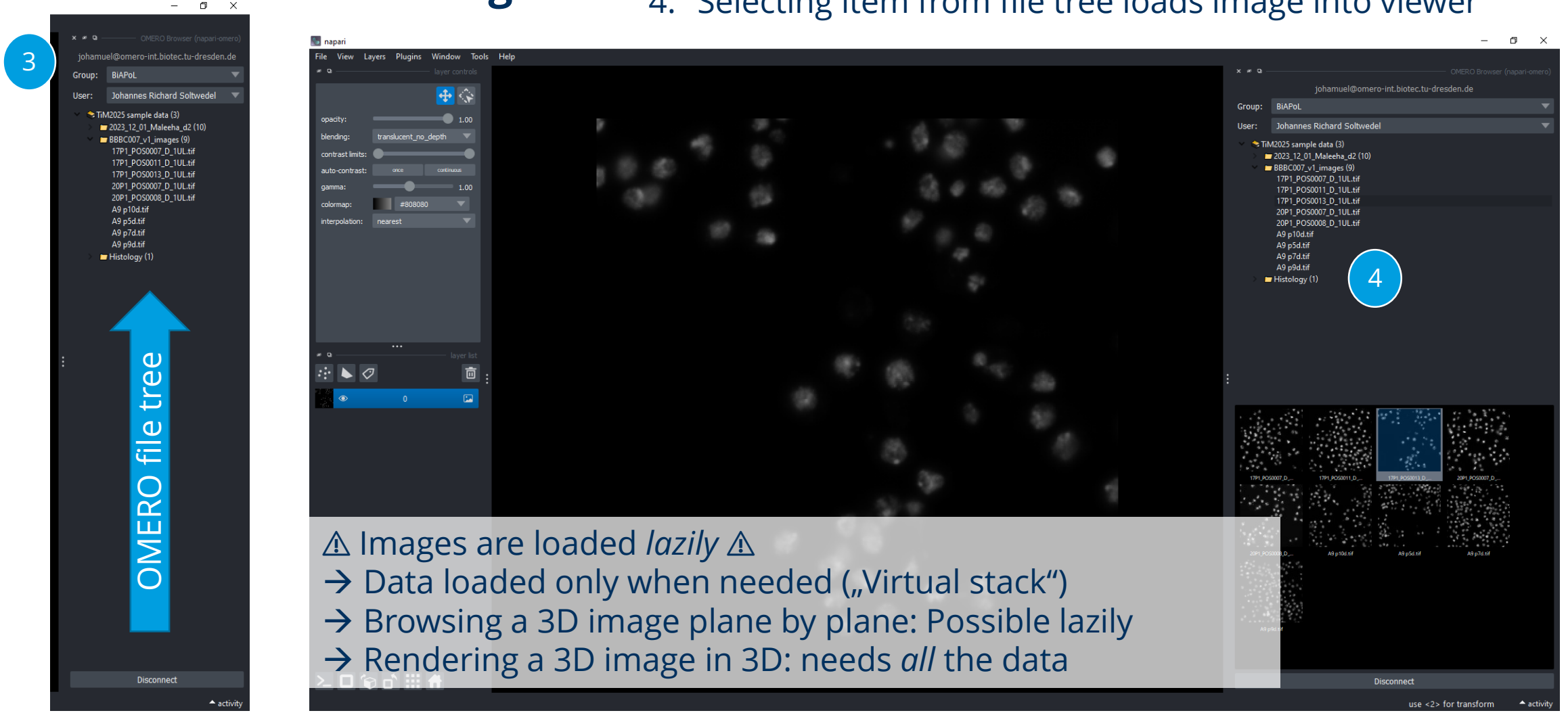
https://github.com/tlambert03/napari-omero



1. Select Plugin from plugin menu
2. Log into TiM OMERO server with your TiM credentials

Napari-OMERO usage

3. Select group and user
4. Selecting item from file tree loads image into viewer



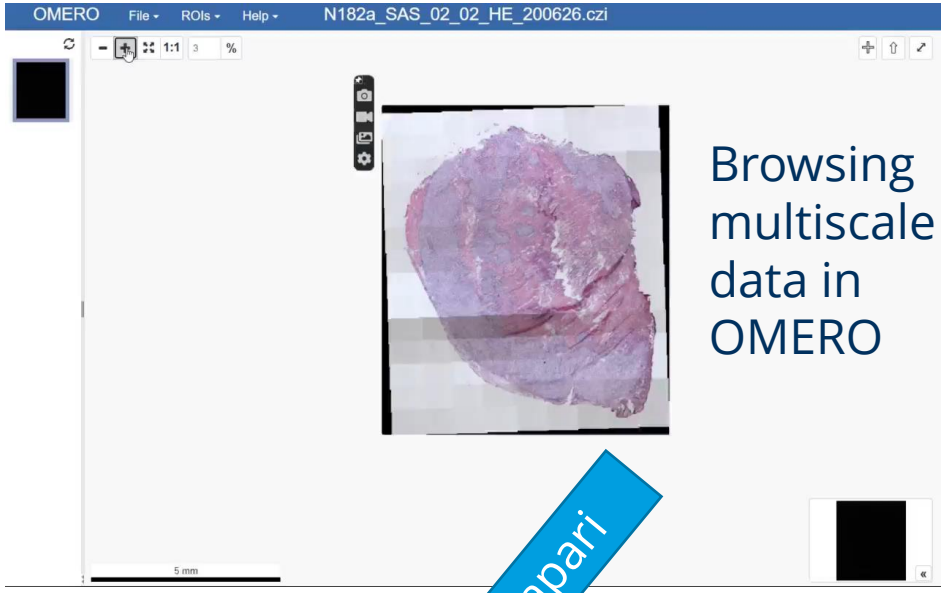
⚠ Images are loaded *lazily* ⚠
→ Data loaded only when needed („Virtual stack“)
→ Browsing a 3D image plane by plane: Possible lazily
→ Rendering a 3D image in 3D: needs *all* the data

Some tips and tricks for OMERO from Phython: https://biapol.github.io/omero-tools

Napari-OMERO usage

Coming soon:

- ROI support
- Multiscale support: Browse larger-than-RAM samples (*3D limitation remains*)



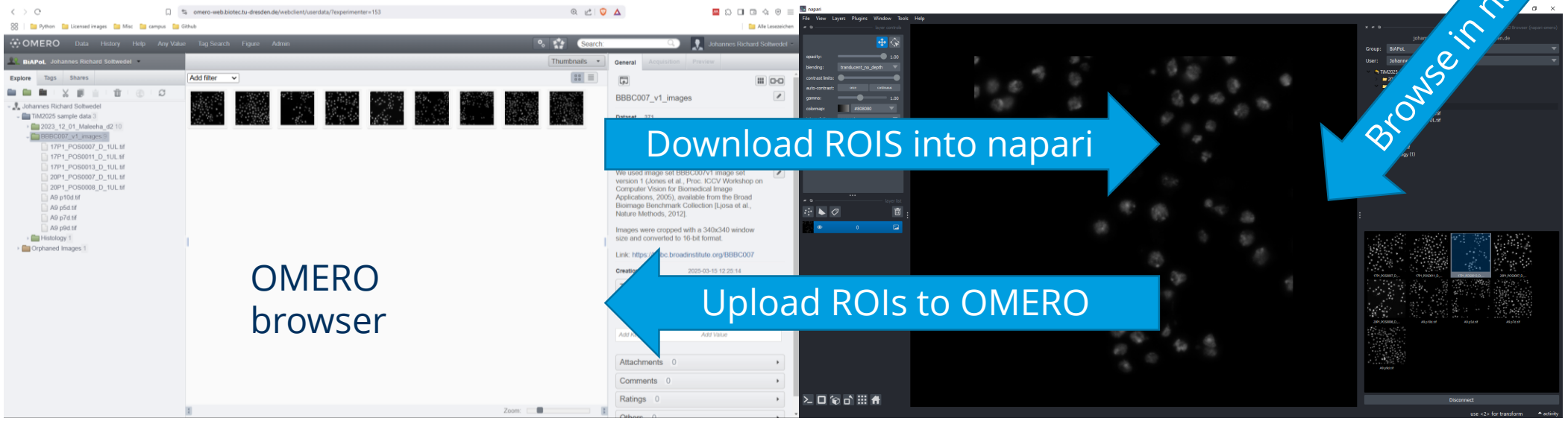
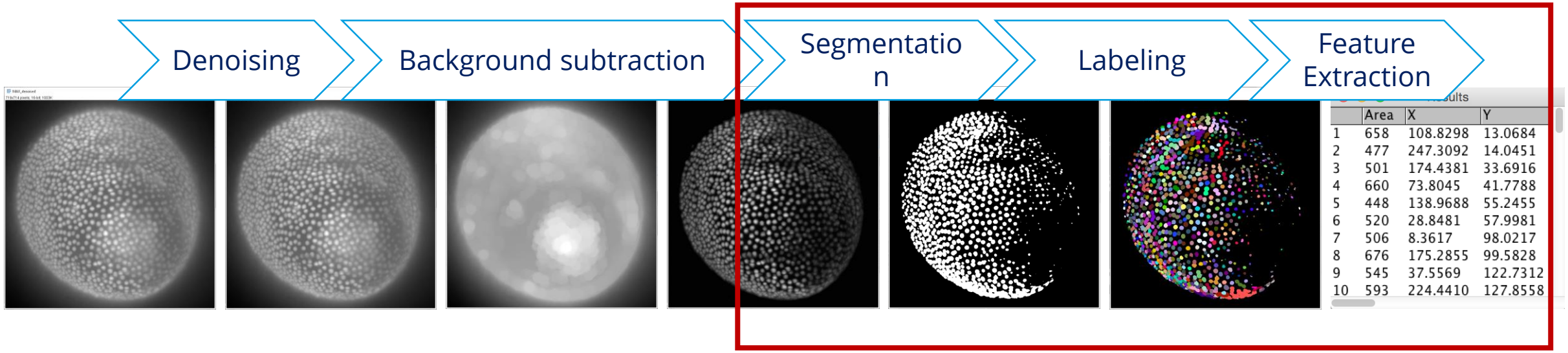


Image Analysis Classic Workflows

Image analysis workflow is a series of processes/functions (not always linear) applied to images to achieve a certain goal (usually some measurements)

Here is a classic example:



Segmentation and Supervised Machine Learning

Random Forest Classifiers

- Pixel Classifier

Application: Segmentation

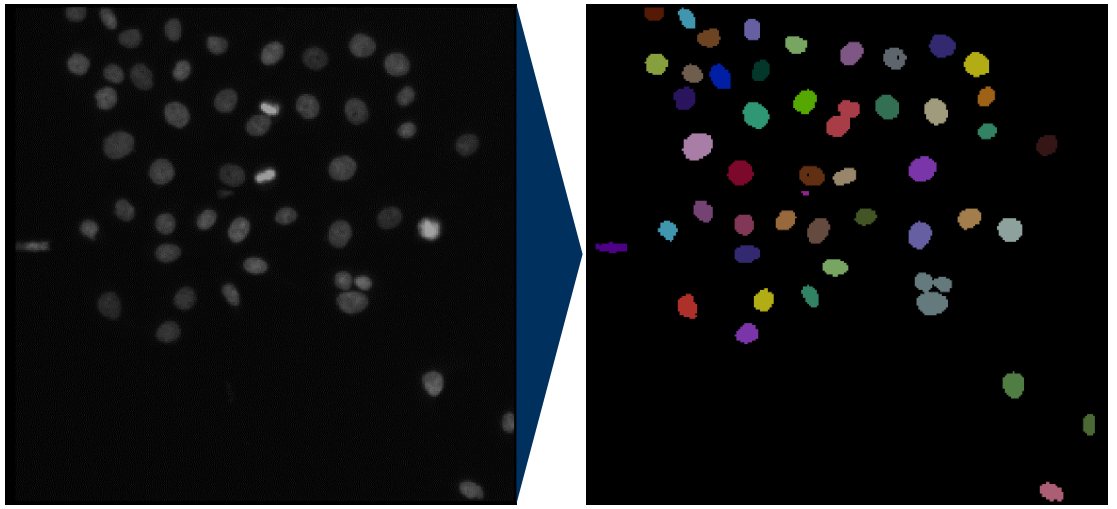
https://scikit-image.org/docs/stable/api/skimage.data.html

Aim:

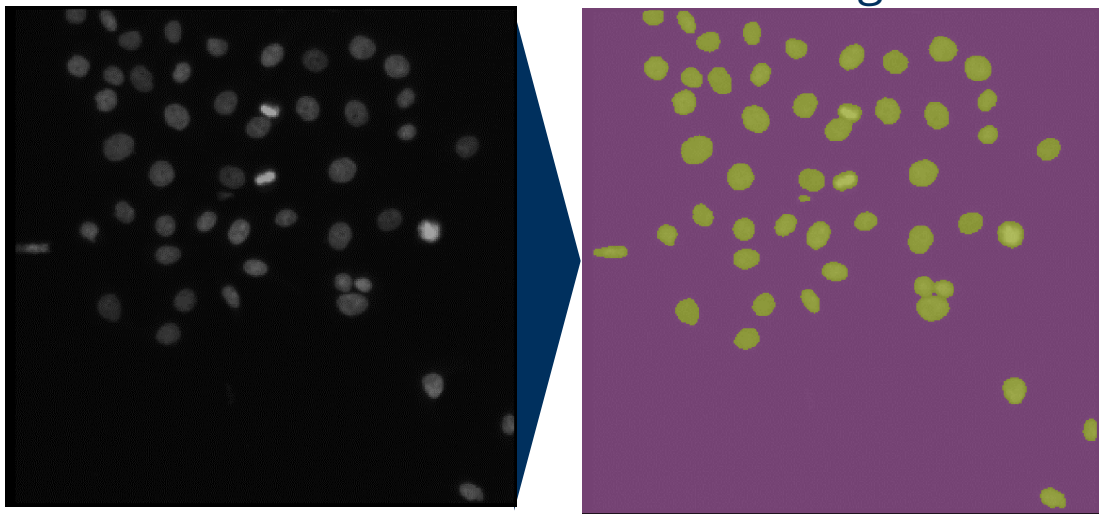
Separate background from foreground

Vocabulary:

- **Segmentation:**
  → Assigning a meaningful *label* to each pixel
  → Segmentation is a *classification* problem
- **Semantic segmentation:**
  Differentiate pixels into multiple *classes* (e.g., membrane, nucleus, cytosol, etc.)
- **Instance segmentation:**
  Differentiate multiple occurrences of the same class into separate instances of this class (e.g., separate *label* for each cell in image)



Instance segmentation



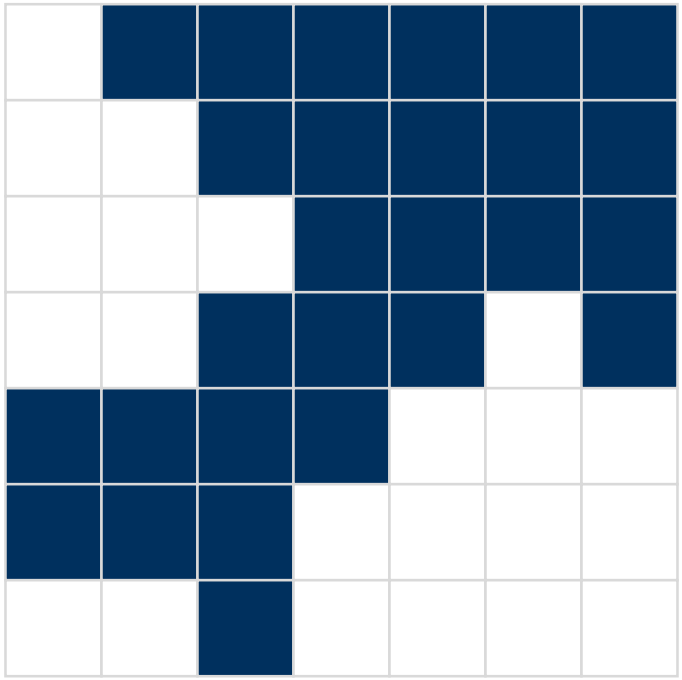
Semantic segmentation

Instance segmentation

In order to allow the computer differentiating objects, connected component analysis (CCA) is used to mark pixels belonging to different objects with different numbers

Background pixels are marked with 0.

The maximum intensity of a labelled map corresponds to the number of objects.



CCA

1	0	0	0	0	0	0
1	1	0	0	0	0	0
1	1	1	0	0	0	0
1	1	0	0	0	3	0
0	0	0	0	3	3	3
0	0	0	3	3	3	3
2	2	0	3	3	3	3

Image segmentation using thresholding

Finding the right workflow towards a good segmentation takes time

A priori, we usually don't know which information in the image is useful for a good segmentation

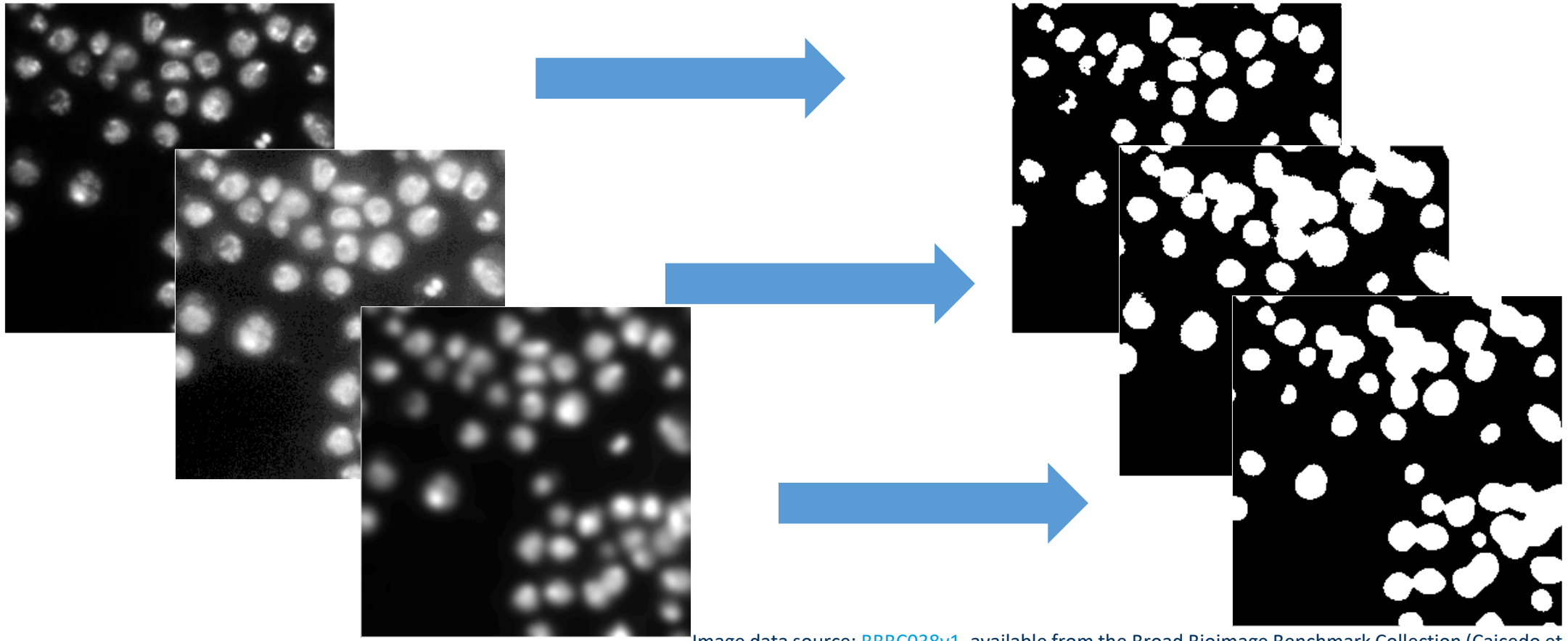
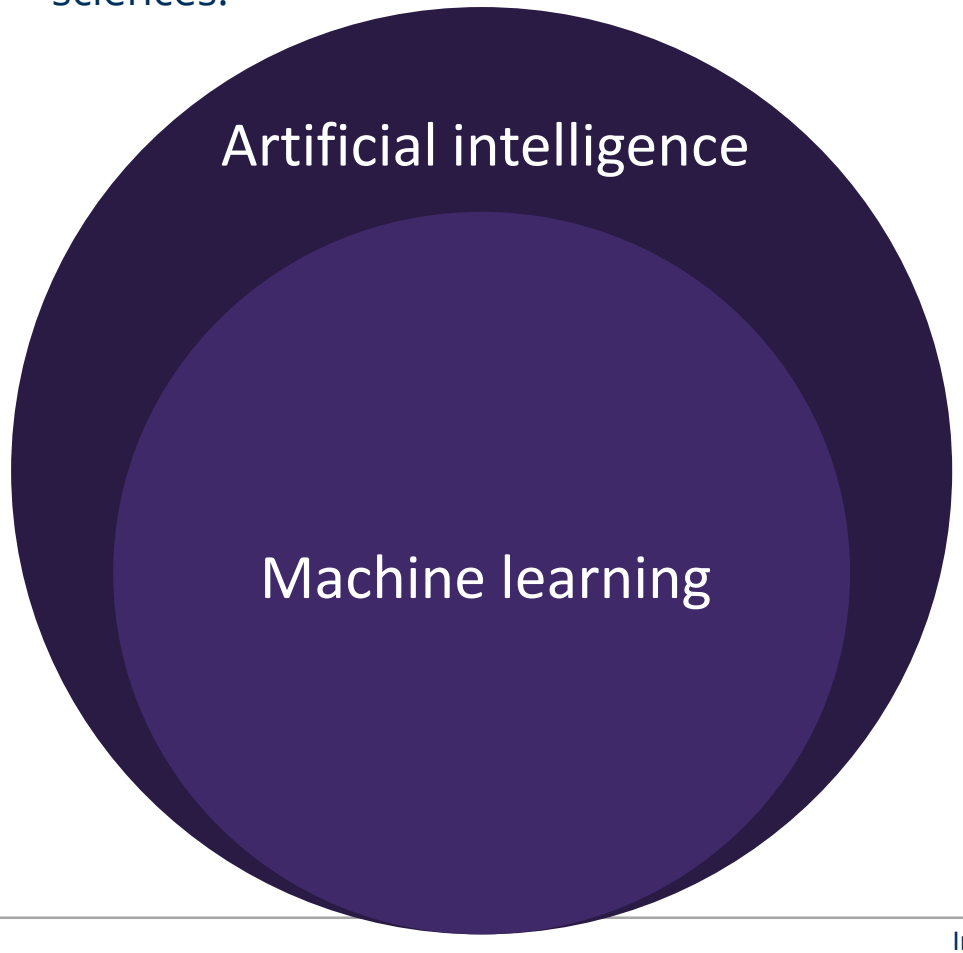
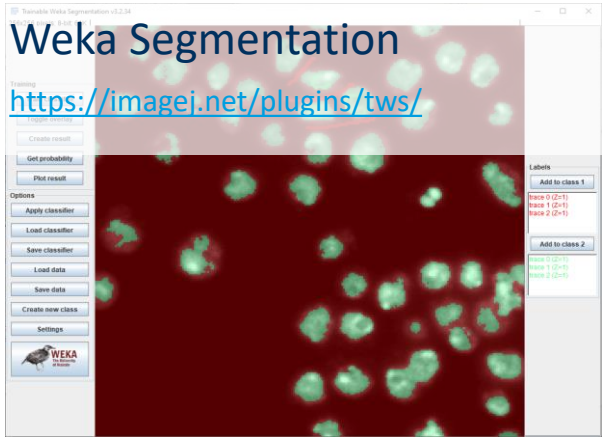


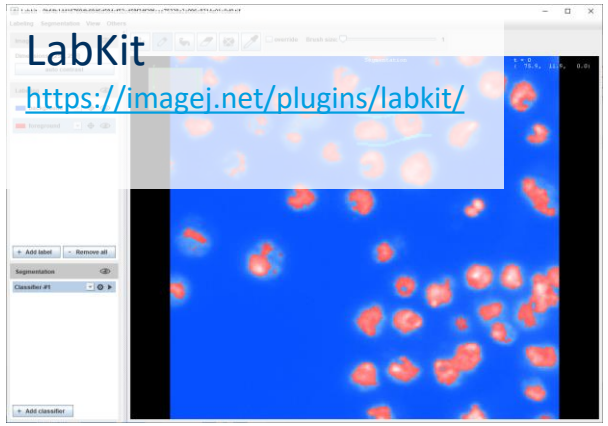
Image data source: BBBC038v1, available from the Broad Bioimage Benchmark Collection (Caicedo et al., Nature Methods, 2019)

Machine learning

- A research field in computer science
- Finds more and more applications, also in life sciences.







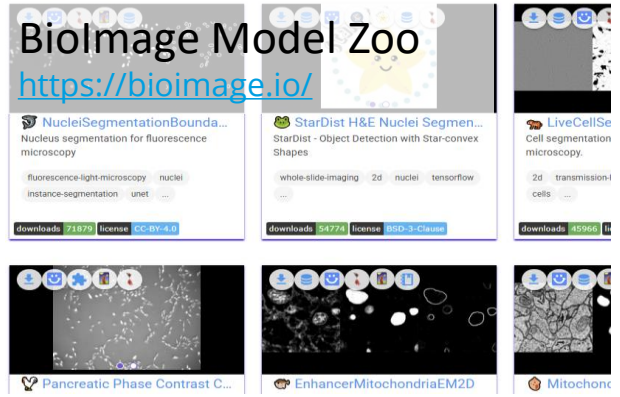
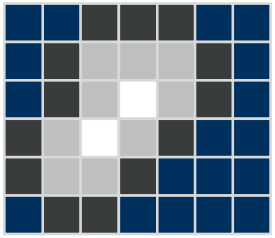


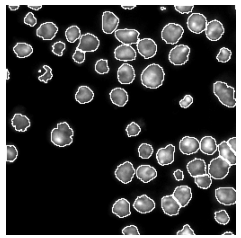
Image data source: BBBC038v1, available from the Broad Bioimage Benchmark Collection (Caicedo et al., Nature Methods, 2019)

Machine learning

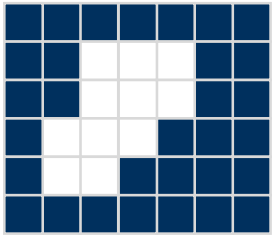
Automatic construction of predictive models from given data

Pixels, Objects, Images, Audio, Text, Measurements, …





Dense Segmentation / Binarization



Object classification

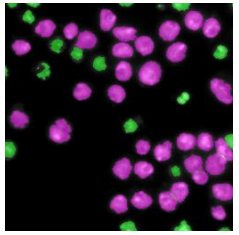
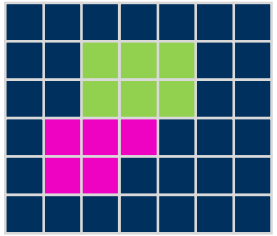


Image classification

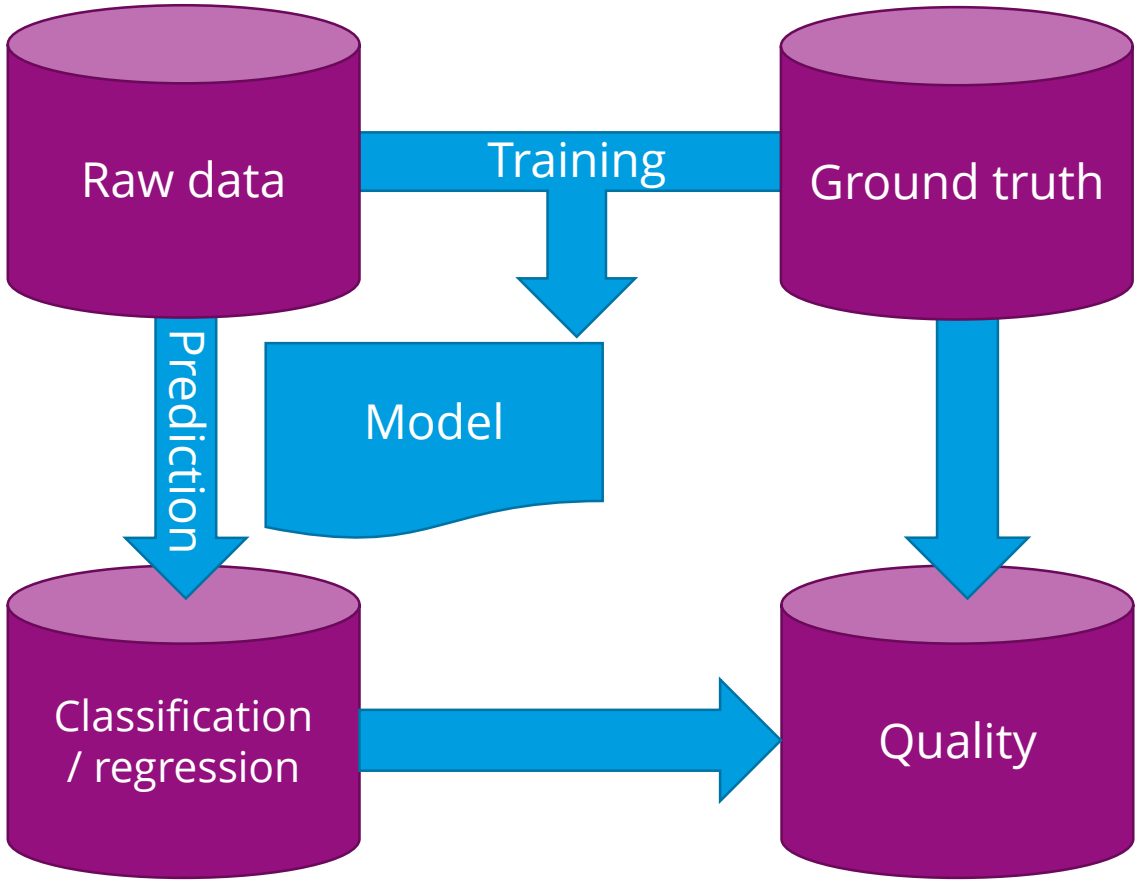
"Cat"

Instance segmentation



Cont. quantity

P_{Cat}= 0.5
$P_{Microscope}$= 0.4
Height = 80 cm



Annotated raw data, usually generated by humans

Precision, Recall

Segmentation: Latest developments

1970s-2010: Filtering, thresholding

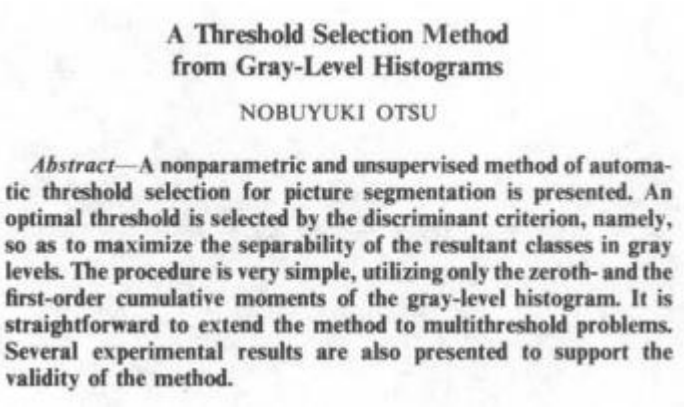
2010s: Random forests et al.

Deep learning

2015: UNet
2018: Stardist

Foundational models

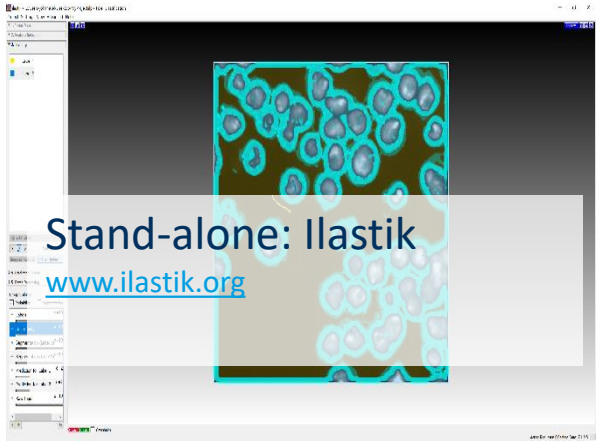
2023: Segment anything (SAM)
2024: SAM 2



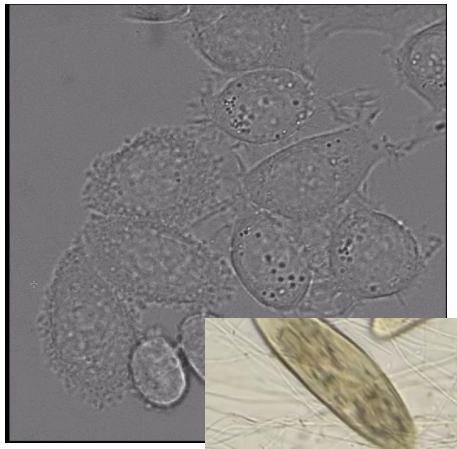
A Threshold Selection Method from Gray-Level Histograms

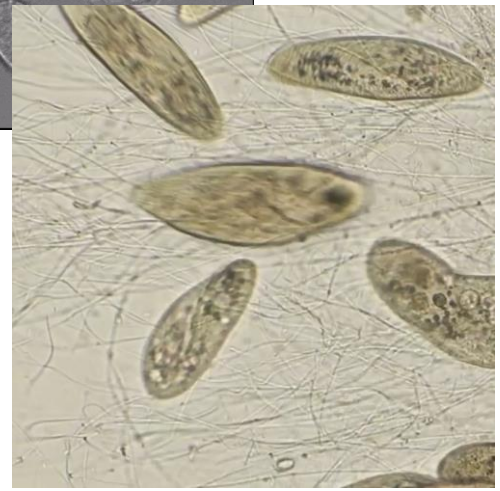
NOBUYUKI OTSU

Abstract—A nonparametric and unsupervised method of automatic threshold selection for picture segmentation is presented. An optimal threshold is selected by the discriminant criterion, namely, so as to maximize the separability of the resultant classes in gray levels. The procedure is very simple, utilizing only the zeroth- and the first-order cumulative moments of the gray-level histogram. It is straightforward to extend the method to multithreshold problems. Several experimental results are also presented to support the validity of the method.



Stand-alone: Ilastik
www.ilastik.org



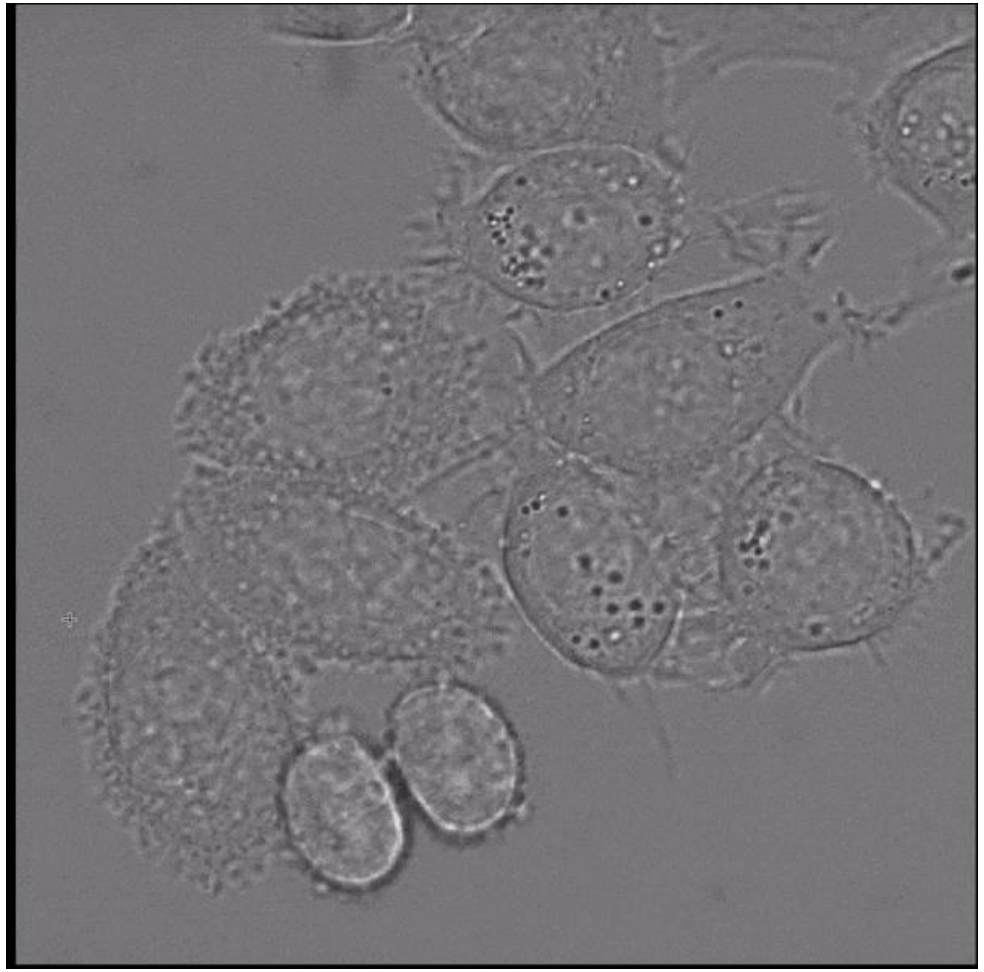


Computational demand

Segmentation and Supervised Machine Learning in napari

- Demonstration: Micro-sam plugin

Micro-sam



https://github.com/computational-cell-analytics/micro-sam

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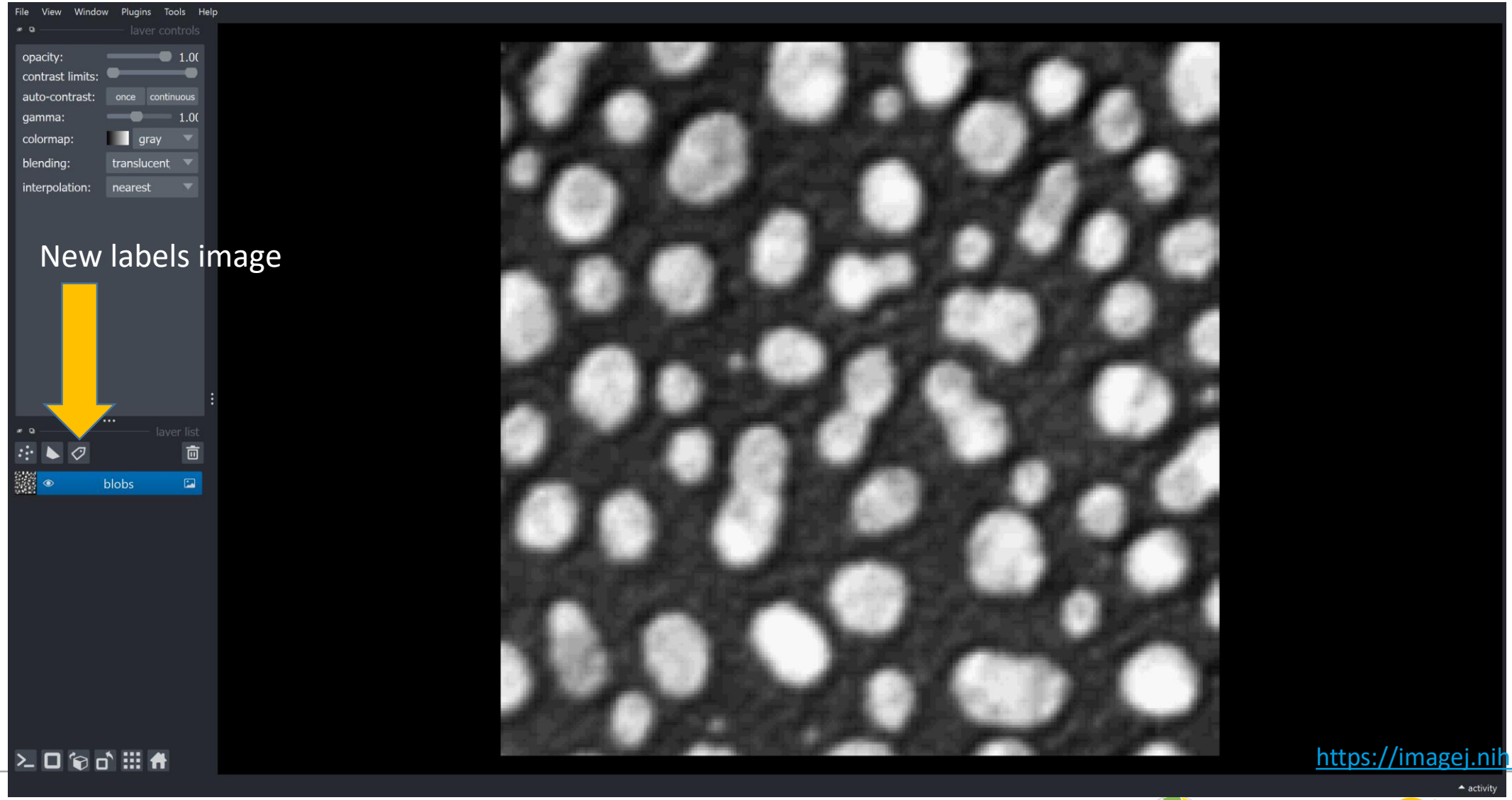
Segmentation and Supervised Machine Learning in napari

Random Forest Classifiers

- Pixel Classifier
- **napari-apoc plugin**

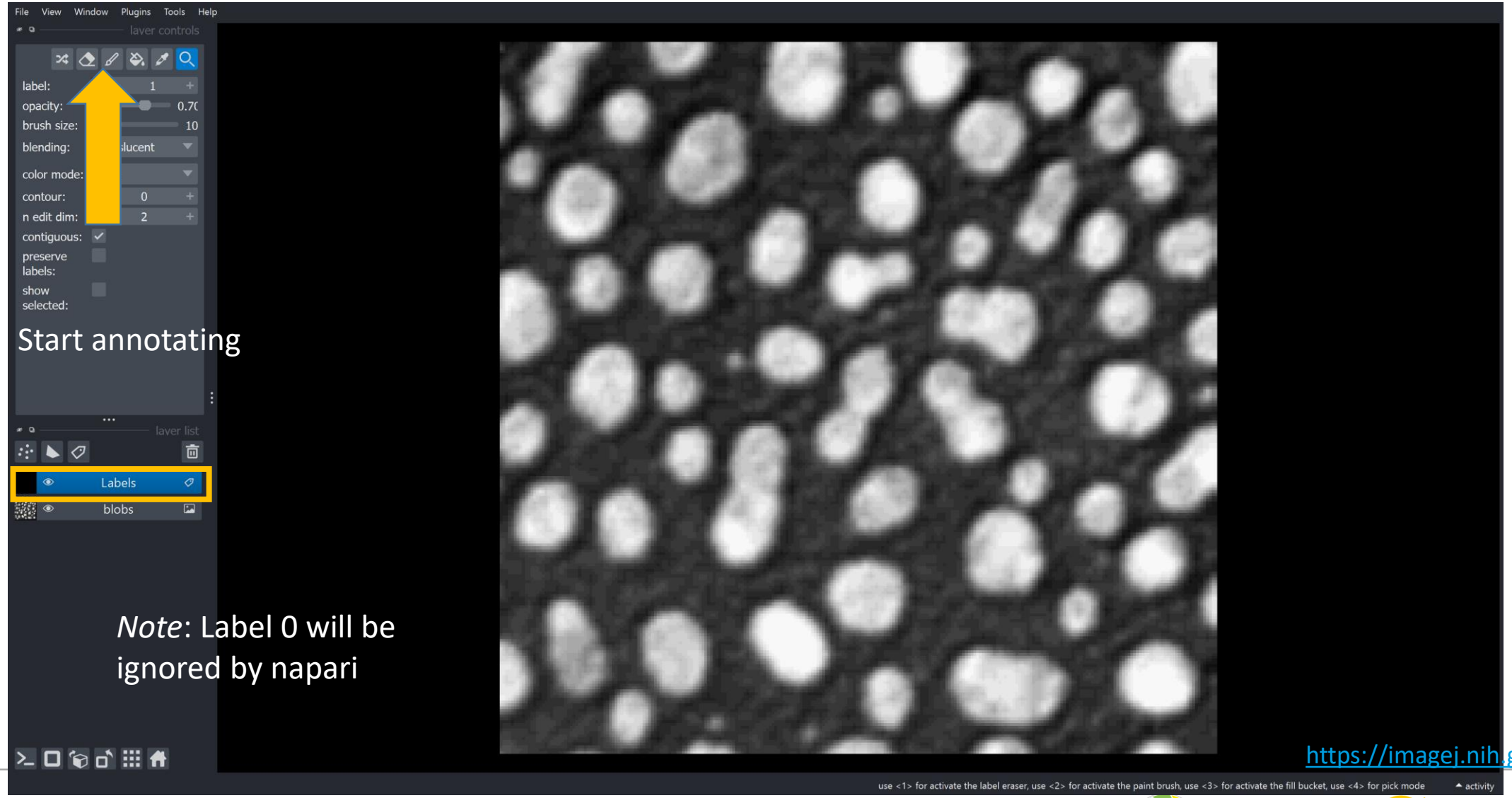
DRESDEN concept

In napari: annotation



https://imagej.nih.gov/ij/images/

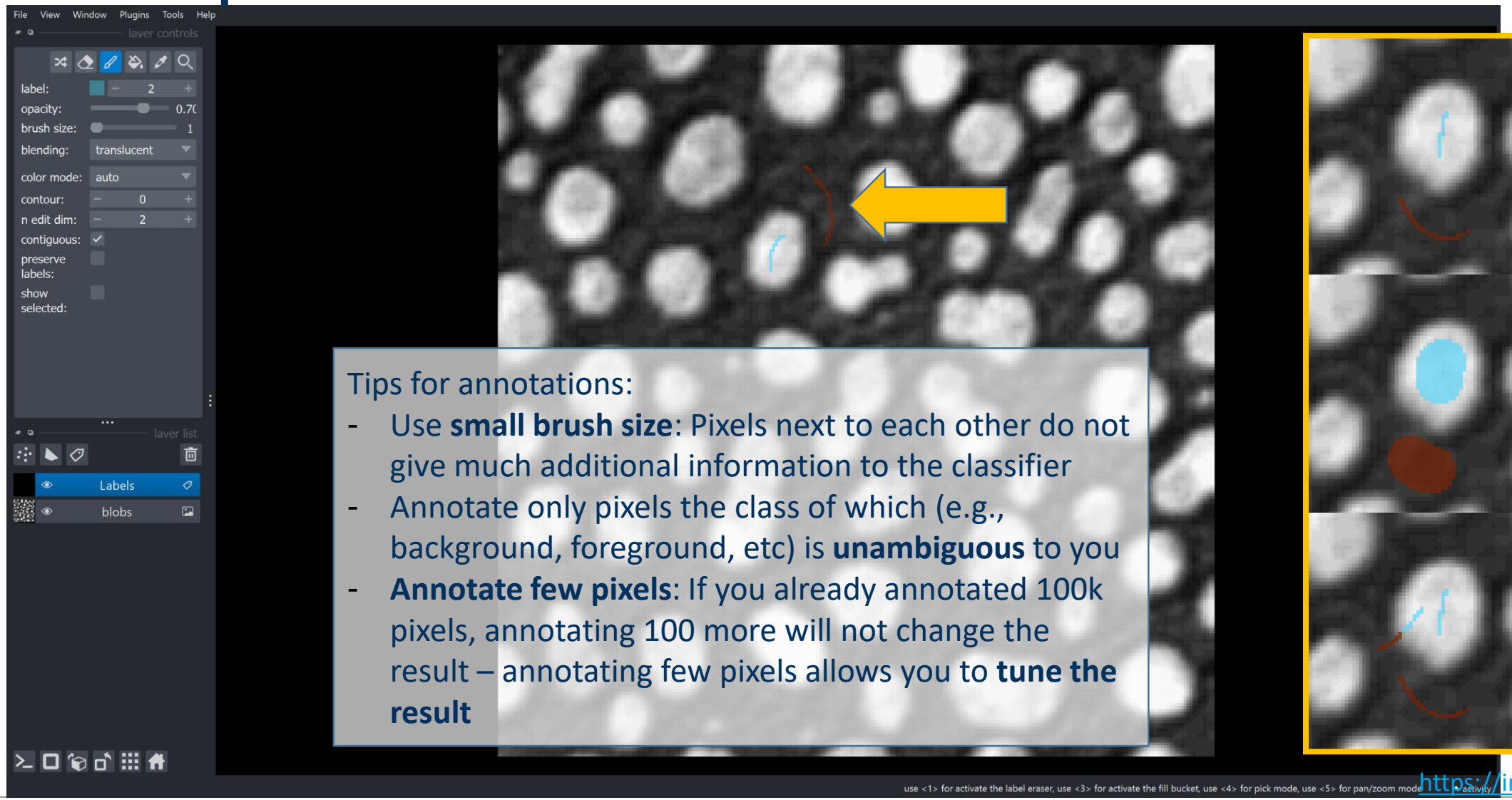
In napari: annotation



Note: Label 0 will be ignored by napari

https://imagej.nih.gov/ij/images/

In napari: annotation



Tips for annotations:

- Use **small brush size**: Pixels next to each other do not give much additional information to the classifier
- Annotate only pixels the class of which (e.g., background, foreground, etc) is **unambiguous** to you
- **Annotate few pixels**: If you already annotated 100k pixels, annotating 100 more will not change the result – annotating few pixels allows you to **tune the result**

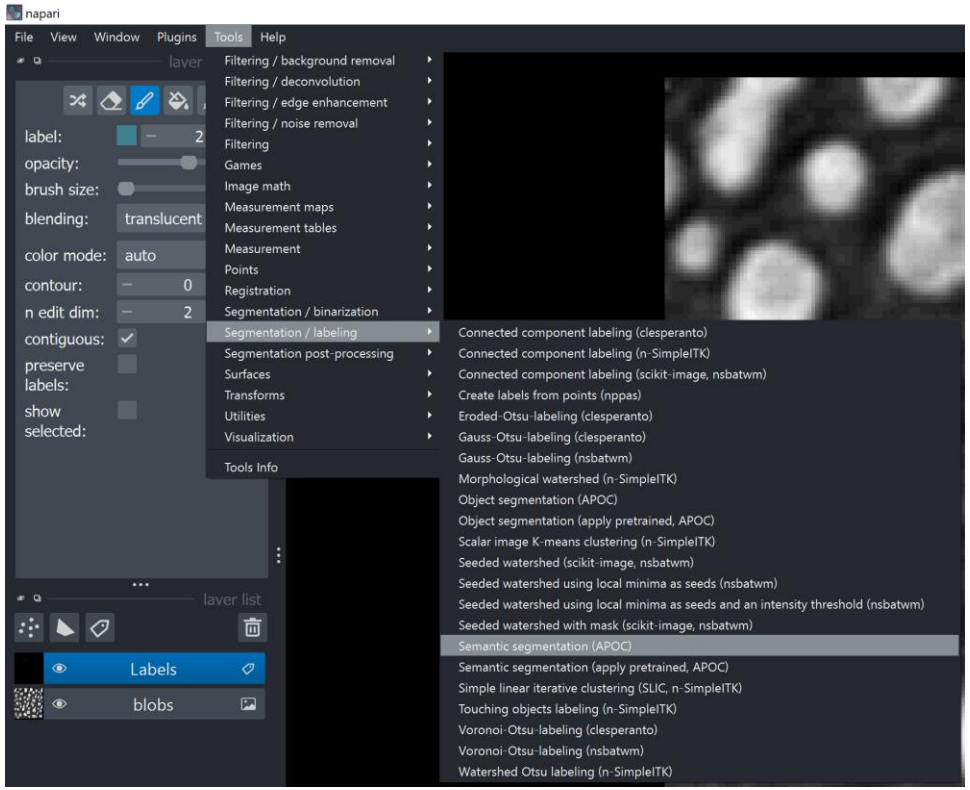
https://imagej.nih.gov/ij/images/

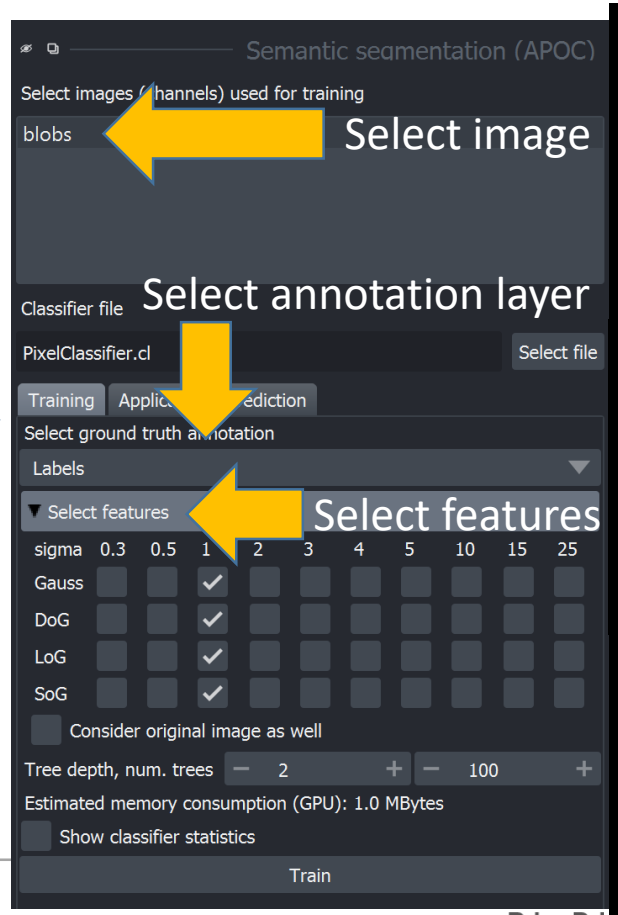
In napari: Semantic segmentation (training)

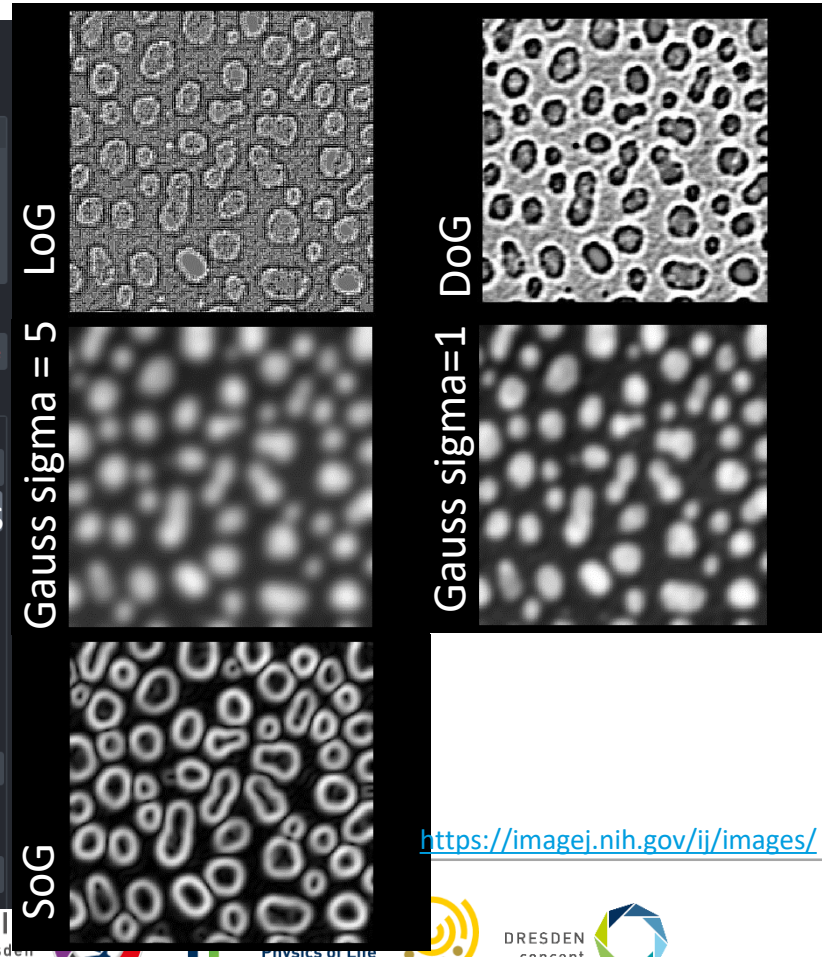
Two options:

Semantic segmentation: Predict class of every pixel according to annotation

Object segmentation: Assumes that class “1” refers to background – applies connected component analysis to other class

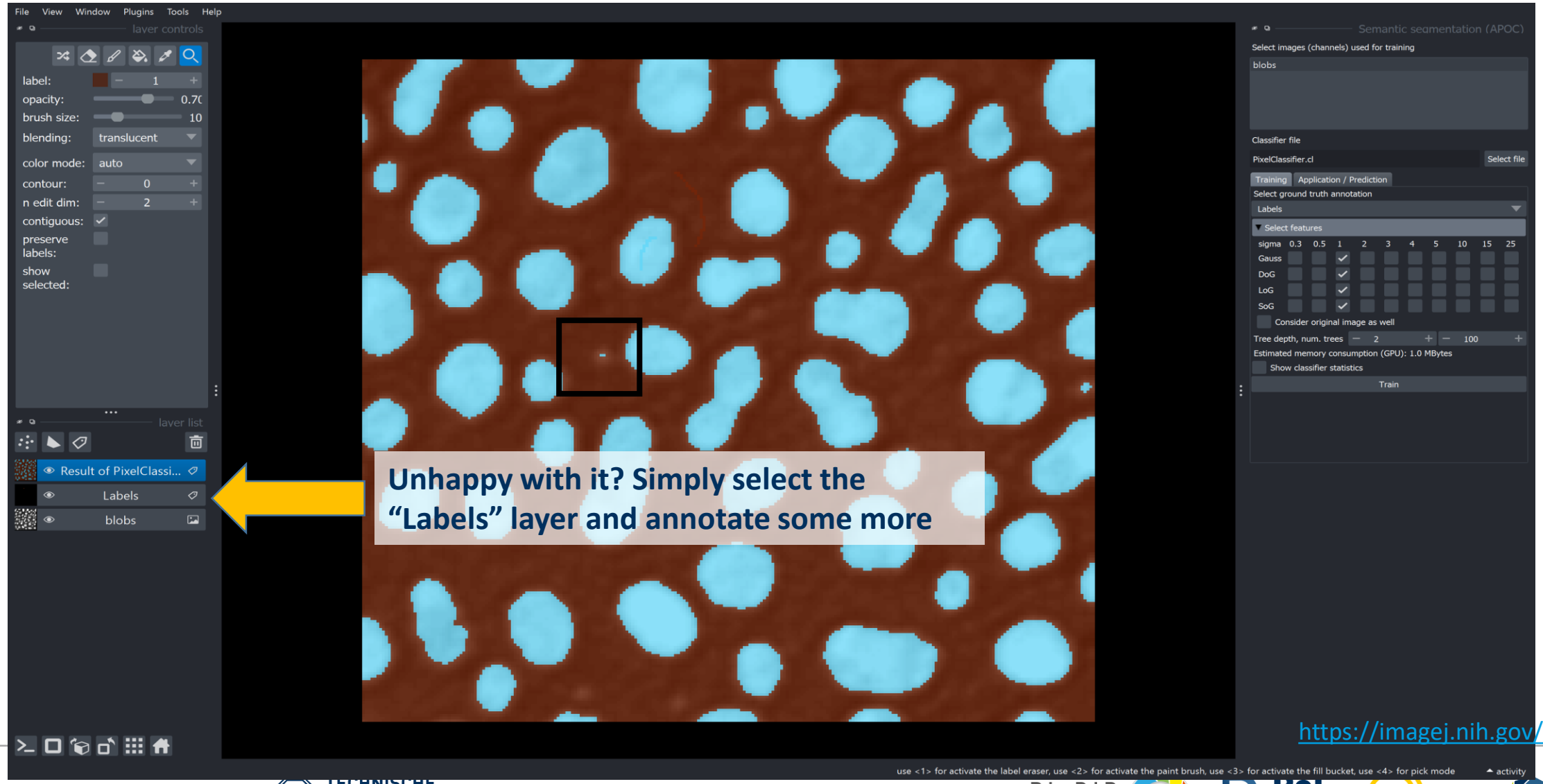






https://imagej.nih.gov/ij/images/

In napari: Semantic segmentation (training)

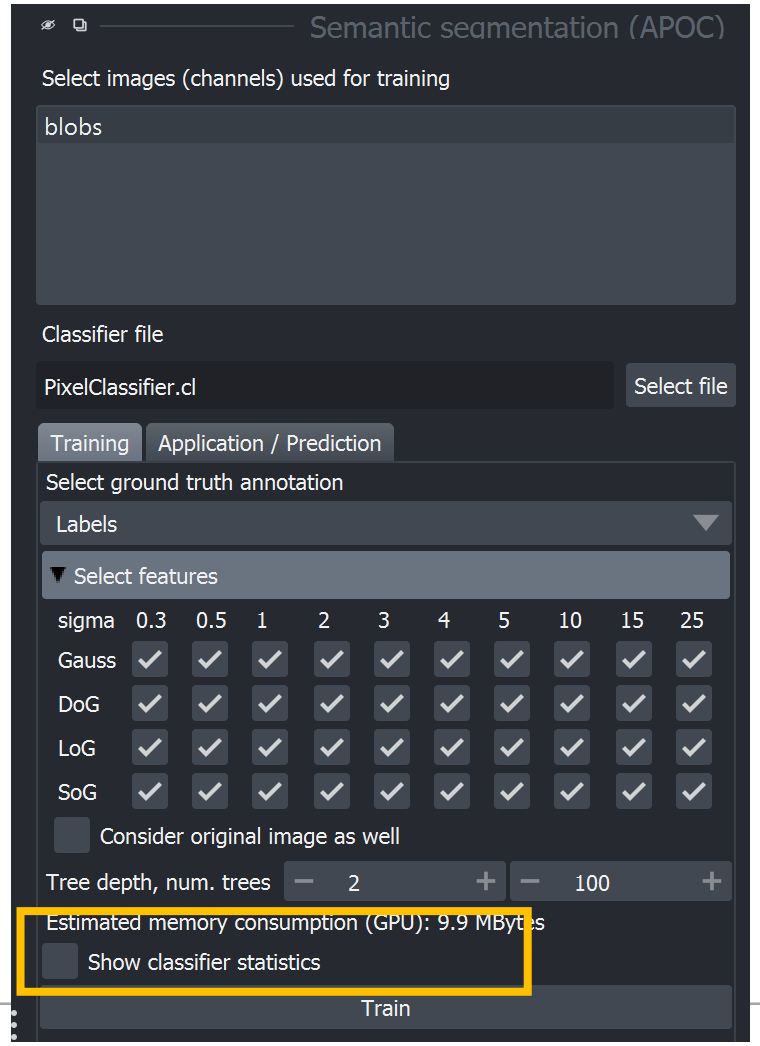


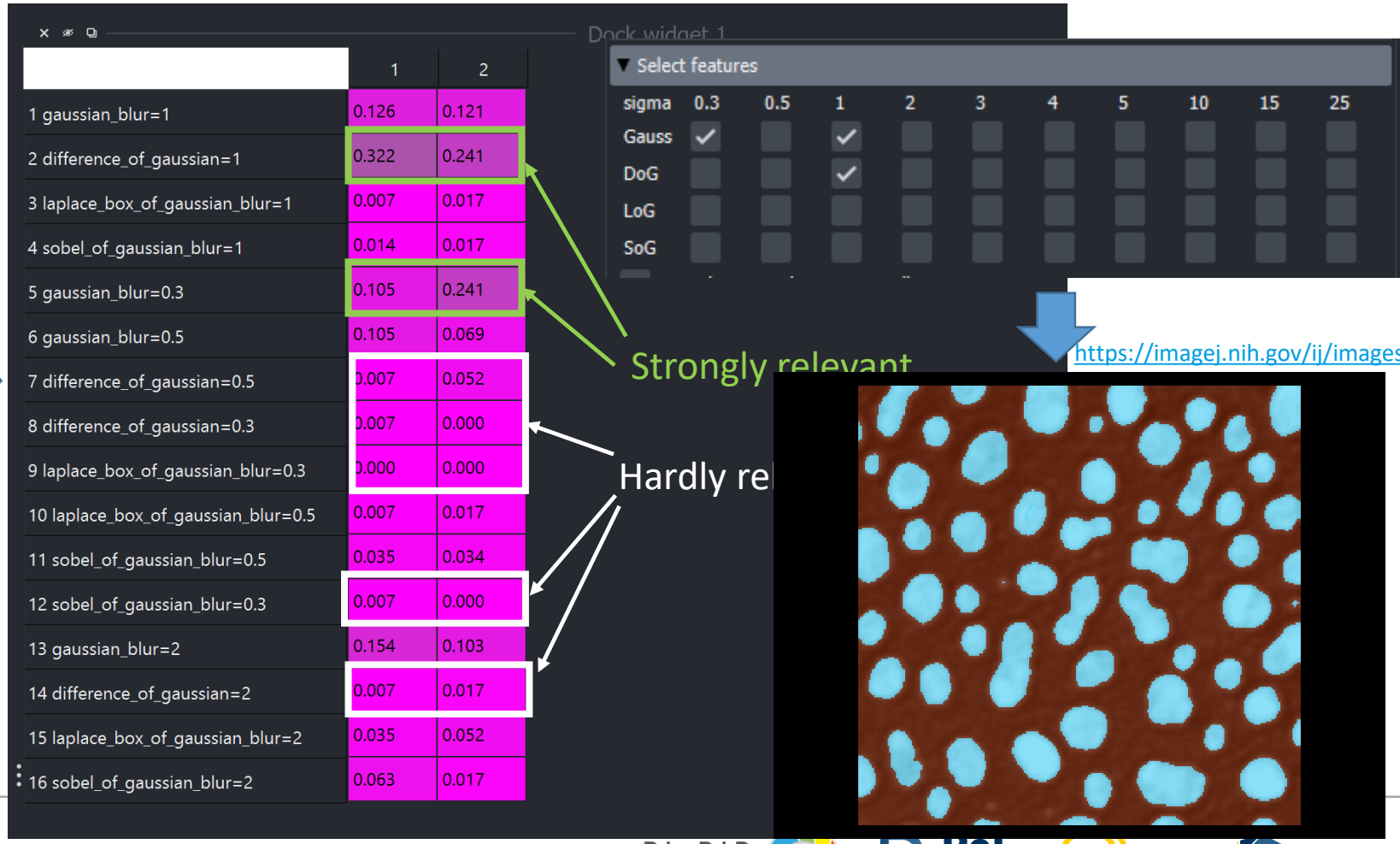
https://imagej.nih.gov/ij/images/

Semantic segmentation: Choosing the right features

Why not just do this?

- Not all features are equally relevant!
- Calculating the features takes time and computation resources!





Feature Extraction

Feature extraction

- A feature is a countable or measurable property of an image or object.
- Goal of feature extraction is finding a minimal set of features to describe an object well enough to differentiate it from other objects.

- **Intensity based**
  - Mean intensity
  - Standard deviation
  - Total intensity
  - Textures
- **Shape based /spatial**
  - Area / Volume
  - Roundness
  - Solidity
  - Circularity / Sphericity
  - Elongation
  - Centroid
  - Bounding box
- **Spatio-temporal**
  - Displacement,
  - Speed,
  - Acceleration
- **Others**
  - Overlap
  - Colocalization
  - Neighborhood
- **Mixed features**
  - Center of mass
  - Local minima / maxima

Further reading:
https://focalplane.biologists.com/2023/05/03/feature-extraction-in-napari/

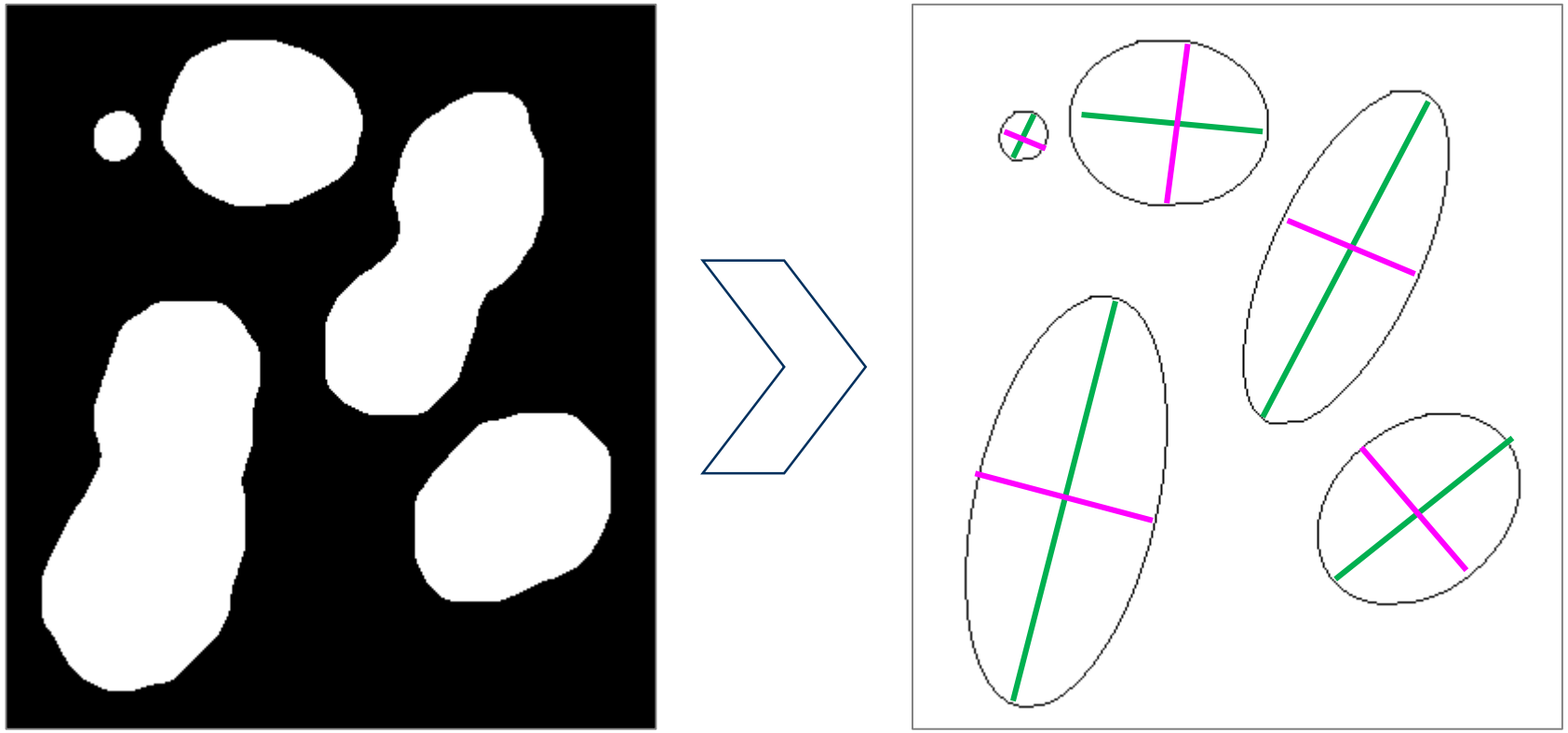
Fit ellipse

For every object, find the optimal ellipse simplifying the object.

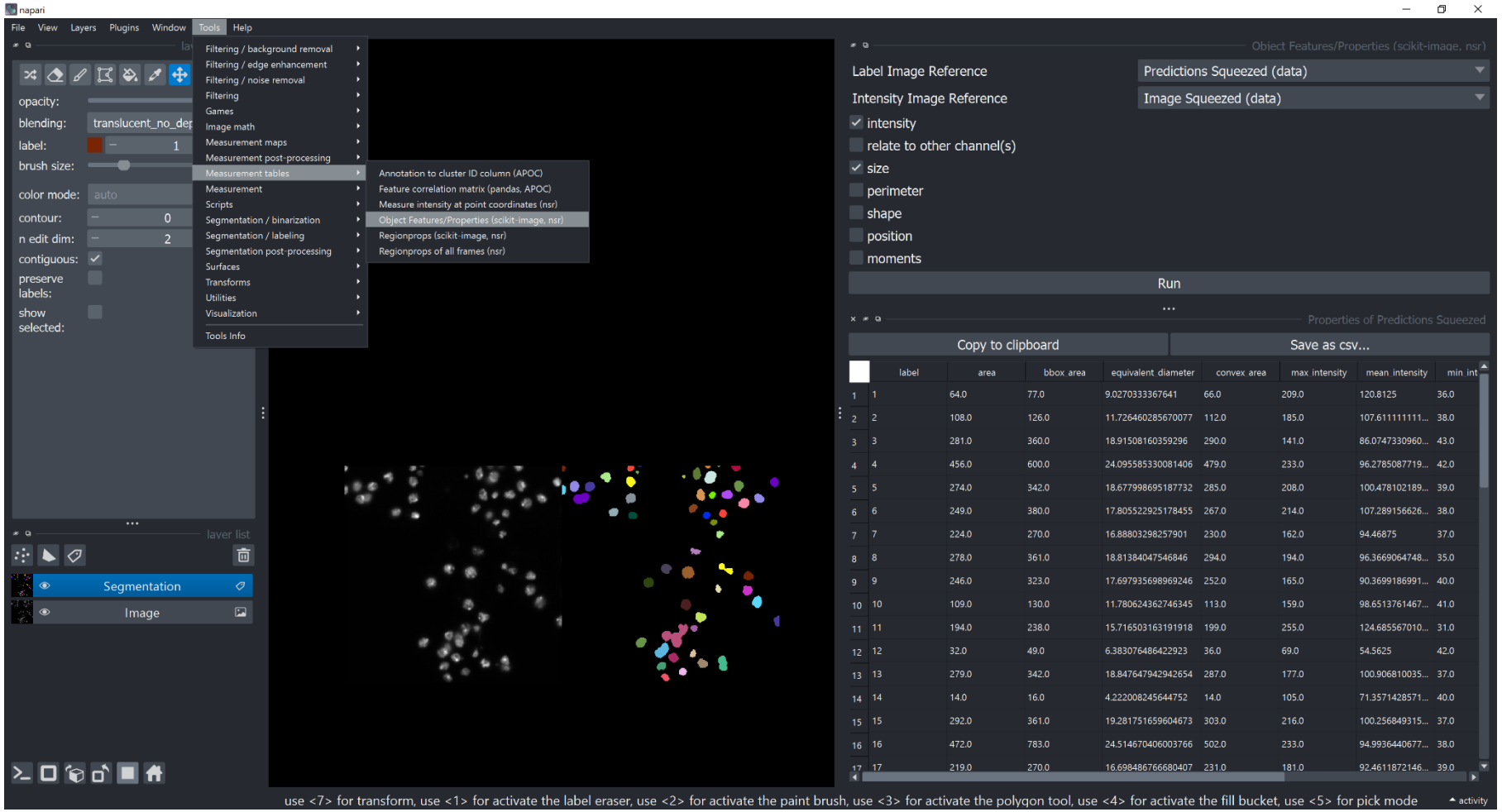
Major axis ... long diameter

Minor axis ... short diameter

Major and minor axis are perpendicular to each other

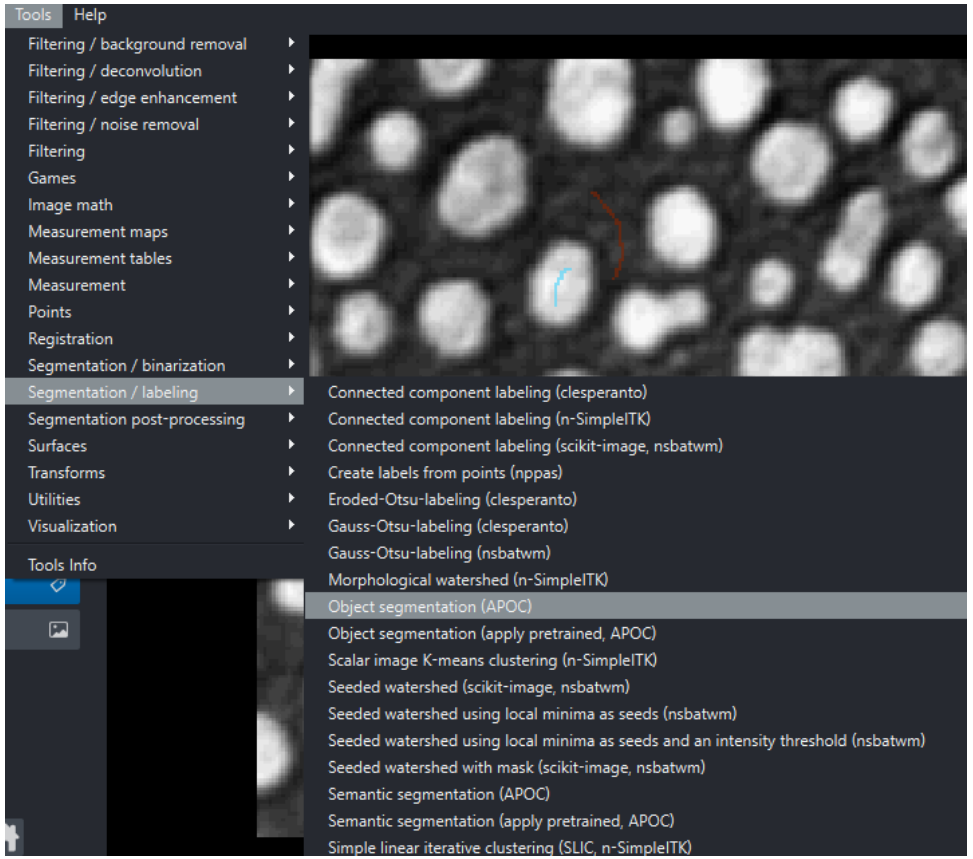


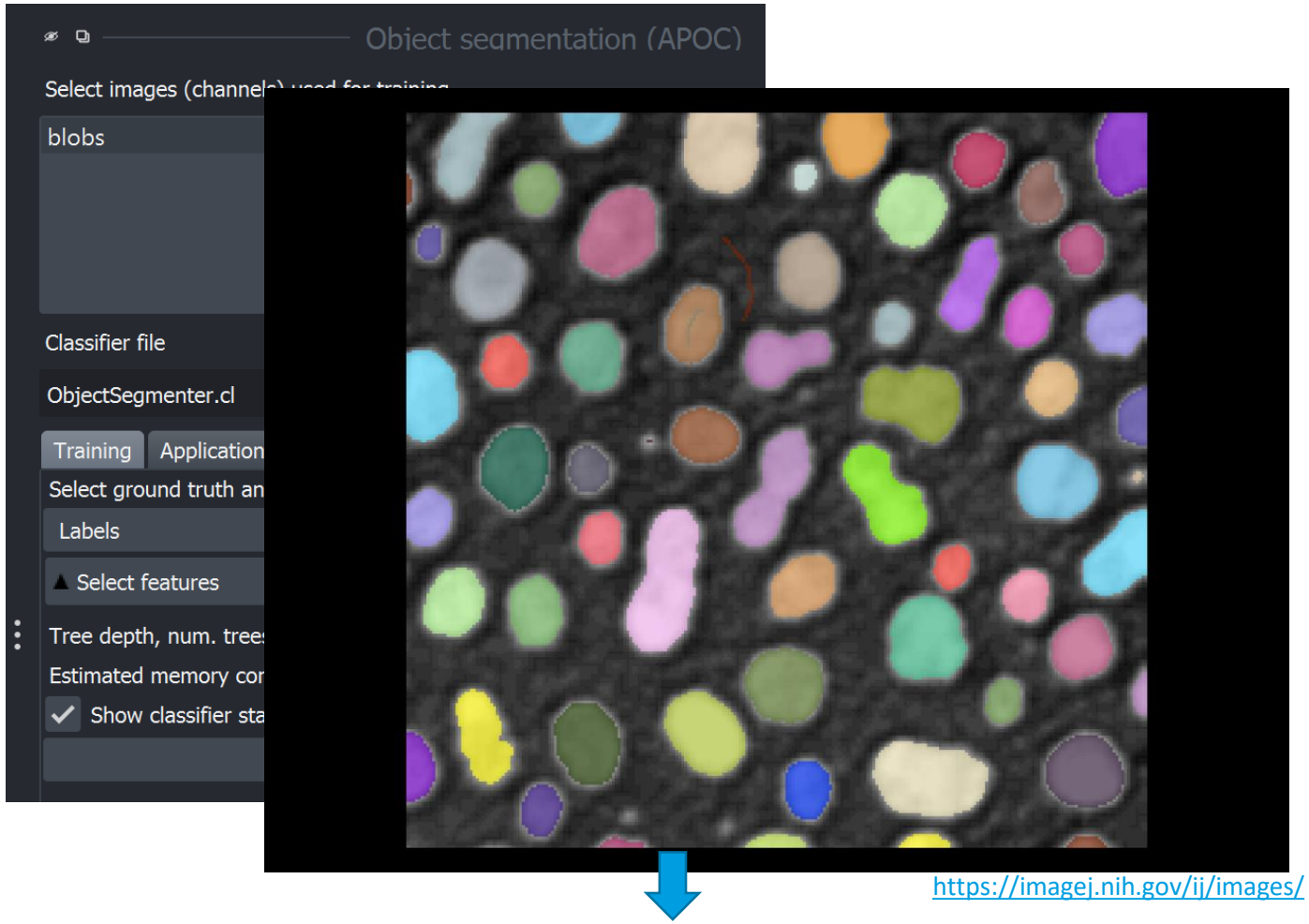
napari-skimage-regionprops



https://github.com/haesleinhuepf/napari-skimage-regionprops

Exercise: Object segmentation





1. Activate the environment and open napari
   ```
   mamba activate napari25
   napari
   ```
2. Download and open the Blobs sample
3. Perform object segmentation on blobs sample dataset

https://imagej.nih.gov/ij/images/

https://biapol.github.io/BioImage-Analysis-and-Data-Processing-Workshop-2025/interactive_pixel_classification/readme.html


Break

Multichannel Analysis

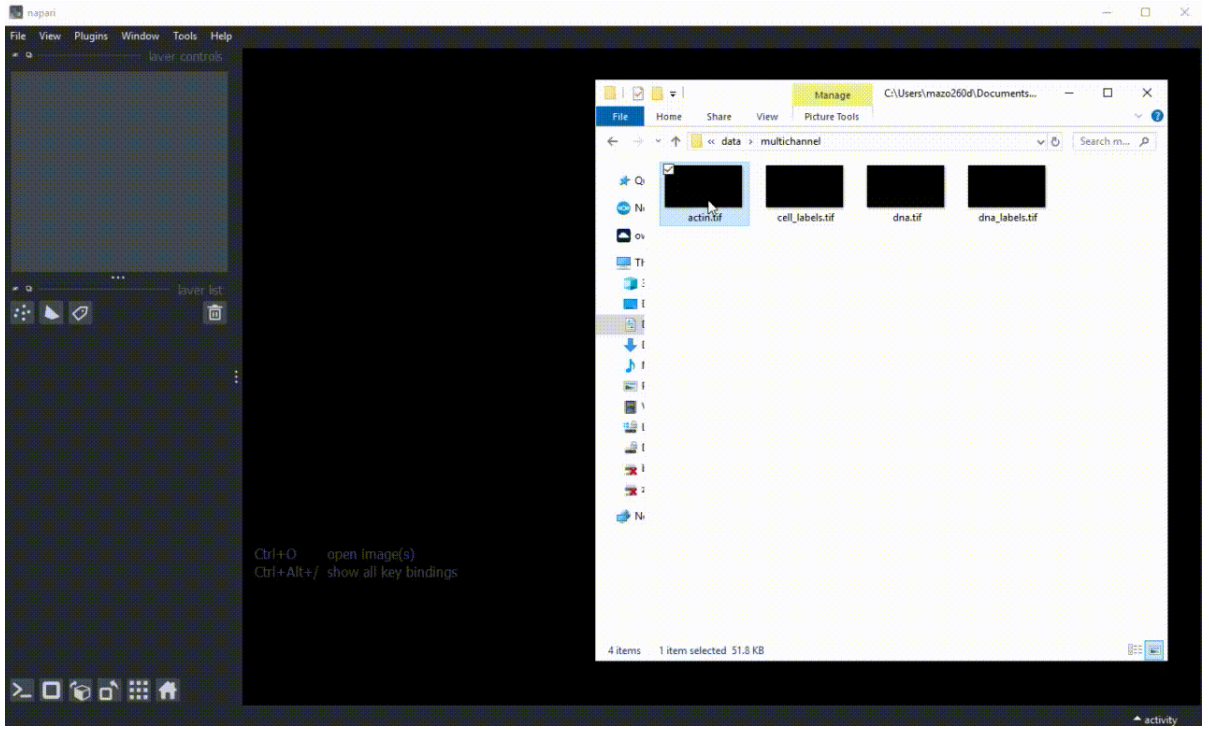
- napari-skimage-regionprops plugin

Exercise: Multichannel Analysis with napari-skimage-regionprops

1. Start up a terminal
2. Activate the environment using:
   `mamba activate napari25`
3. Run:
   `napari`
4. Open the following images in napari

   (located in the data/multichannel folder from the GitHub repository):

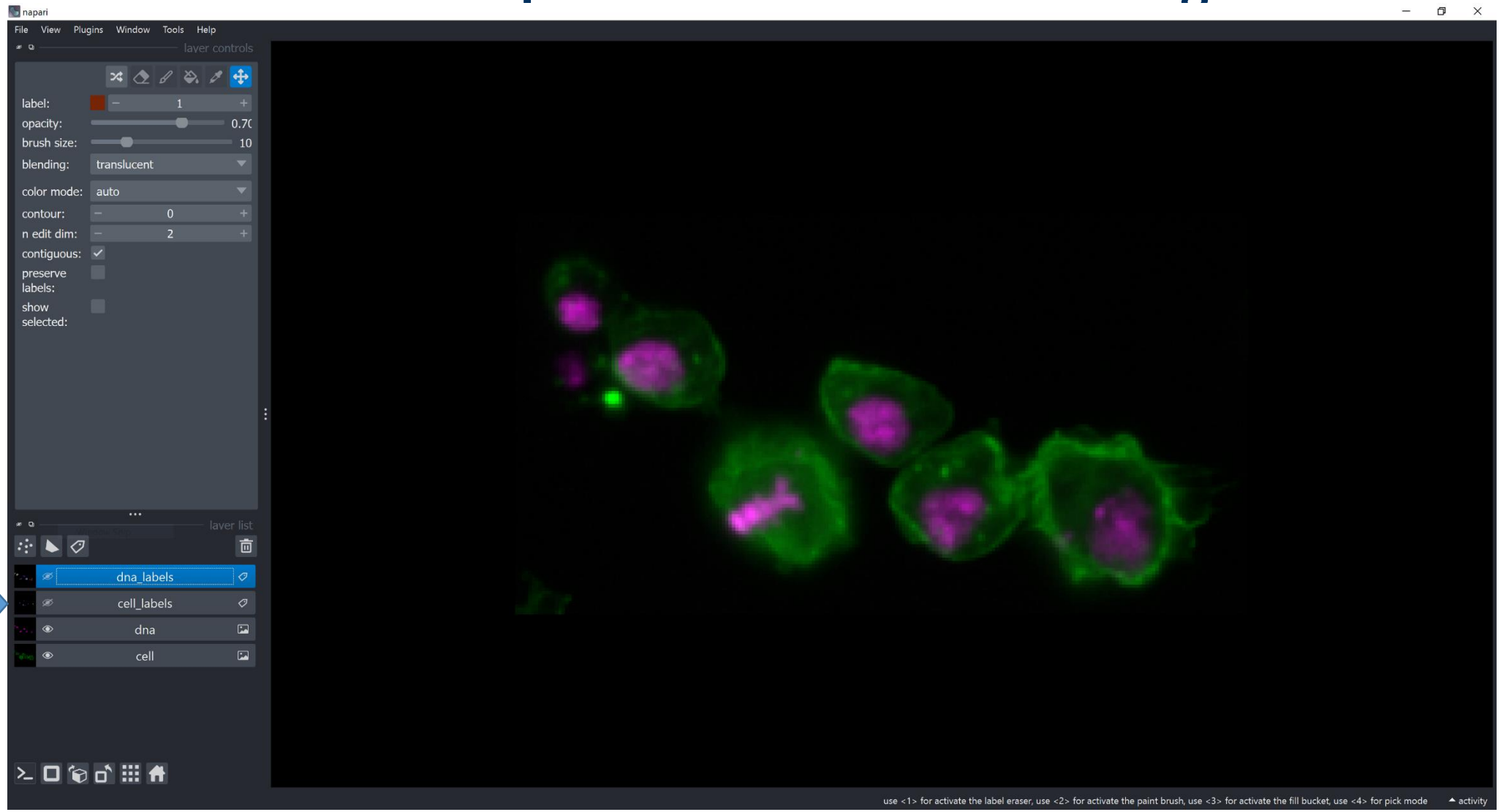
   — actin.tif
   — dna.tif
   — cell_labels.tif
   — dna_labels.tif



5. Change blending mode of ‘actin’ and ‘dna’ Image layers to ‘additive’ (it is a dropdown in the Image layer controls)
6. Select the ‘cell_labels’ Labels layer, and change the contour parameter from 0 to 1 (it is a spinbox in the Labels layer controls)
7. Change the colormaps of these layers to green and magenta, respectively

https://biapol.github.io/BioImage-Analysis-and-Data-Processing-Workshop-2025/multichannel_analysis/readme.html

Feature extraction in napari with multichannel images



https://github.com/haesleinhuepf/napari-skimage-regionprops

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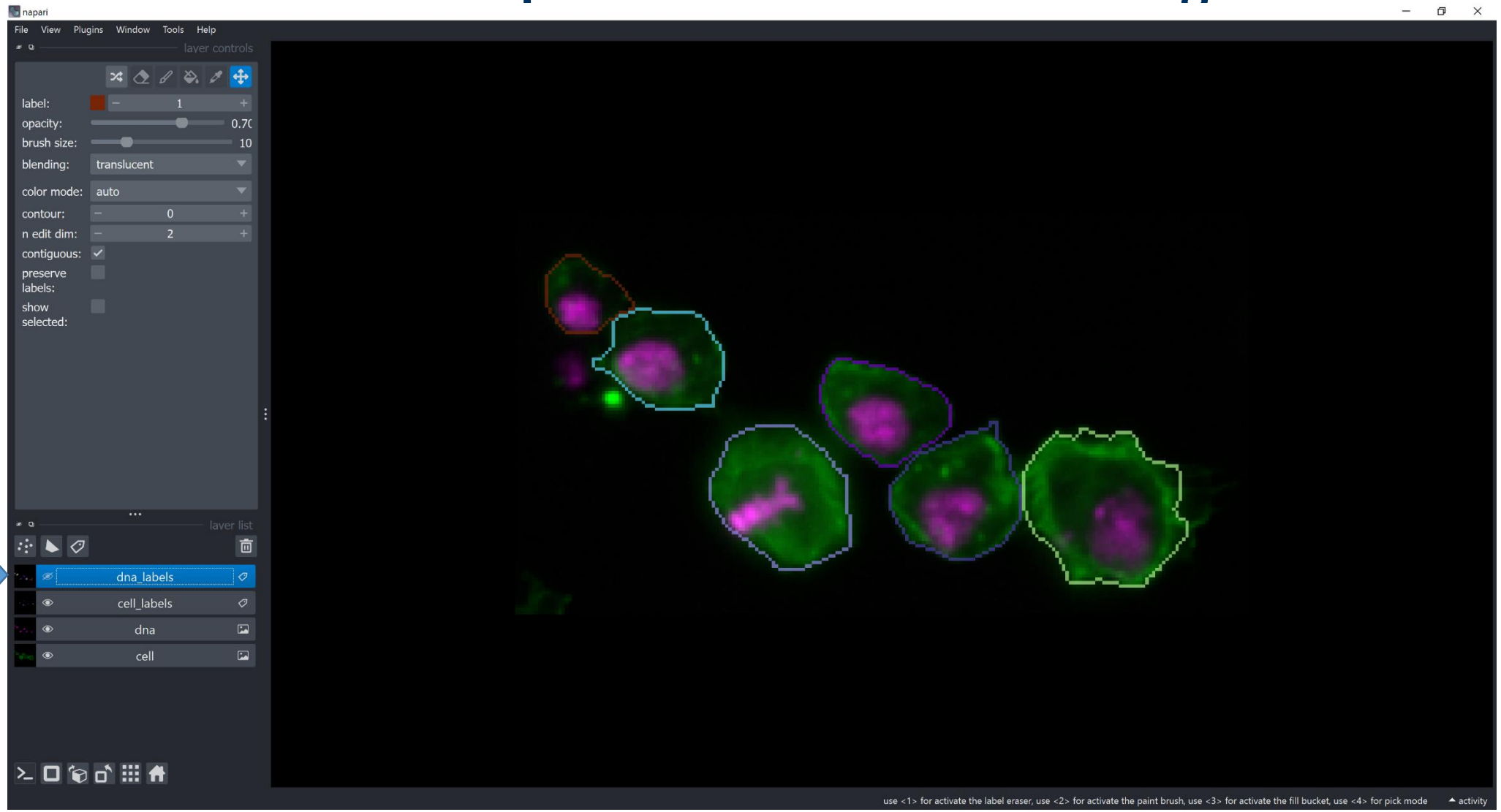
DRESDEN concept

Feature extraction in napari with multichannel images



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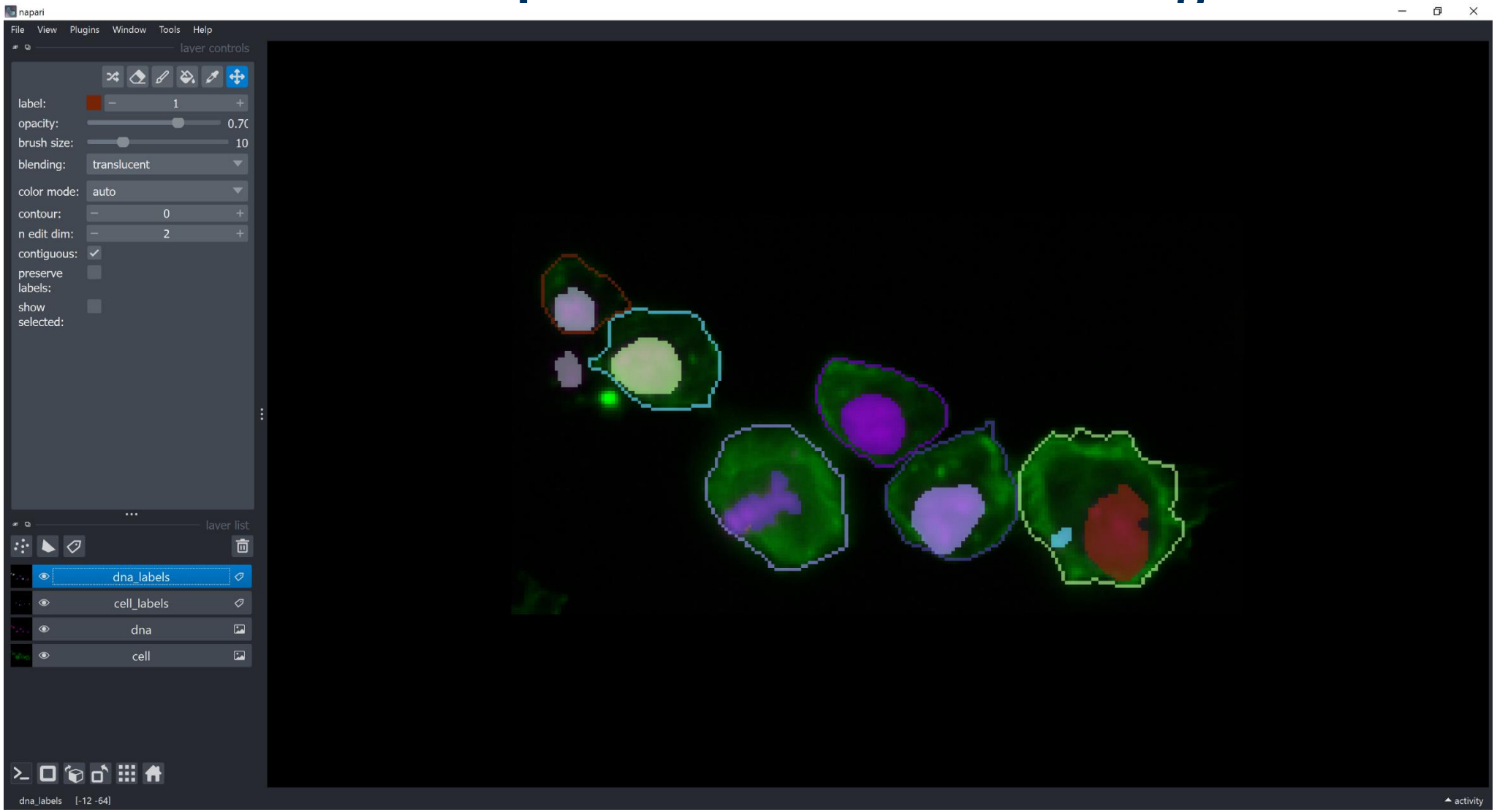

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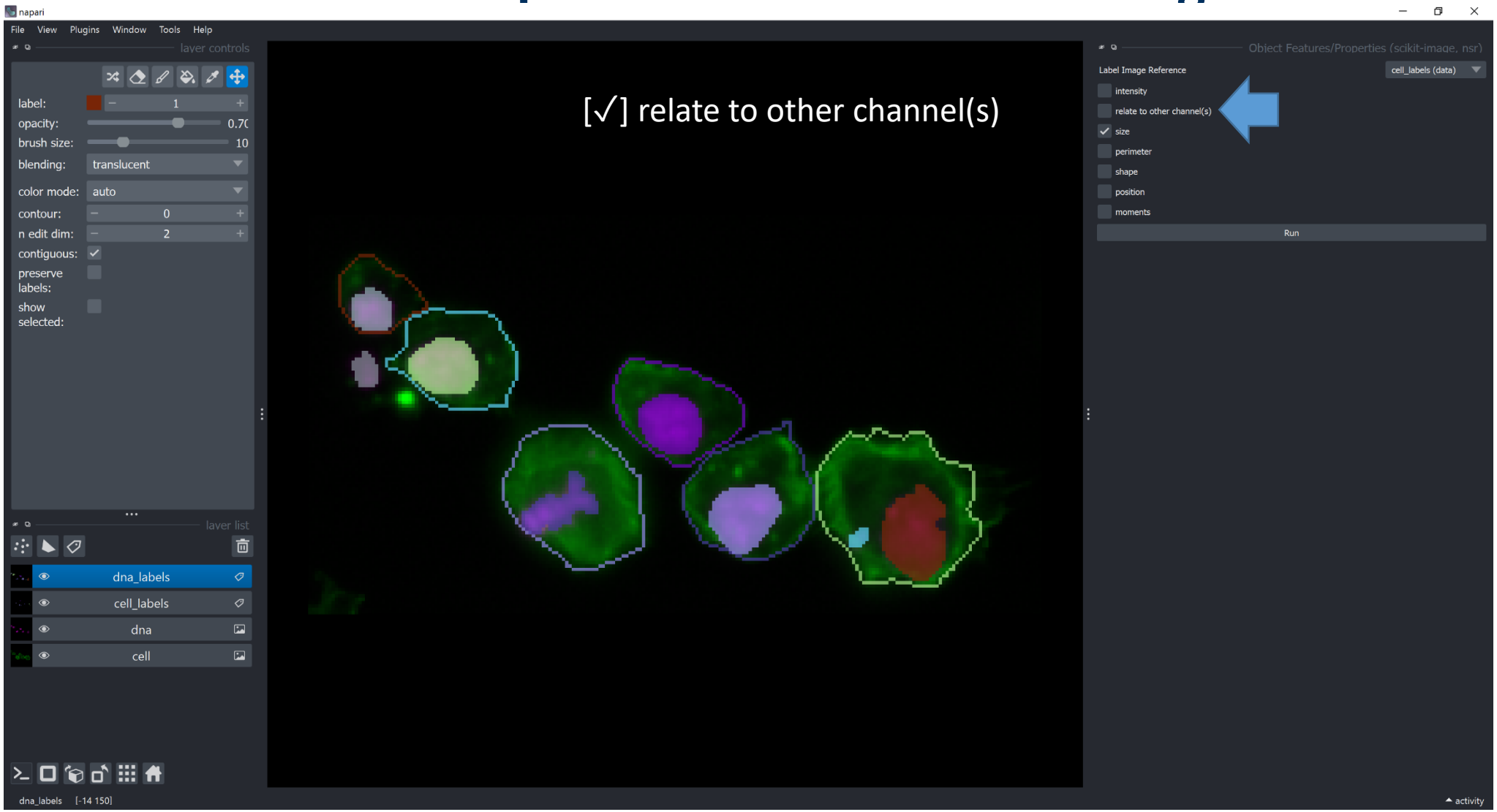

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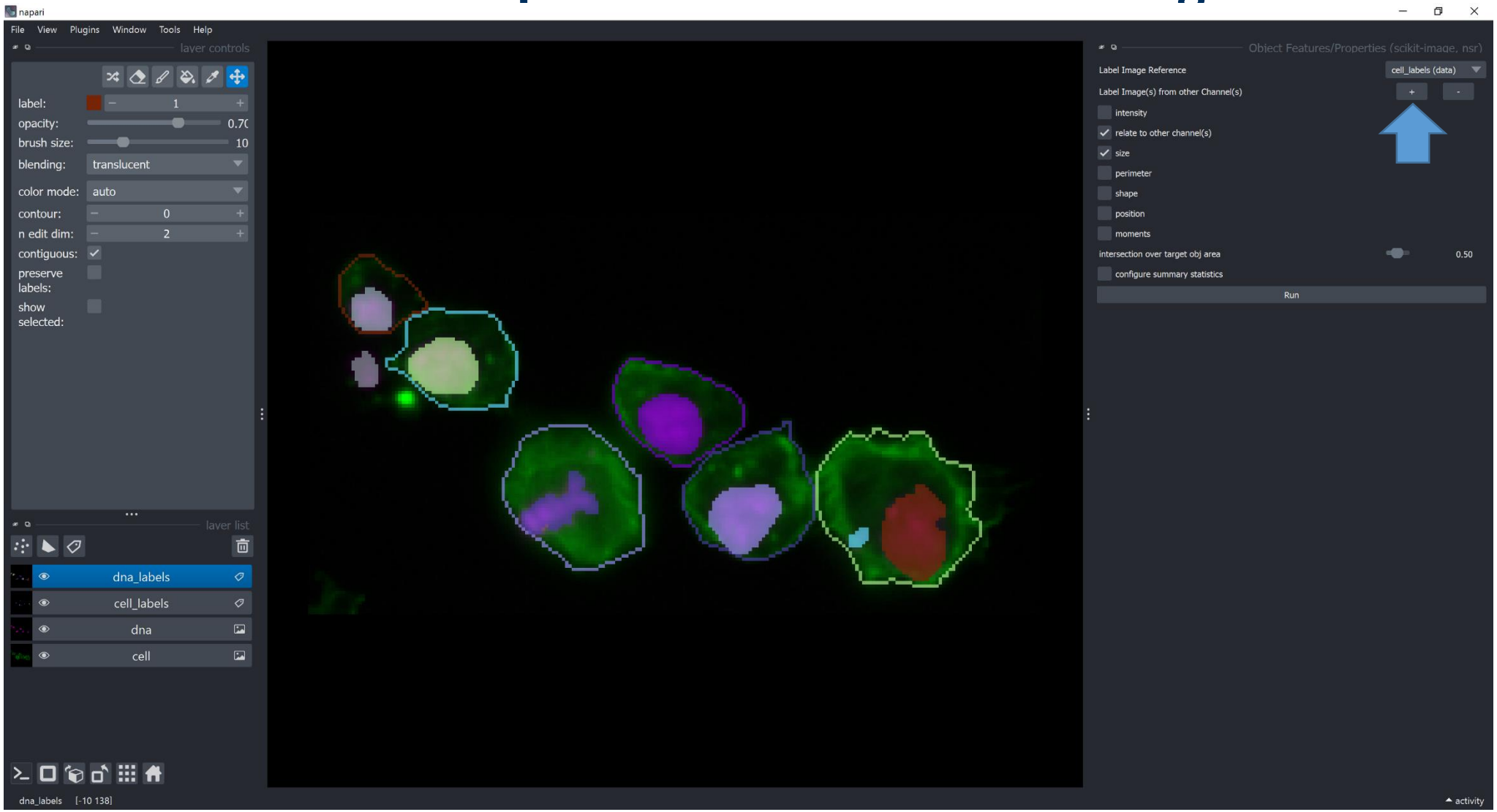

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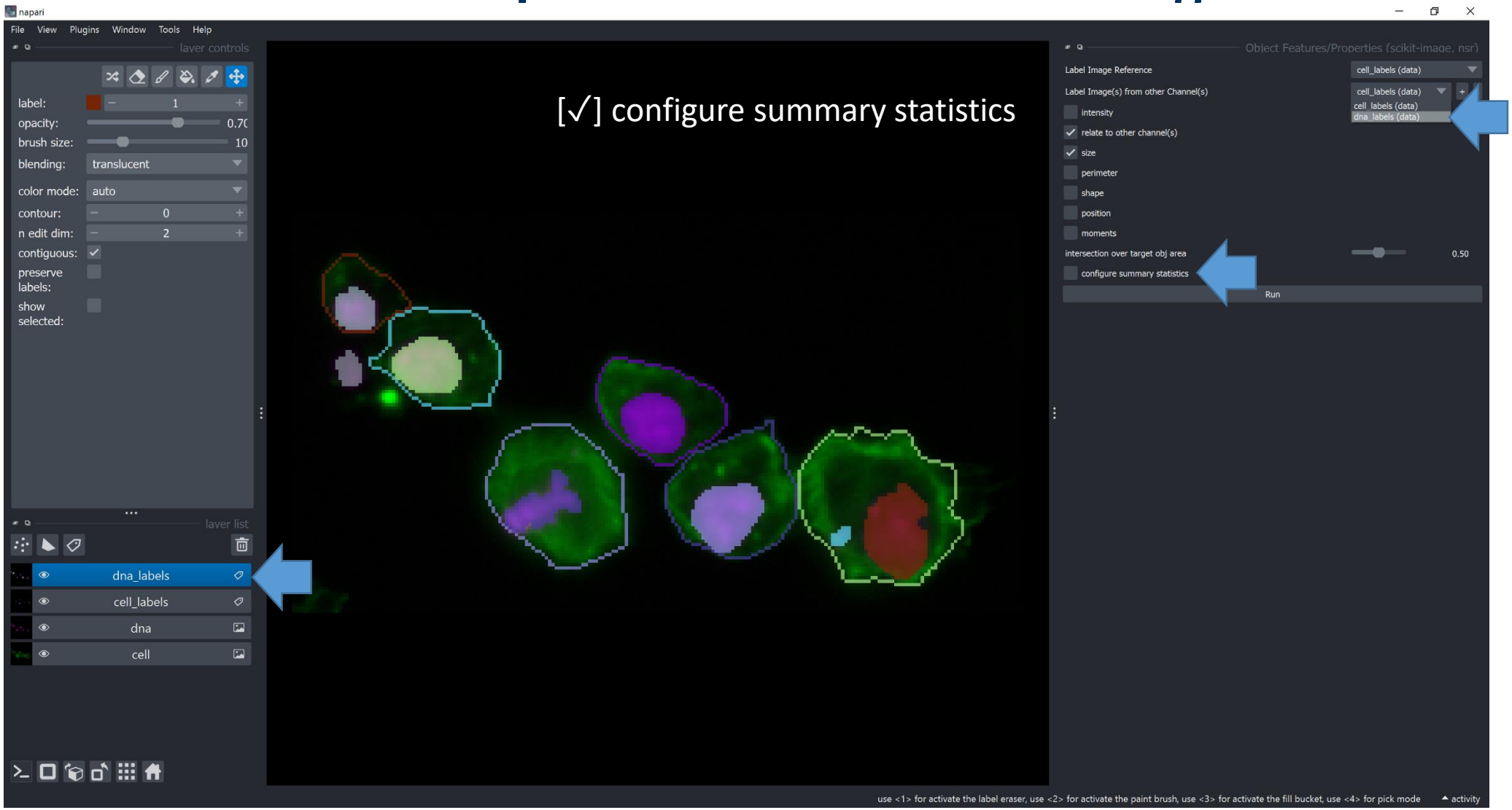

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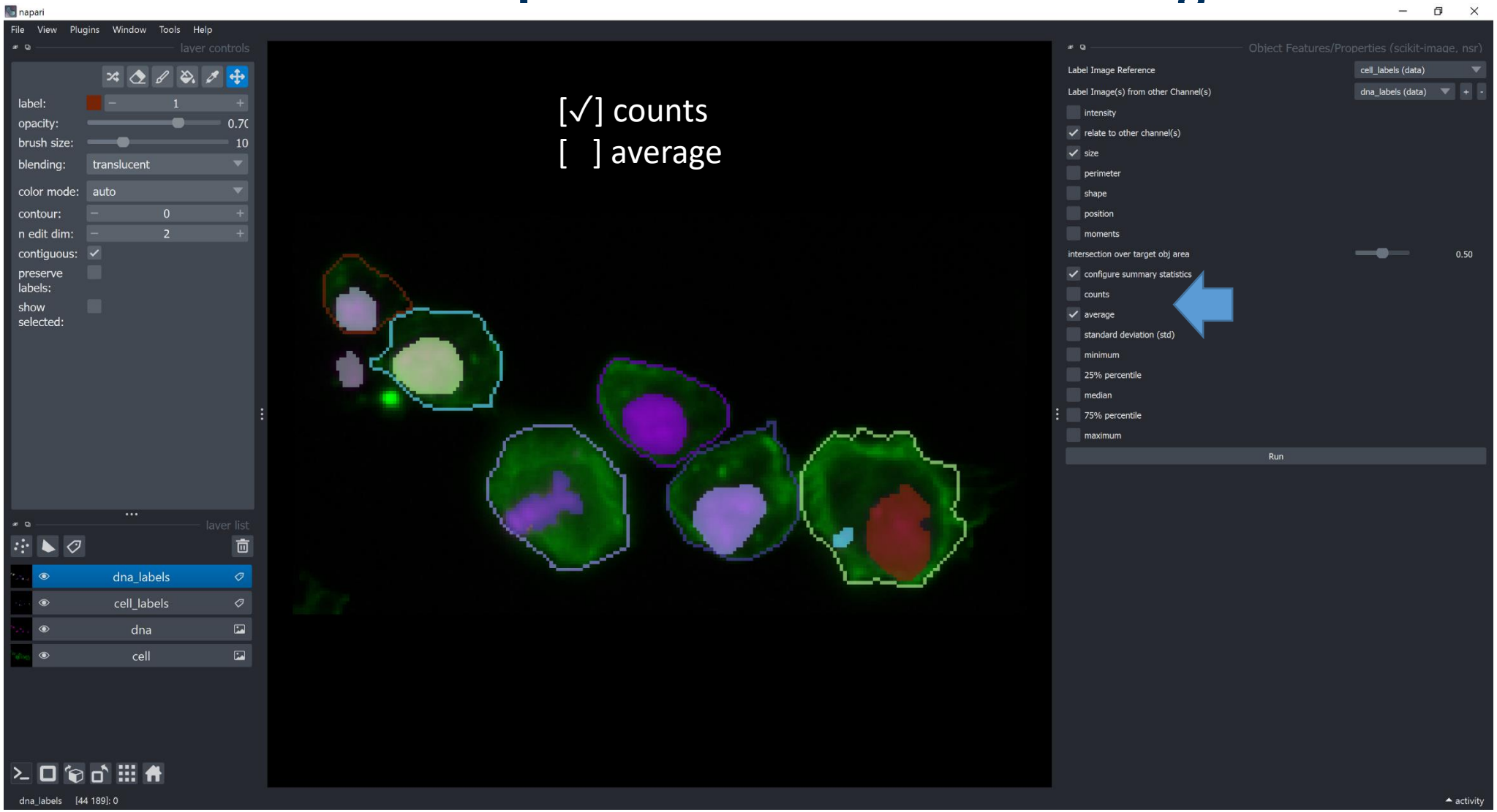

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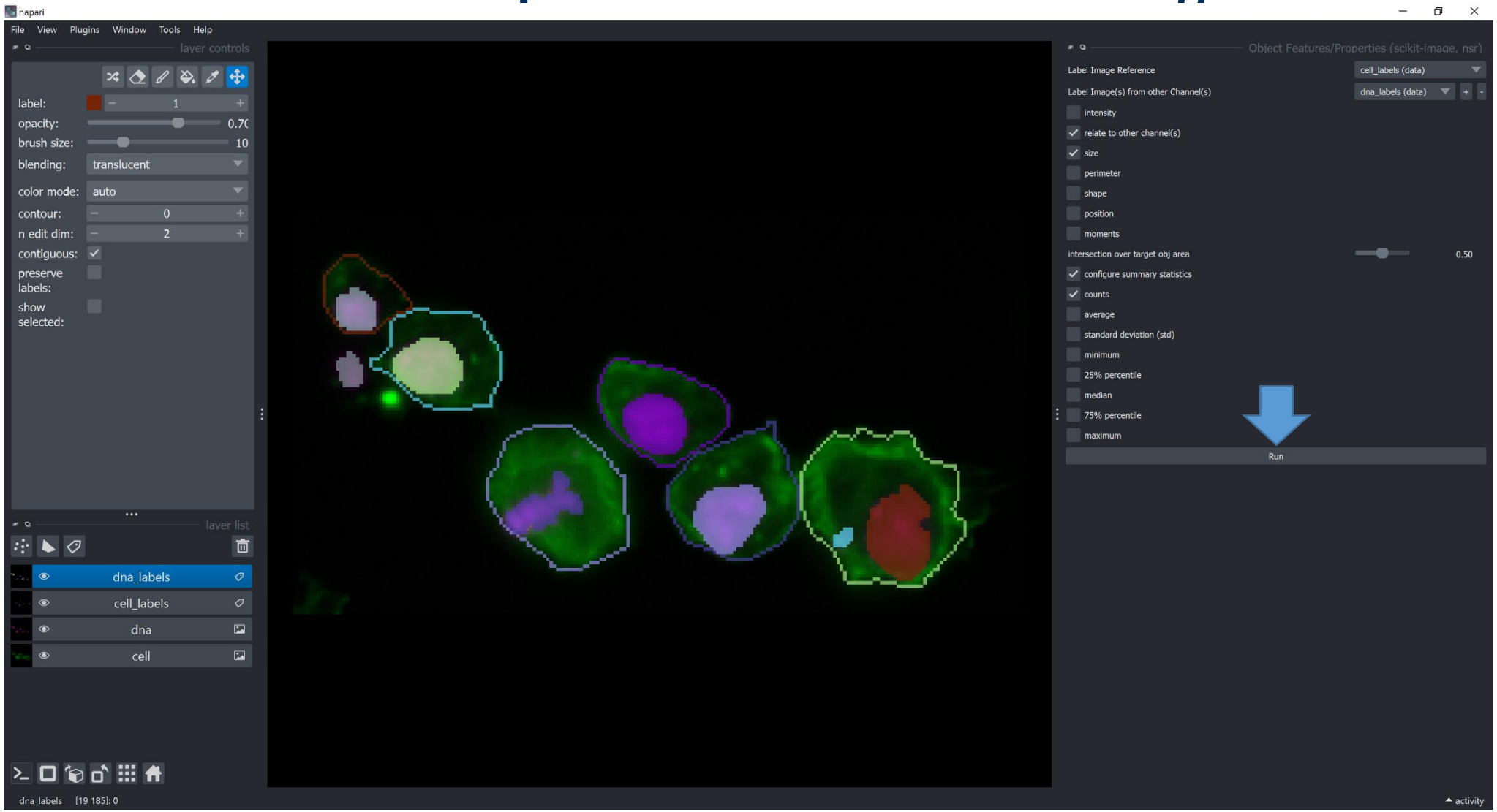

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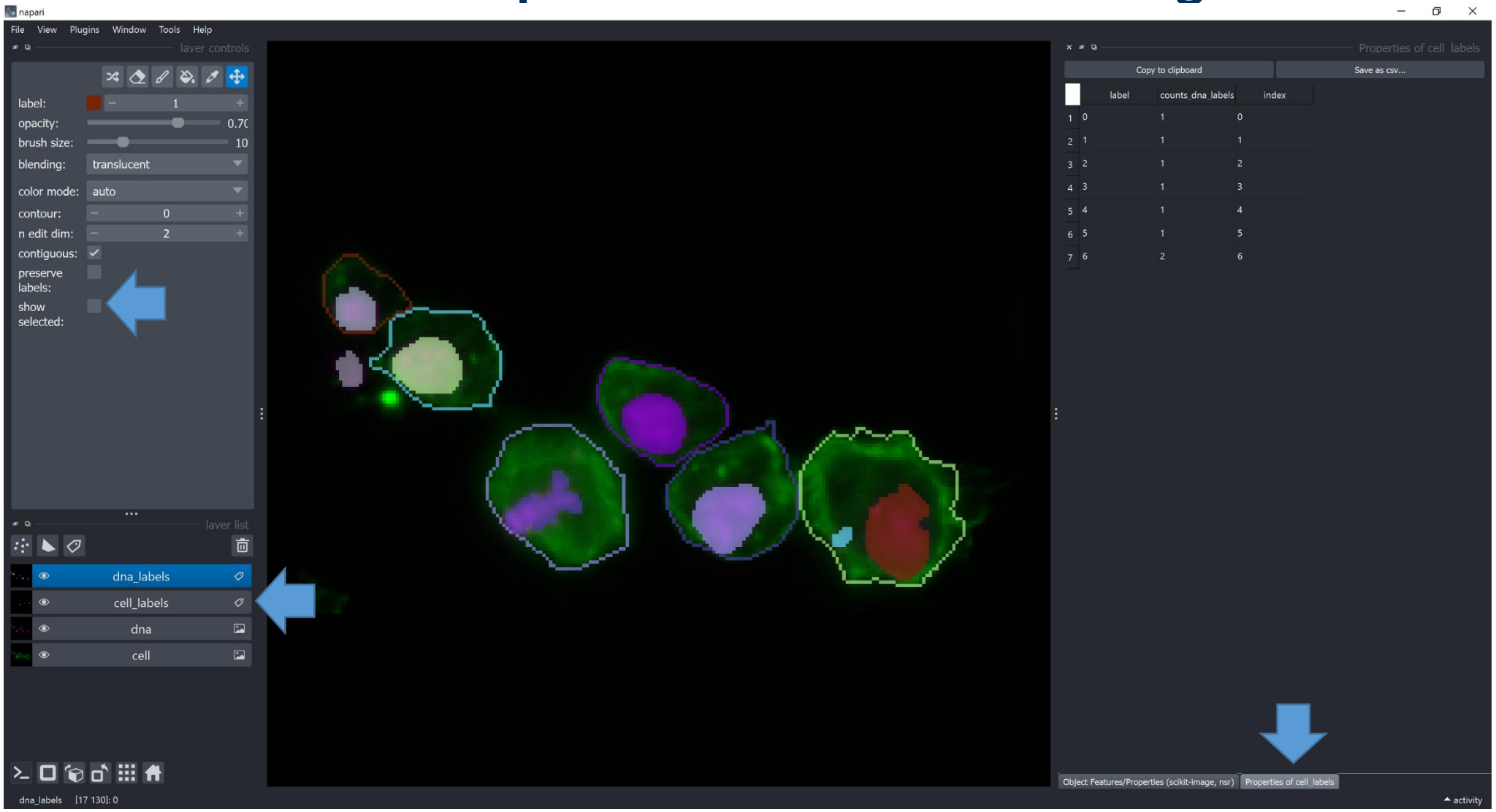

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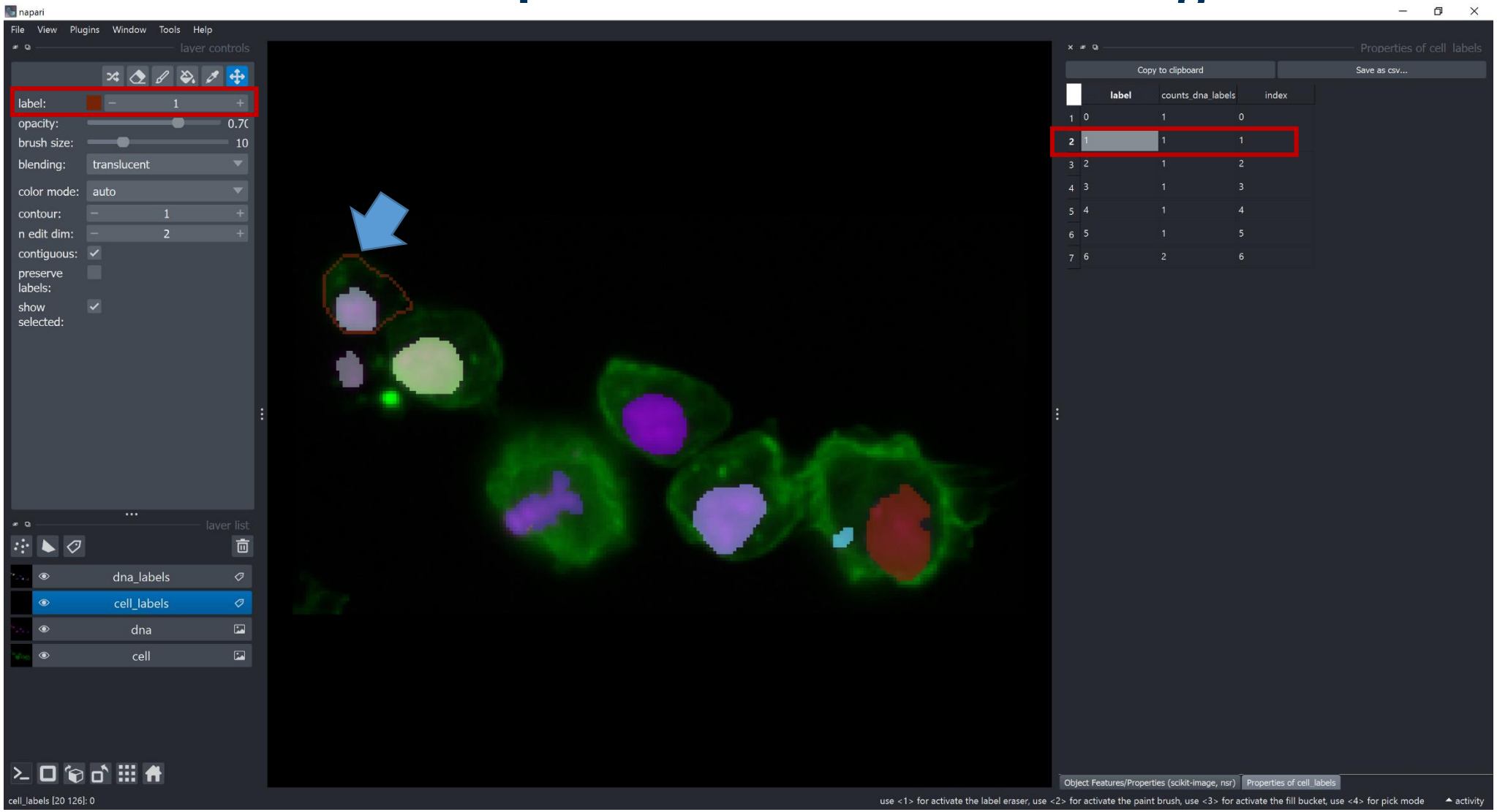

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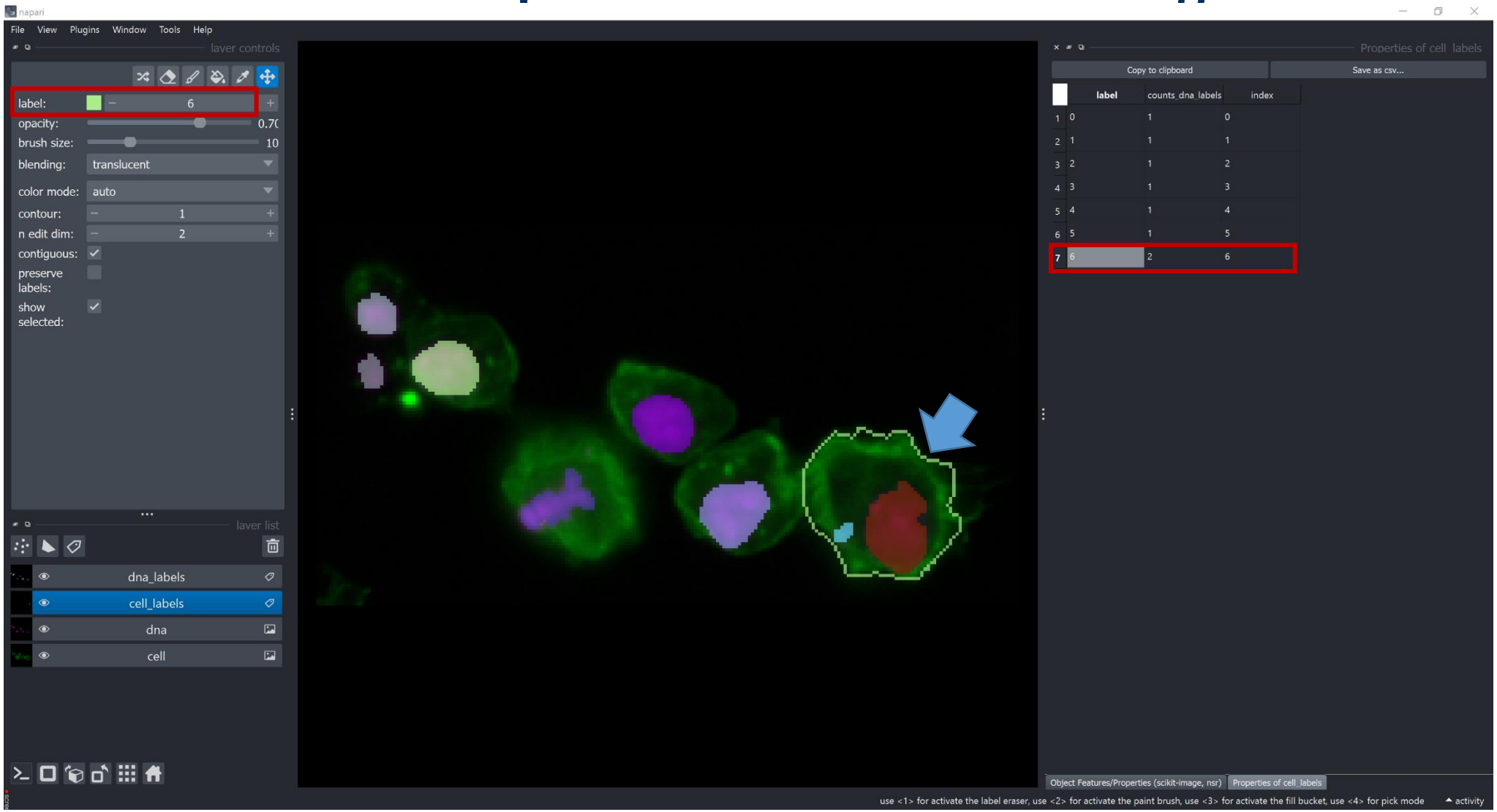

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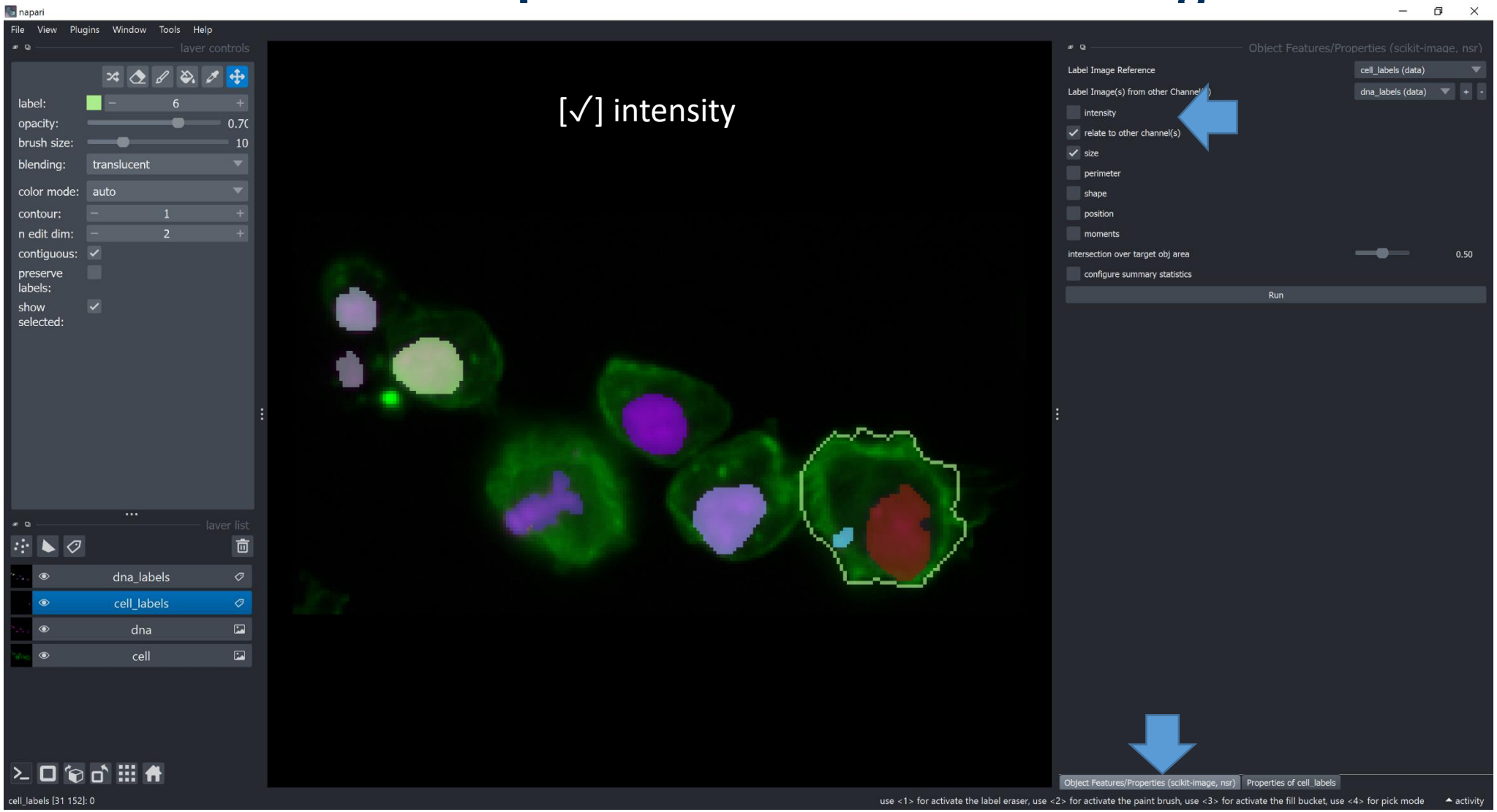

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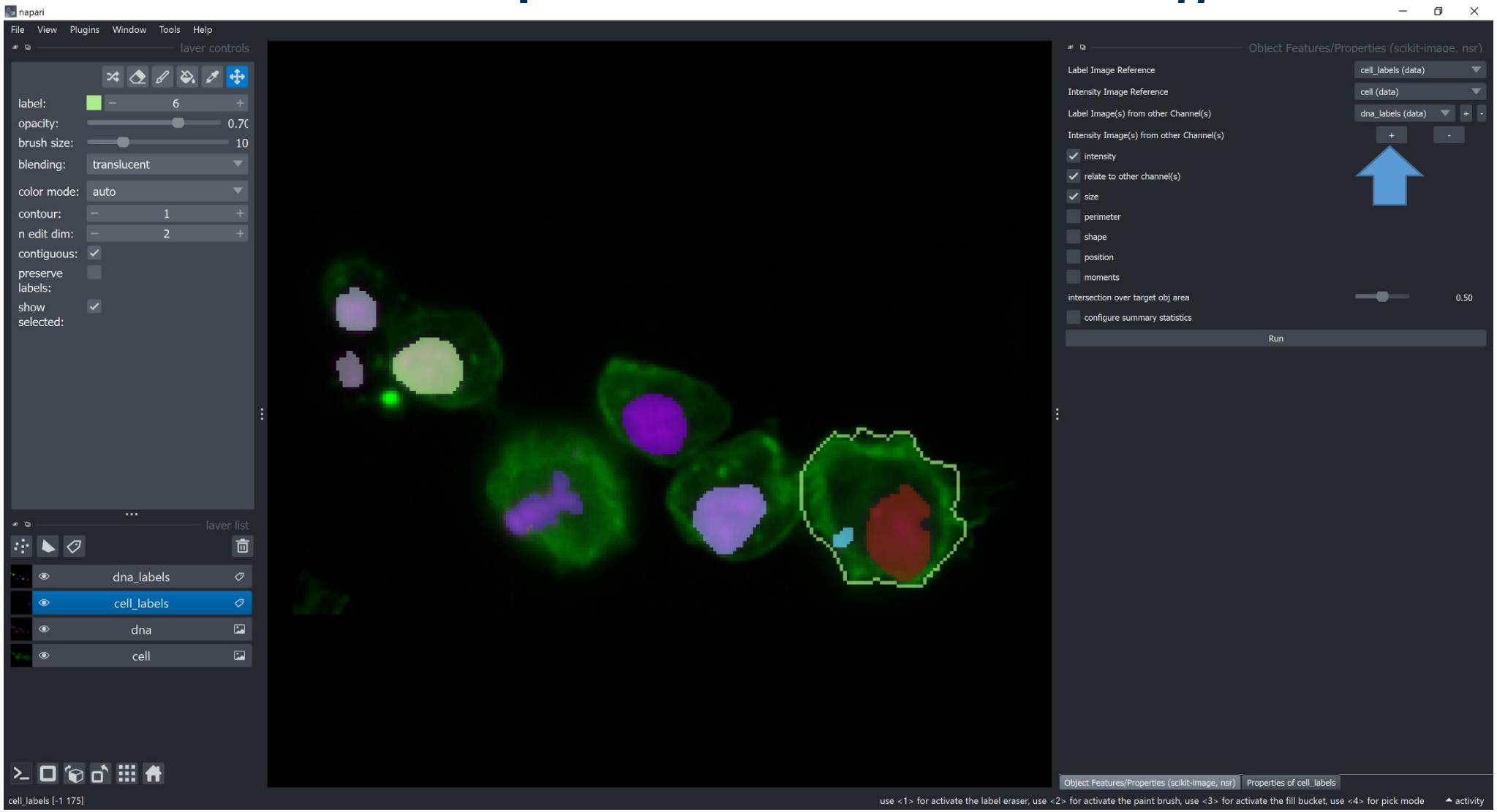

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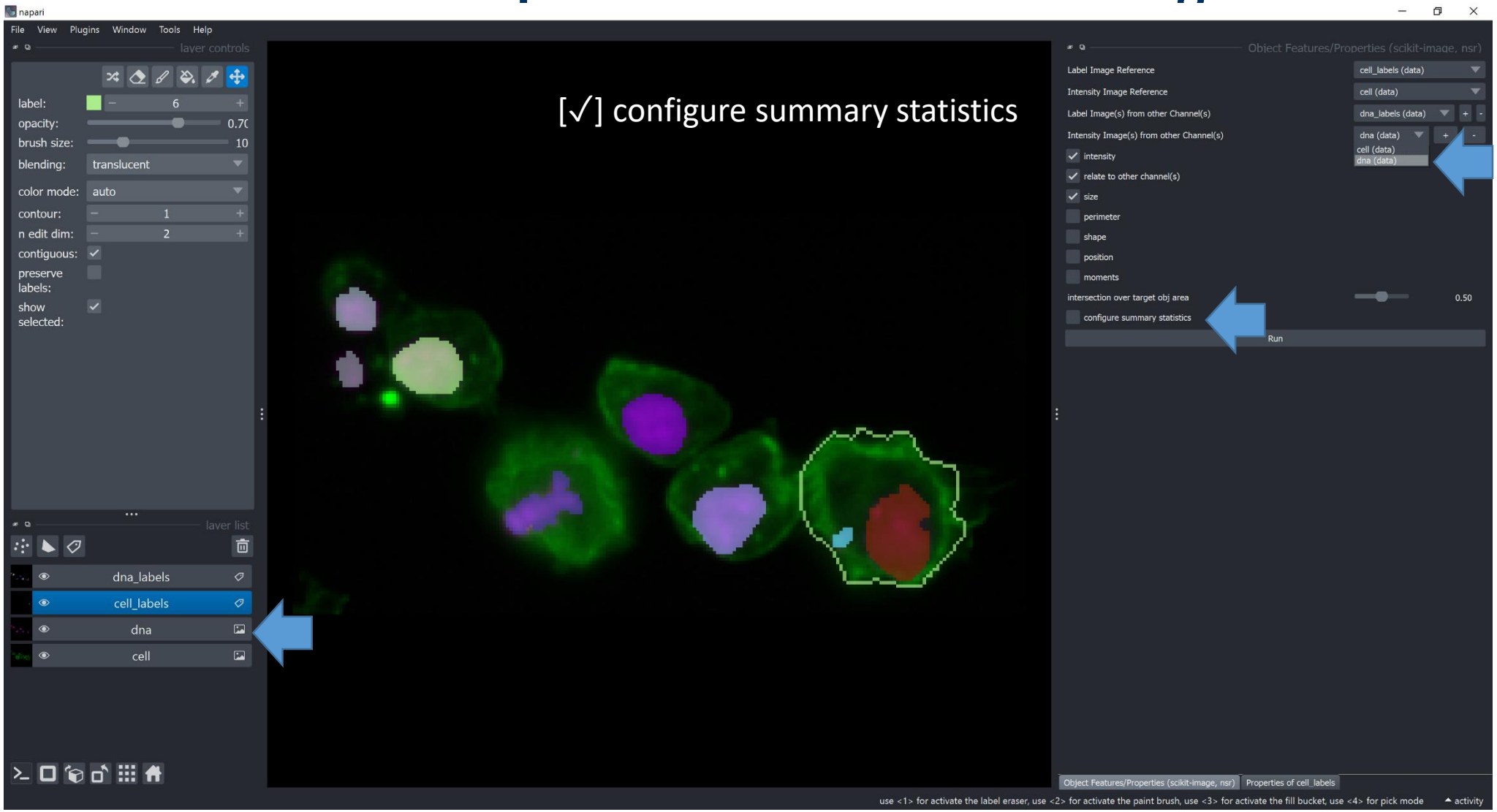

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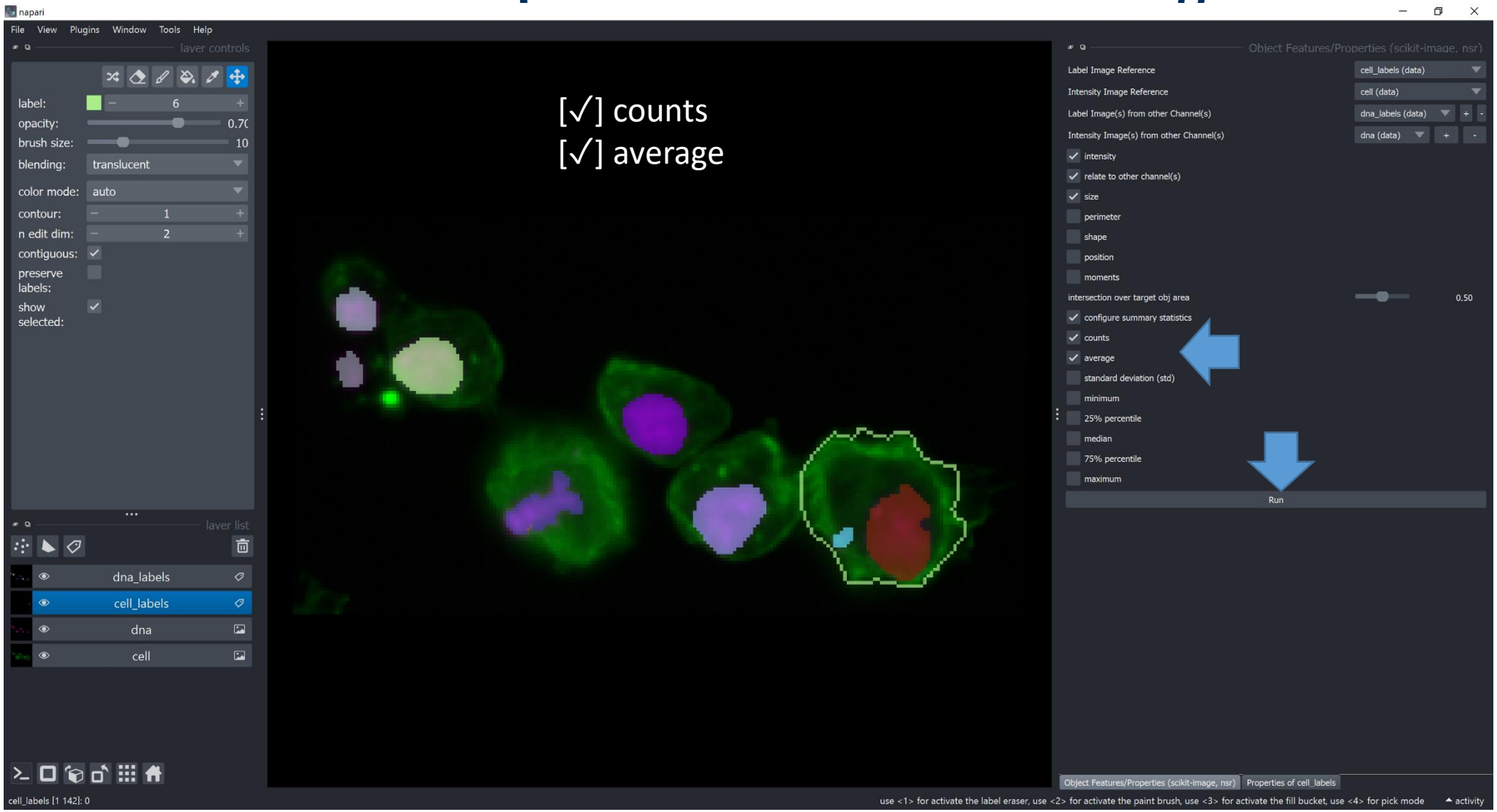

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DRESDEN concept

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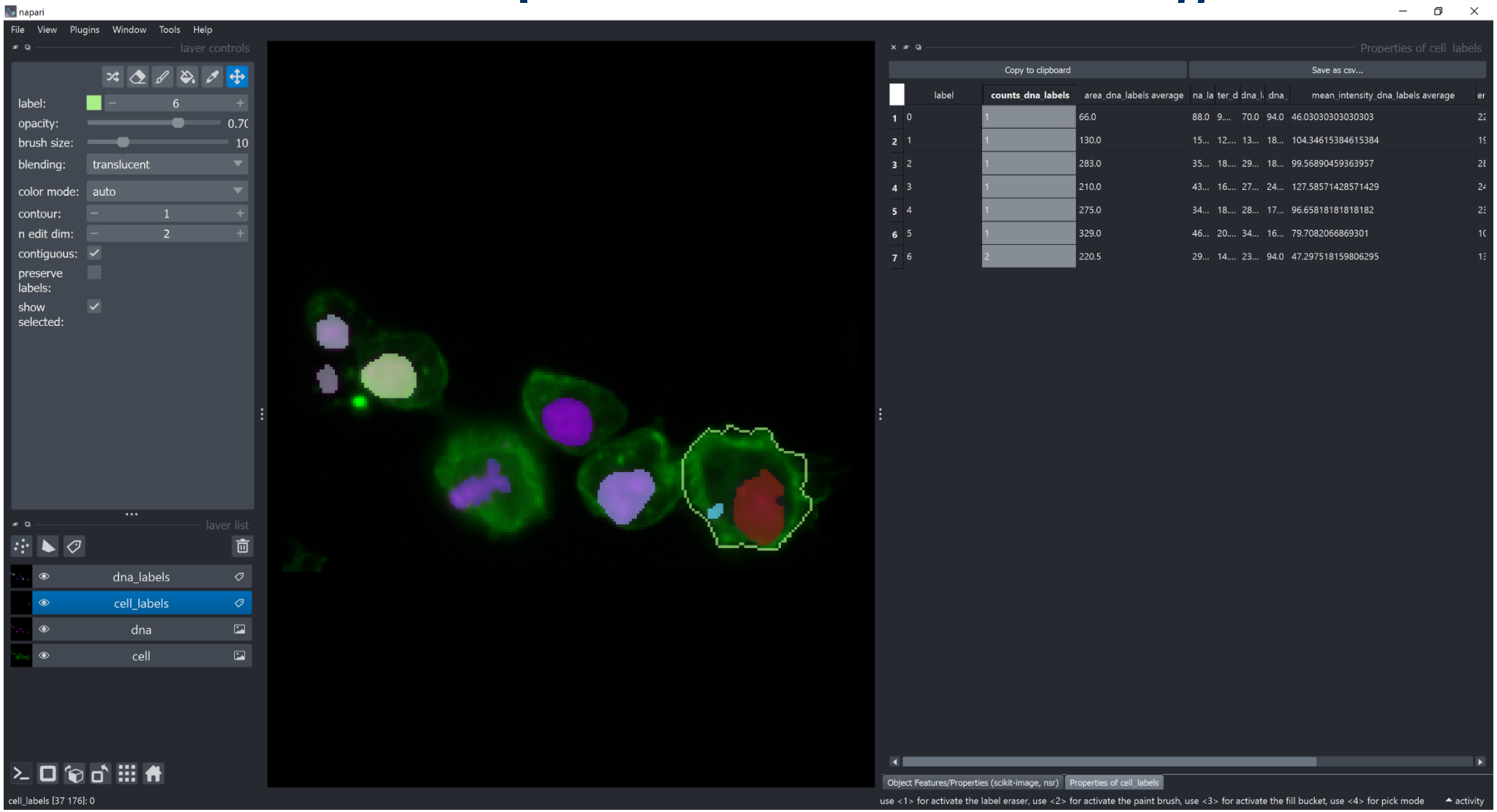

https://github.com/haesleinhuepf/napari-skimage-regionprops

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DRESDEN concept

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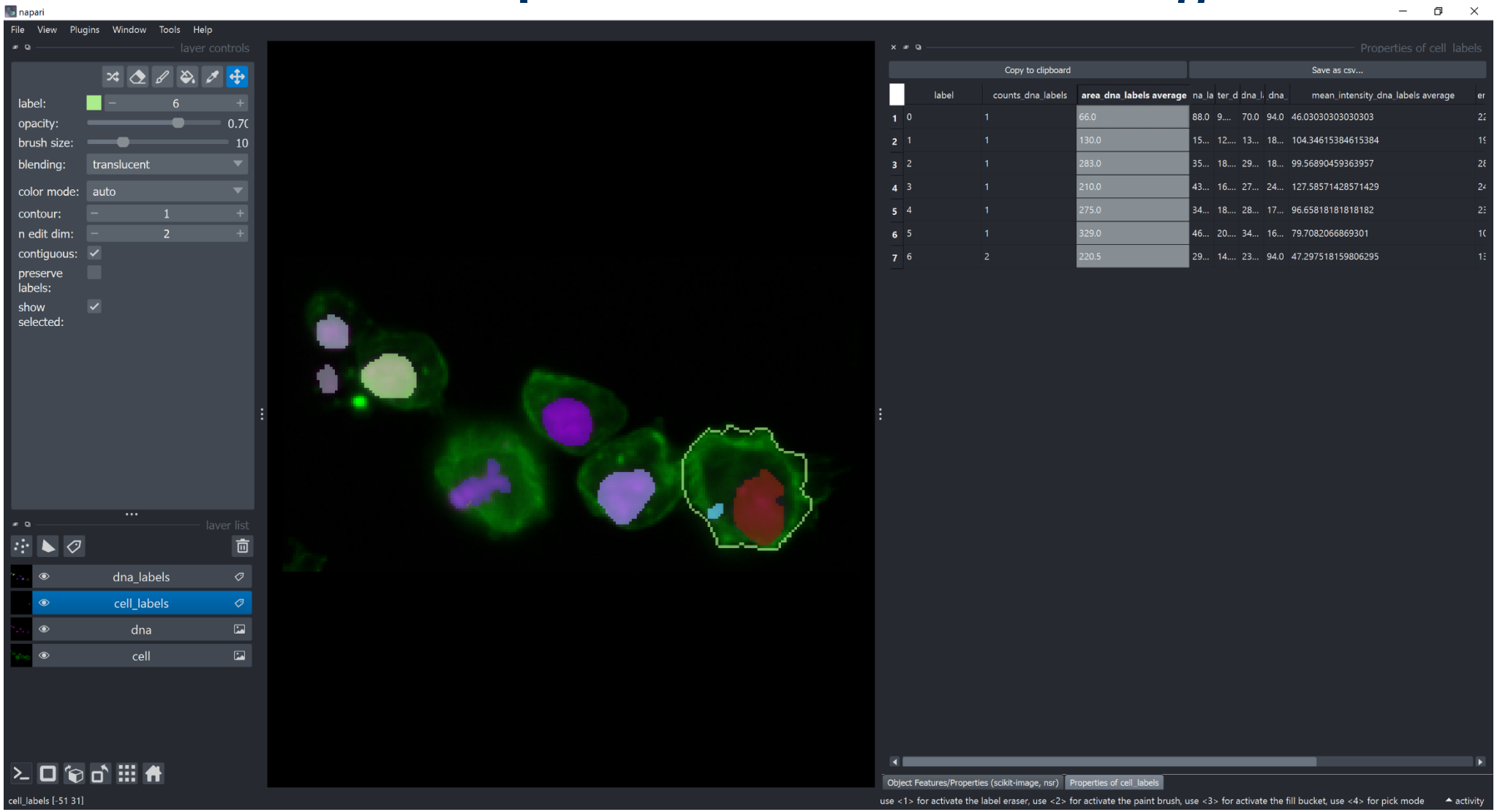

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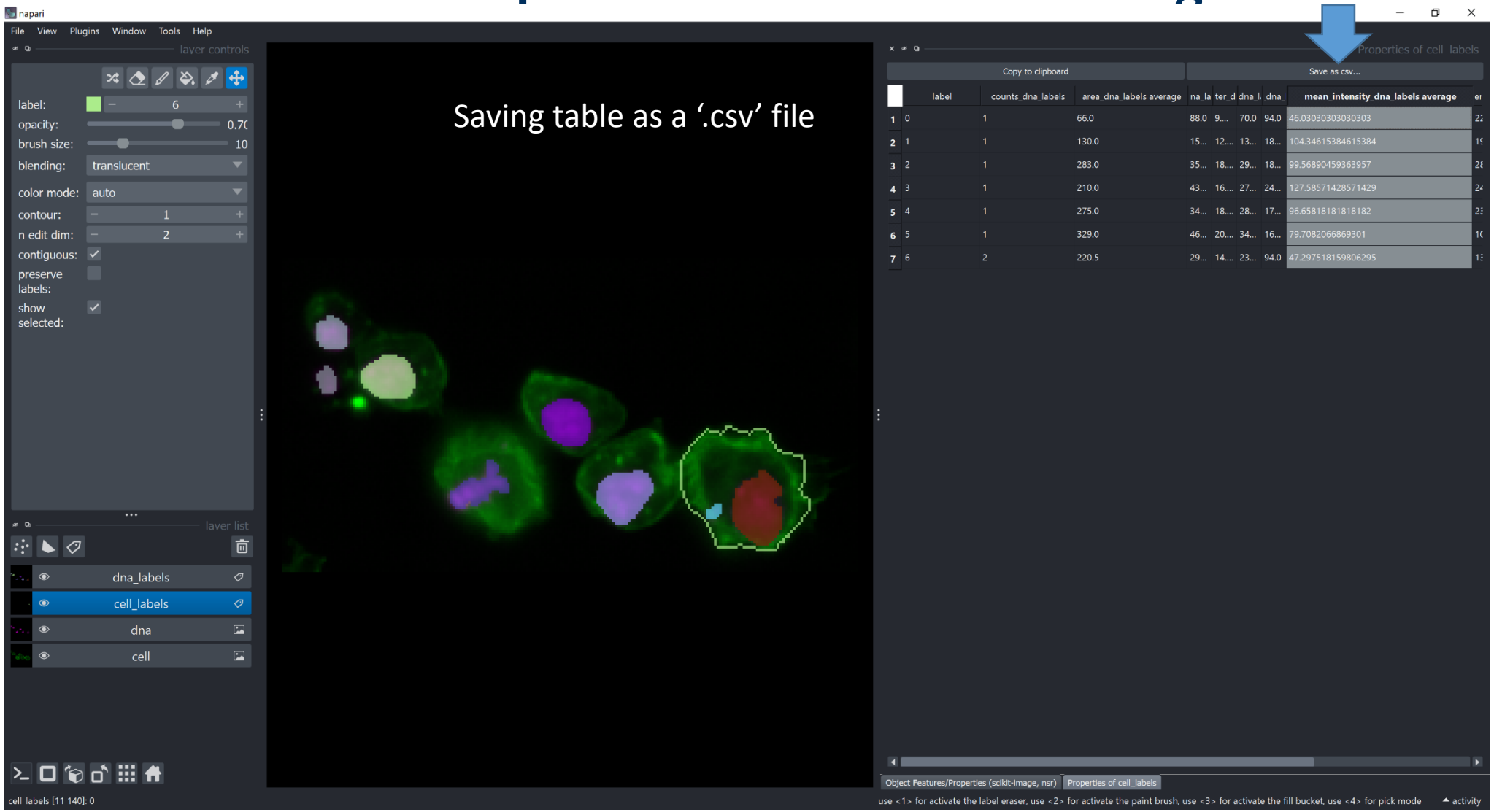

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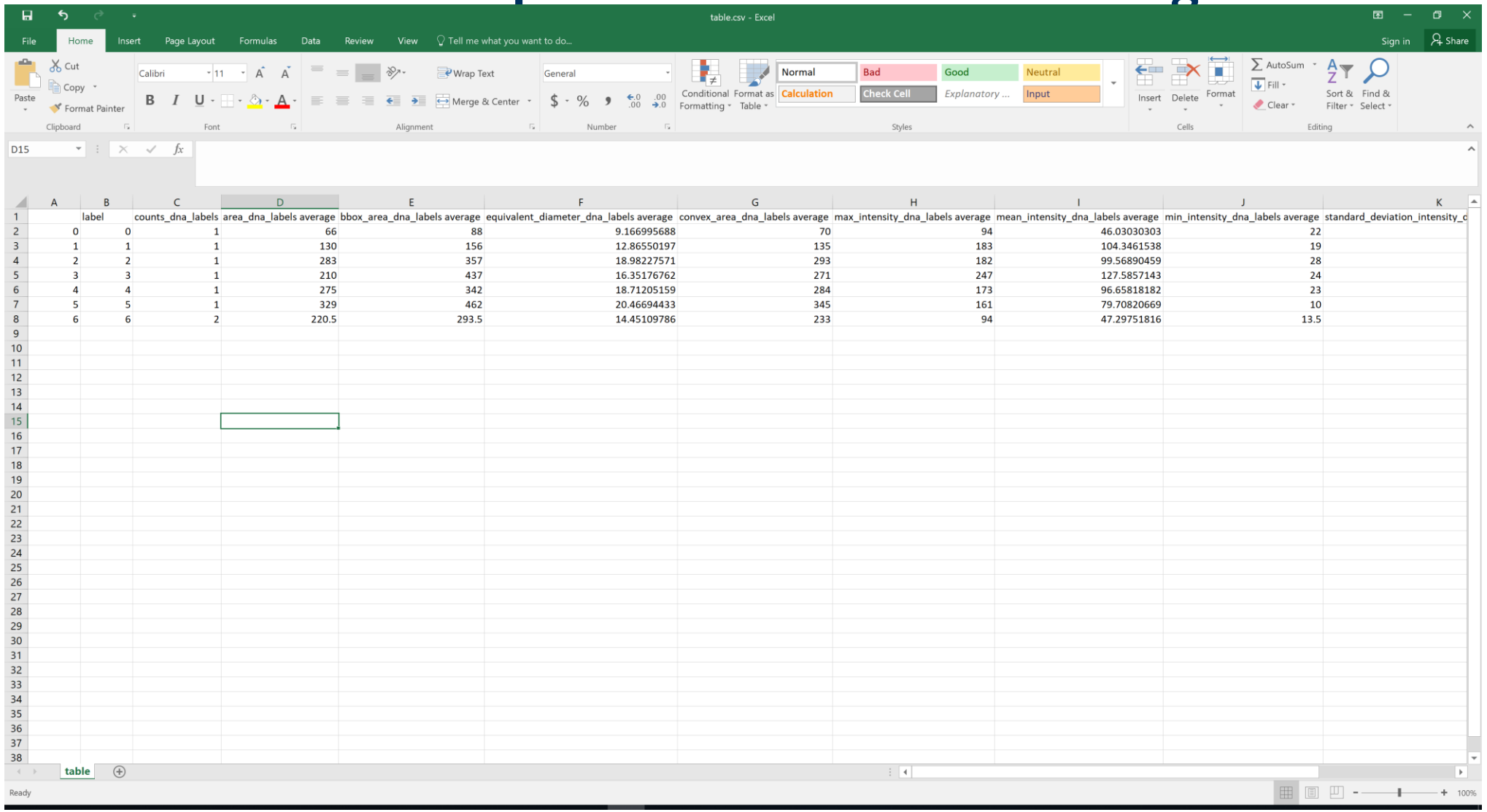

table.csv - Excel

	A	B	C	D	E	F	G	H	I	J	K
1		label	counts_dna_labels	area_dna_labels average	bbox_area_dna_labels average	equivalent_diameter_dna_labels average	convex_area_dna_labels average	max_intensity_dna_labels average	mean_intensity_dna_labels average	min_intensity_dna_labels average	standard_deviation_intensity_c
2	0	0	1	66	88	9.166995688	70	94	46.03030303	22	
3	1	1	1	130	156	12.86550197	135	183	104.3461538	19	
4	2	2	1	283	357	18.98227571	293	182	99.56890459	28	
5	3	3	1	210	437	16.35176762	271	247	127.5857143	24	
6	4	4	1	275	342	18.71205159	284	173	96.65818182	23	
7	5	5	1	329	462	20.46694433	345	161	79.70820669	10	
8	6	6	2	220.5	293.5	14.45109786	233	94	47.29751816	13.5	

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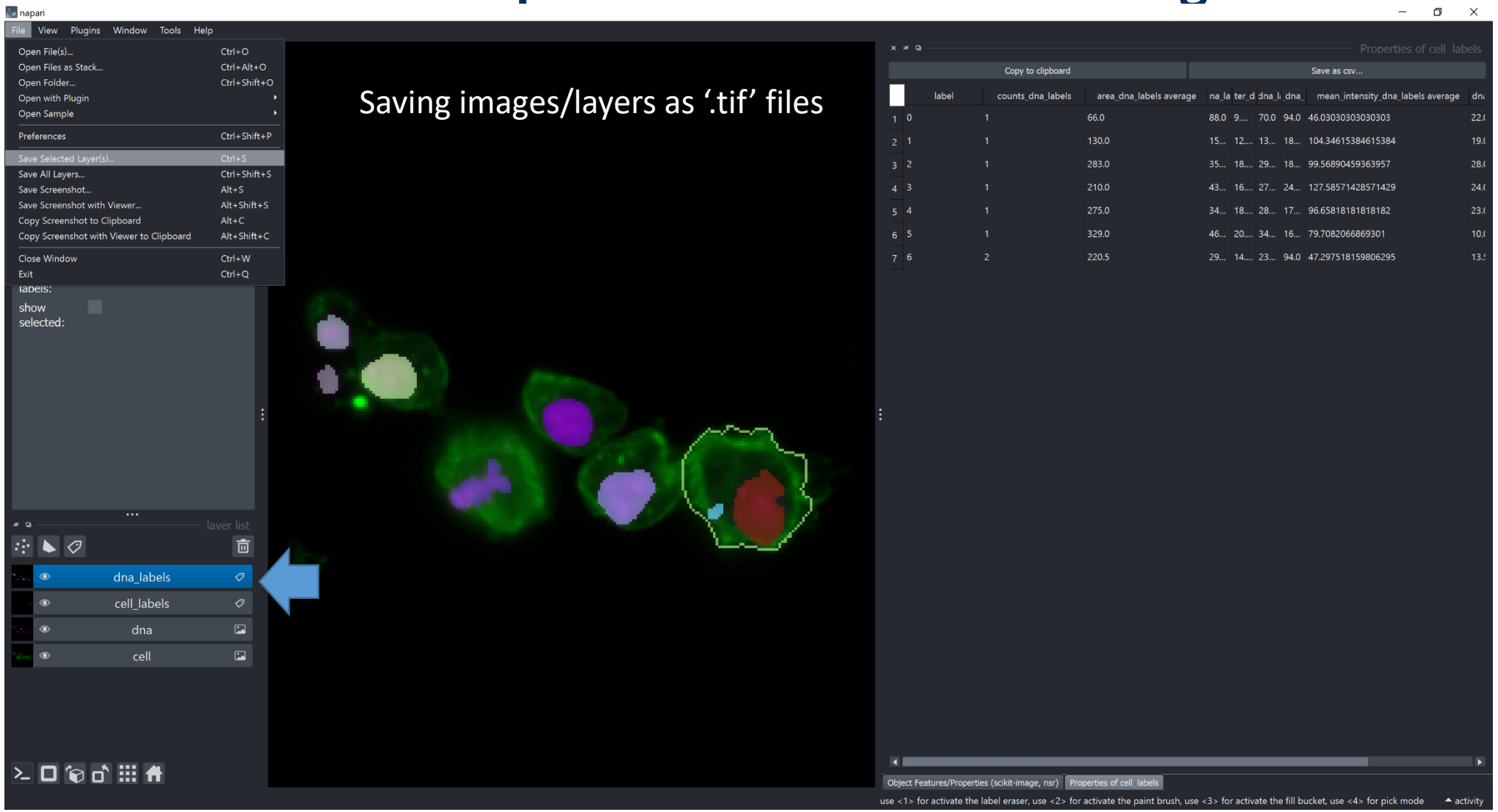

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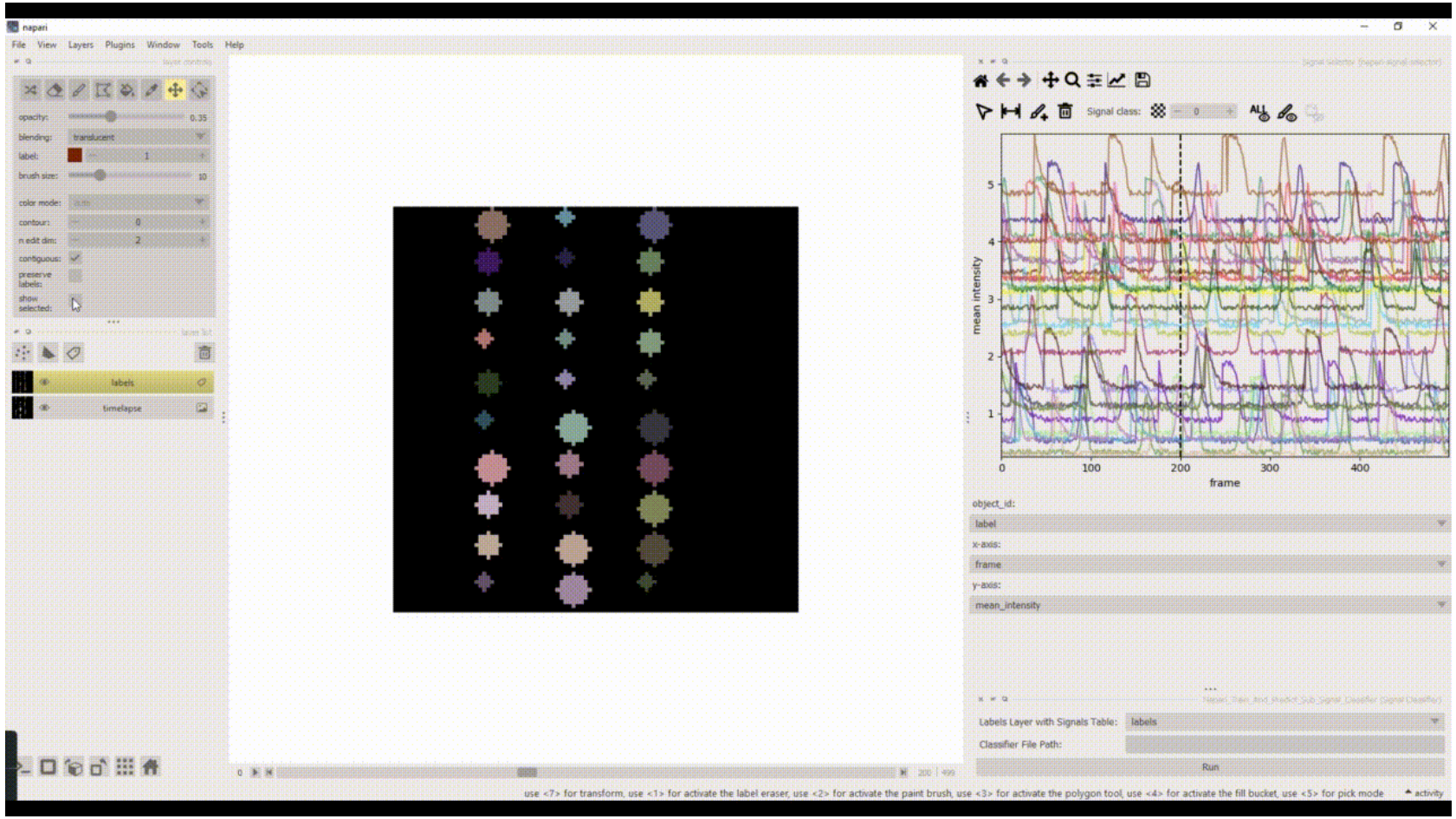
Outlook: Temporal Features Annotation and Classification



- https://github.com/zoccoler/napari-signal-selector
  - https://github.com/zoccoler/napari-signal-classifier

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Object Classification and Supervised Machine Learning in napari

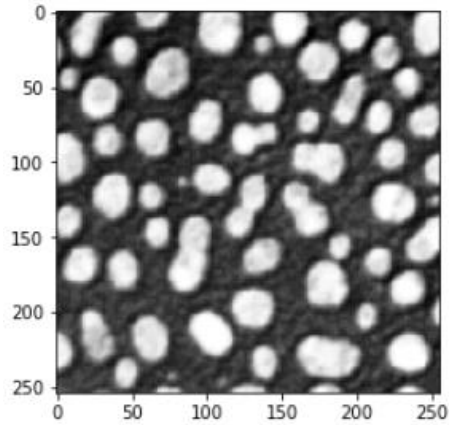
Random Forest Classifiers

- Object Classifier
- **napari-apoc plugin**

DRESDEN concept

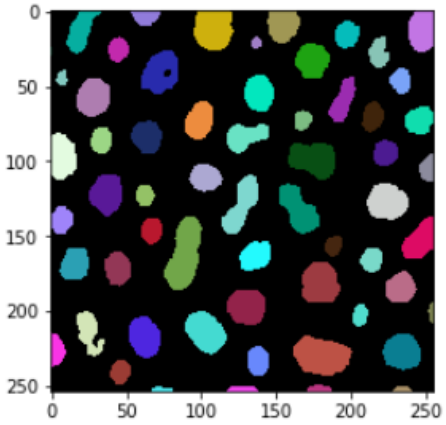
Object classification

- Object classification is a task that can be applied to an existing instance segmentation in order to identify a particular group of objects



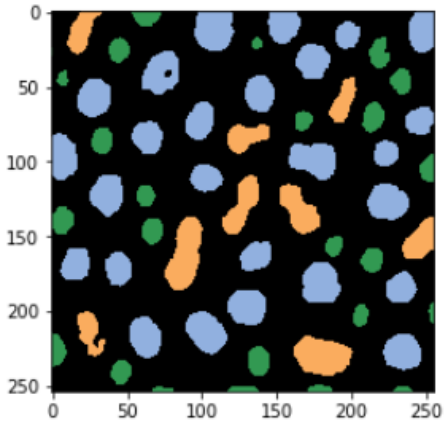
Raw image

Object segmentation

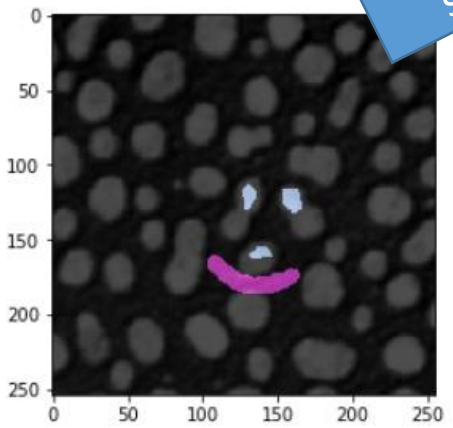


Object label image

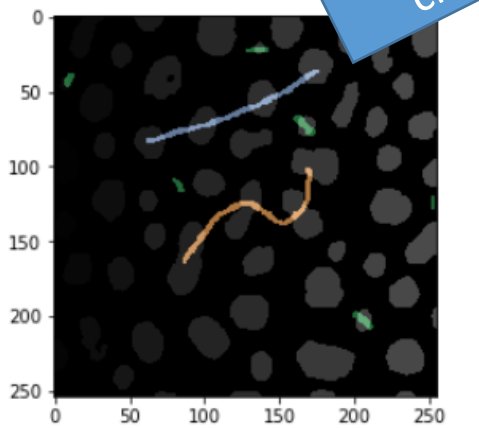
Object classification



Class label image



Pixel annotation



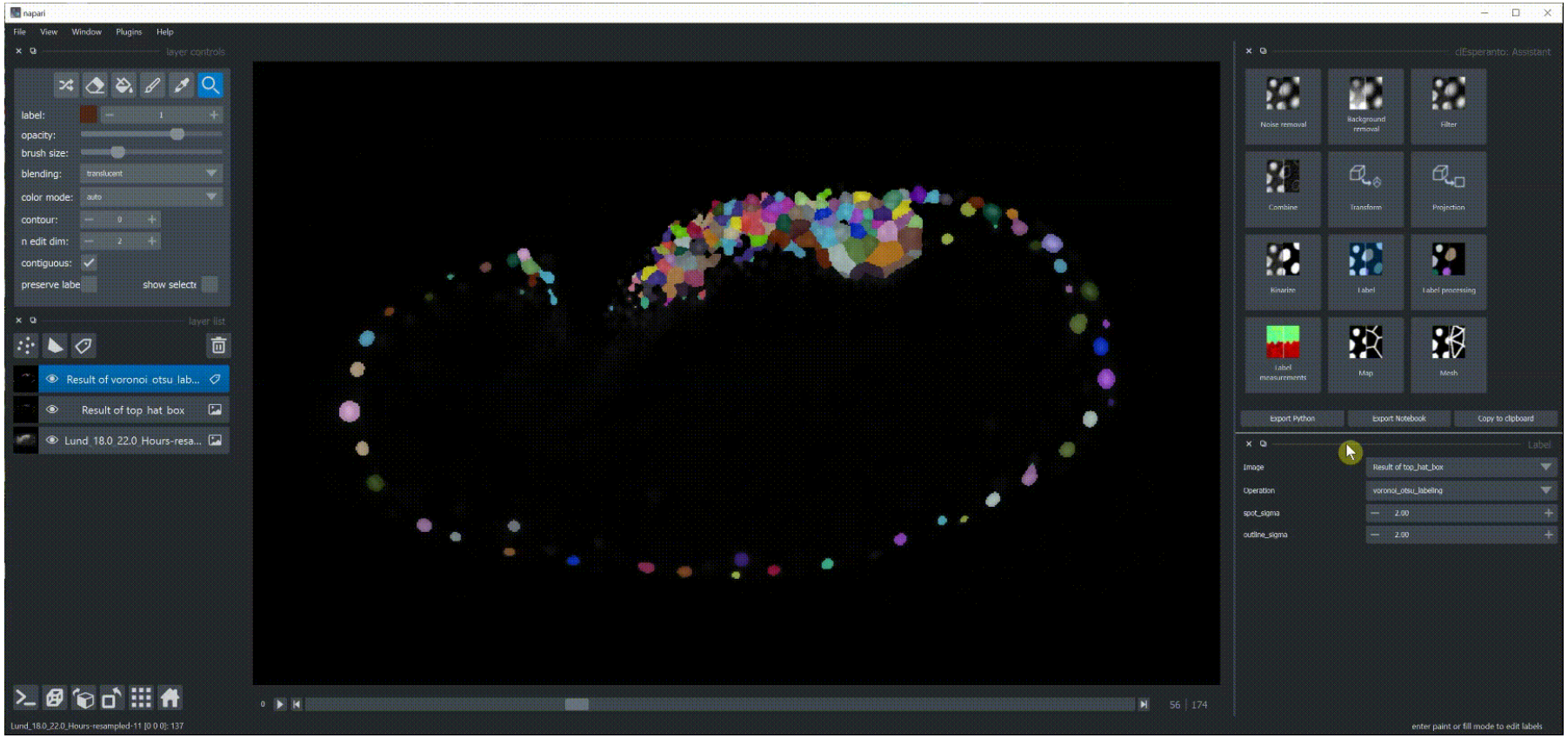
Object annotation

https://imagej.nih.gov/ij/images/

Object classification

Random Forest Classifiers based on

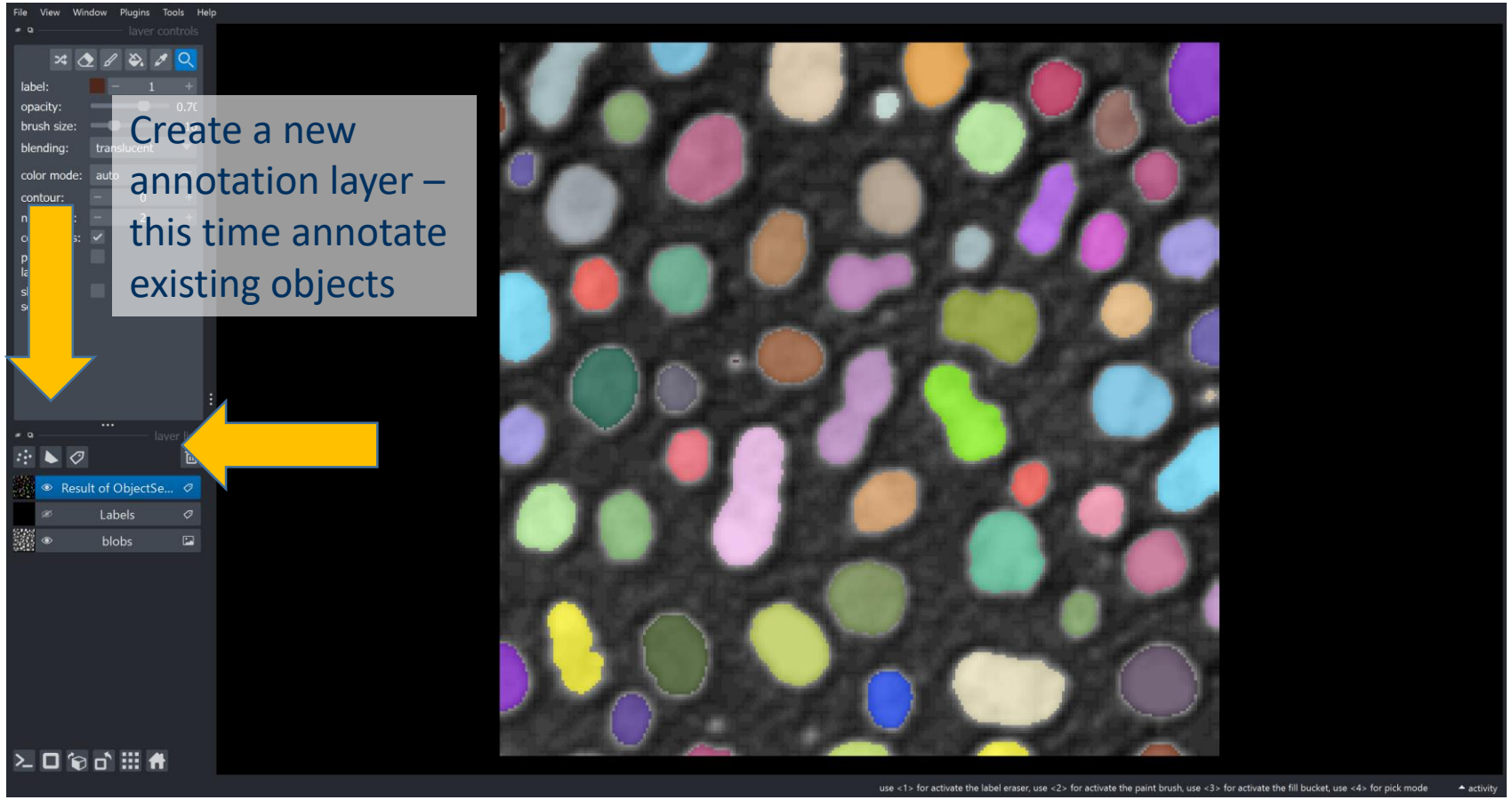
- scikit-learn and
- clesperanto



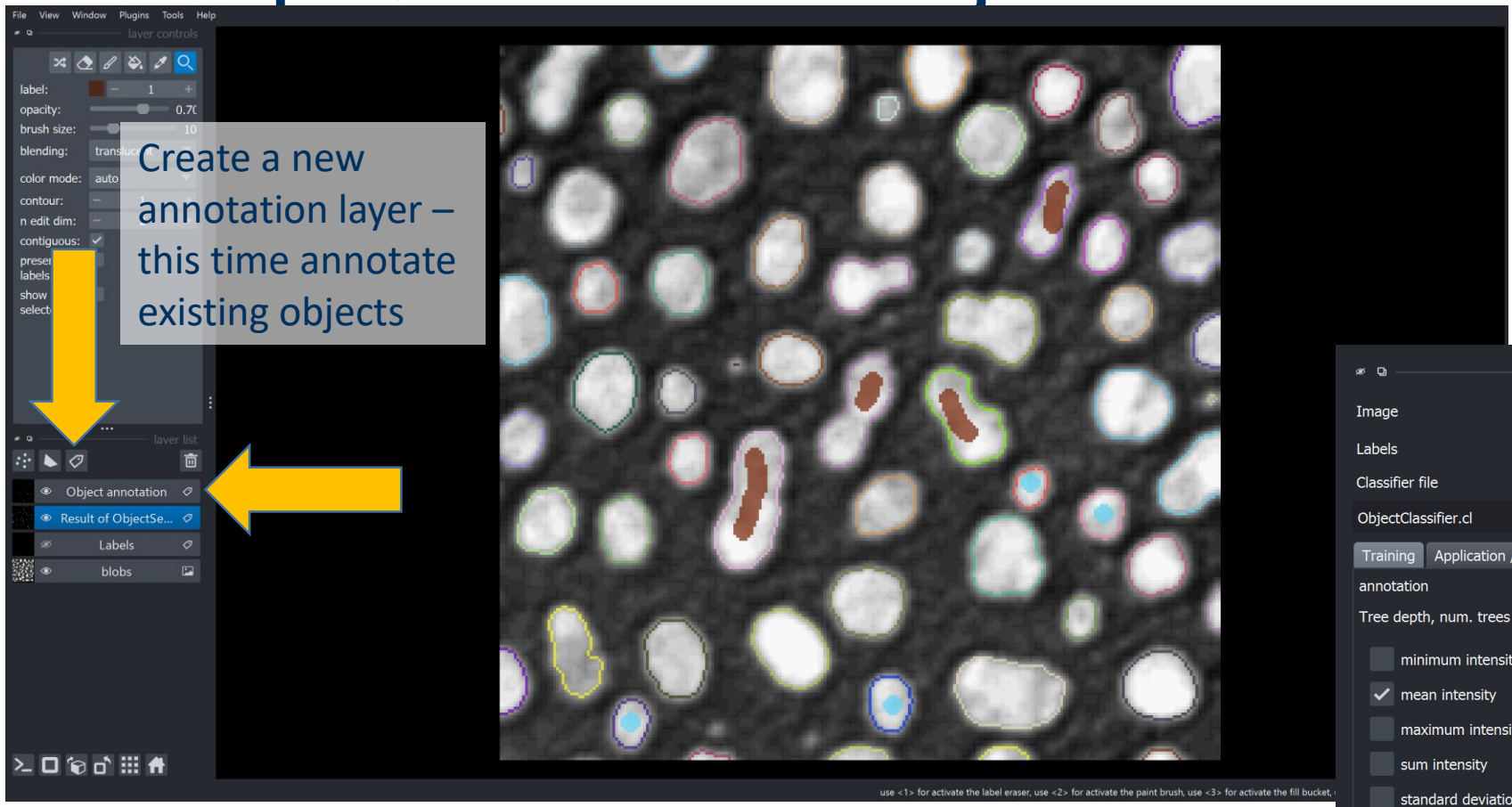
https://github.com/haesleinhuepf/napari-accelerated-pixel-and-object-classification

Image data source: Daniela Vorkel, Myers lab, MPI-CBG/CSBD

In napari: annotation

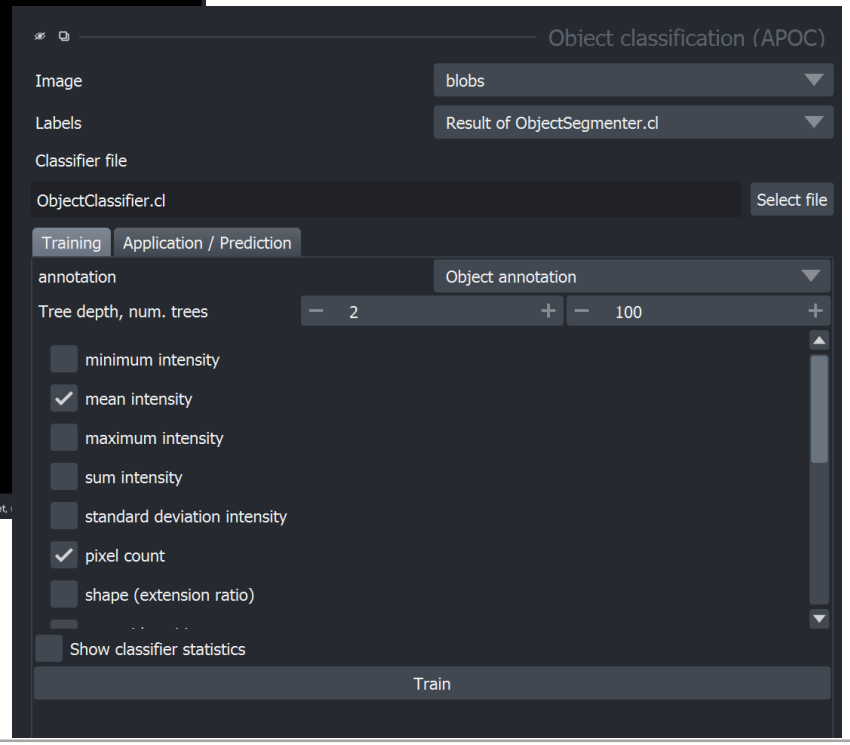


In napari: annotation and object classification



Create a new annotation layer – this time annotate existing objects

Tools ->
Segmentation post-processing ->
Object Classification (APOC)



Telling different classes of objects apart requires:

- An **annotation** for some example objects
- **Features** for each objects upon which to make a prediction

https://imagej.nih.gov/ij/images/

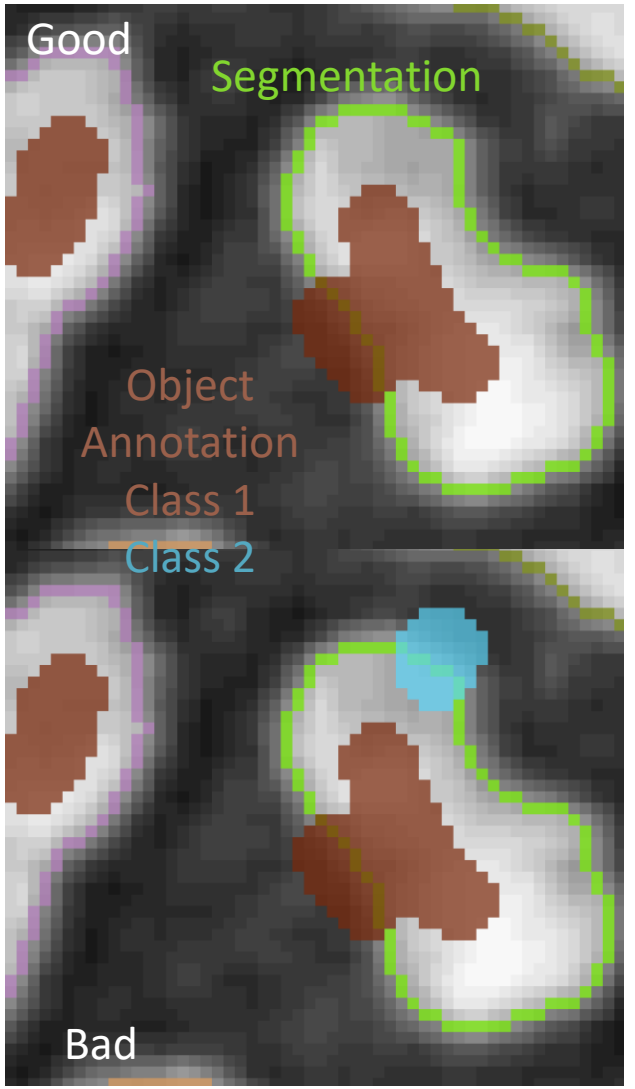
Exercise: Object Classification

1. Activate the environment and open napari
   ```
   mamba activate napari25
   napari
   ```
2. Train an object classifier on sample data: Differentiate **good nuclei** (nicely separated from other objects) from **bad nuclei** (two nuclei sticking together)

•Hints for annotation:

- The annotation **does not have to overlap exactly** with the painted object
- Every annotation should **only touch** the correct objects
- **Save your annotations**
- In this case, it makes no difference whether we annotate many or few pixels. The number of annotated objects is more important in this context.

https://biapol.github.io/BioImage-Analysis-and-Data-Processing-Workshop-2025/interactive_object_classification/readme.html



https://imagej.nih.gov/ij/images/

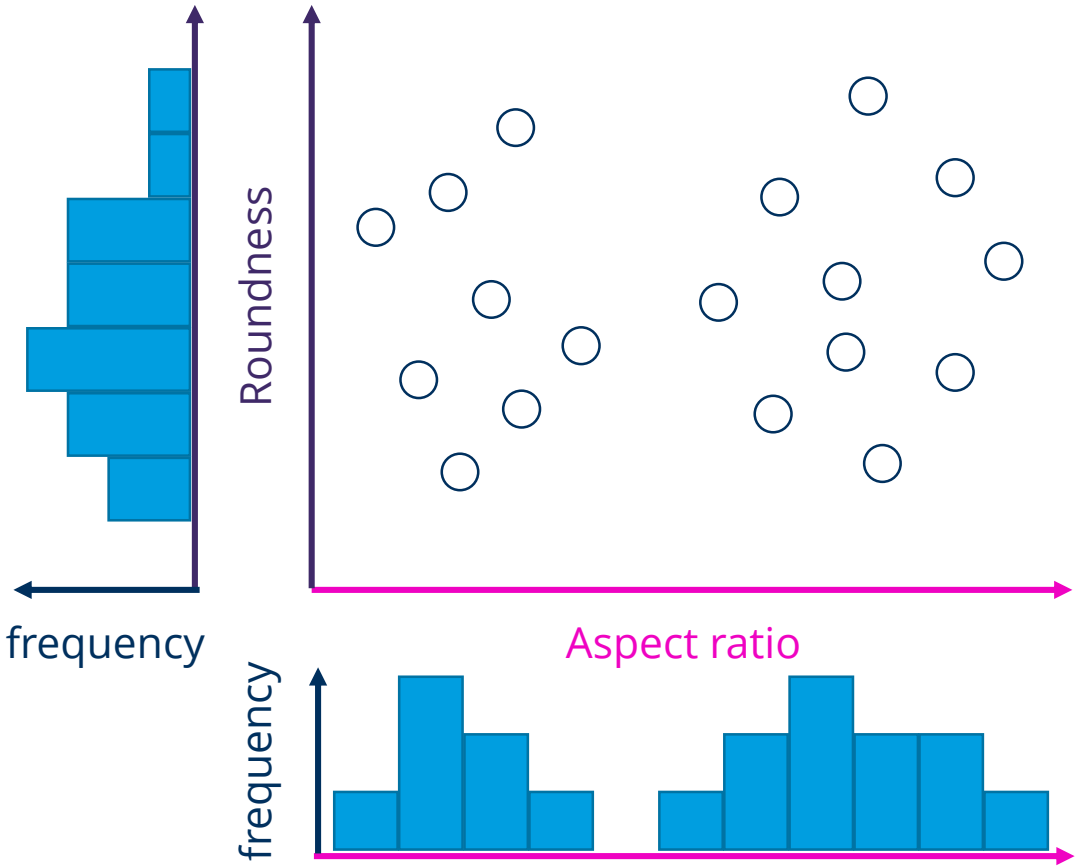

A smaller
break

Object Classification and Unsupervised Machine Learning

- Dimensionality Reduction
- Clustering
- **Napari-clusters-plotter plugin**

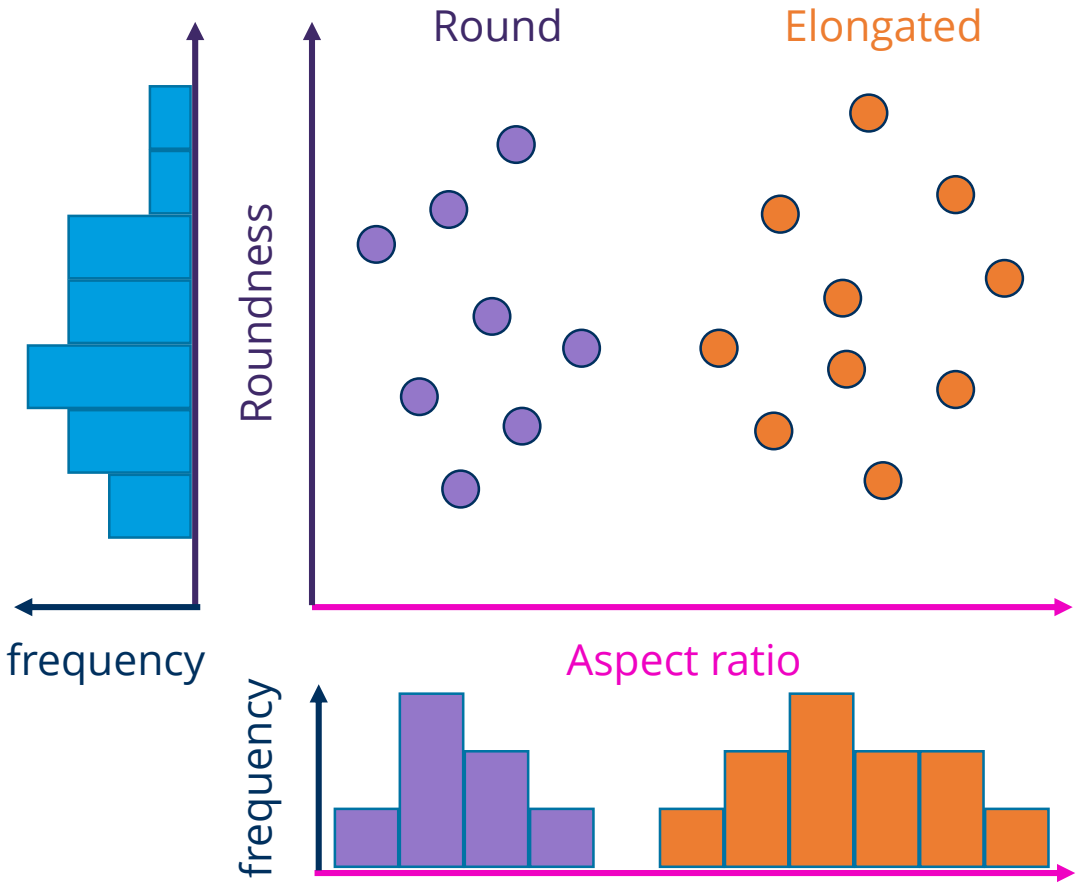
Unsupervised machine learning

If you don’t provide ground truth, the algorithm is *unsupervised*.

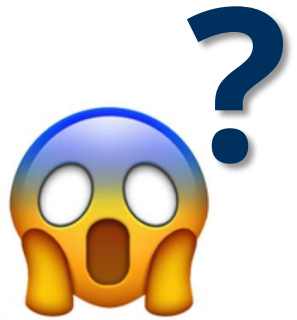


Unsupervised machine learning

If you don’t provide ground truth, the algorithm is unsupervised.
Nevertheless, algorithms can tell us something about the data



- Mean intensity
- Standard deviation
- Total intensity
- Textures
- Area / Volume
- Roundness
- Solidity
- Circularity / Sphericity
- Elongation
- Centroid
- Bounding box
- …



Dimensionality reduction

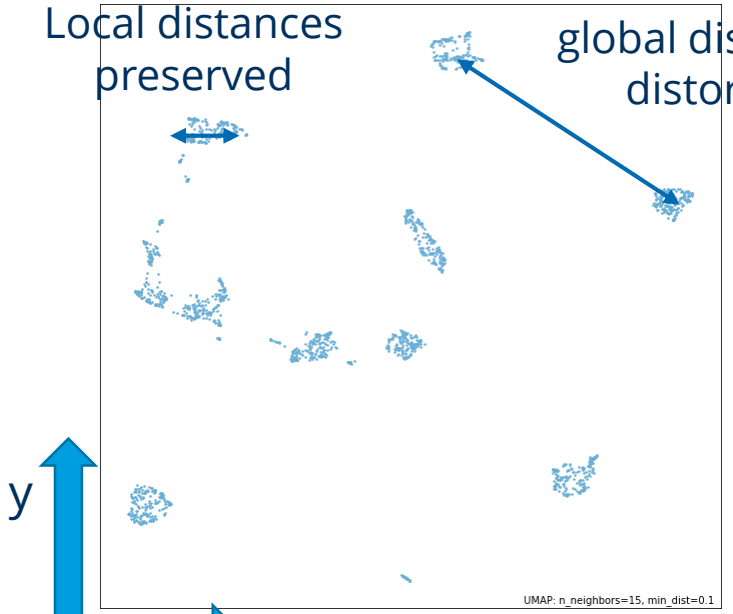
Challenge: Find a representation (embedding) of your data that represents the data in fewer dimensions

Preserve local distances at the expense of global distortions

x y z ...

	label	area	bbox_area	convex_area	quivalent_diamete	max_intensity	mean_intensity	min_intensity	solidity	extent	eret_diameter_ma	local_centroid-0	l
1	1	3379	13949	5120	18.61786412639...	613.0	345.6717963894...	259.0	0.6599609375	0....	37.3496987939662	15.77952056821...	18
2	2	2319	7448	3491	16.42230229224...	421.0	297.8434670116...	240.0	0....	0....	38.65229618017...	4....	17
3	3	2304	14415	4281	16.38681751812...	456.0	300.8298611111...	245.0	0....	0....	34.19064199455...	17.73828125	13
4	4	3278	13804	5139	18.43048549951...	467.0	316.1446003660...	249.0	0....	0....	34.84250278036...	15.52287980475...	10
5	5	1501	3315	1681	14.20563625190...	458.0	302.147235176549	236.0	0....	0....	17.97220075561...	6....	6.
6	6	2341	6061	2714	16.47407088948...	594.0	355.4446817599...	261.0	0....	0....	30.67572330035...	16.54250320375...	6.
7	7	1725	3584	1940	14.87979081163...	568.0	343.7866666666...	257.0	0....	0....	17.72004514666...	7.804637681159 42	7.
8	8	1502	3840	1753	14.20879025650...	431.0	290.0659121171...	235.0	0....	0....	18.57417562100...	8....	6.
9	9	1602	4080	1894	14.51737058294...	475.0	297.8008739076...	241.0	0....	0....	18.70828693386...	8....	8.
10	10	1395	3600	1624	13.86304166283...	424.0	304.8494623655...	247.0	0....	0.3875	17.60681686165...	7....	7.
11	11	609	1100	697	10.51654029260...	323.0	274.2528735632...	241.0	0....	0....	13.45362404707...	3....	4.
12	12	1686	3757	1894	14.76679738567...	460.0	303.8303677342...	240.0	0....	0....	17.97220075561...	9....	7.
13	13	2157	5184	2531	16.03062694504...	576.0	339.990264255911	270.0	0....	0....	19.54482028569...	8....	8.
14	14	863	2340	1032	11.81237949737...	327.0	272.4449594438...	237.0	0....	0....	16.0312195418814	6....	5.

Many dimensions



Few dimensions

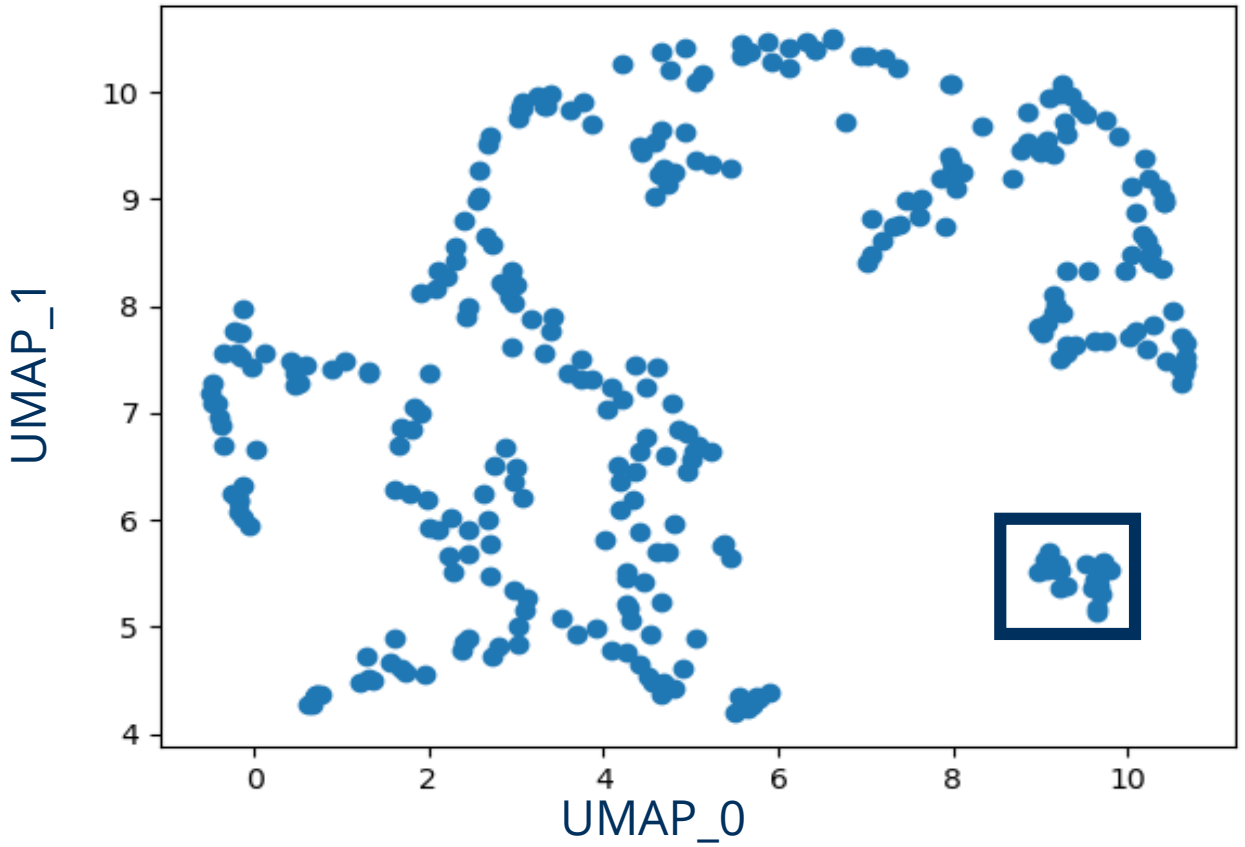
UNIFORM MANIFOLD UMAP APPROXIMATION & PROJECTION

https://umap-learn.readthedocs.io/en/latest/index.html

Clustering

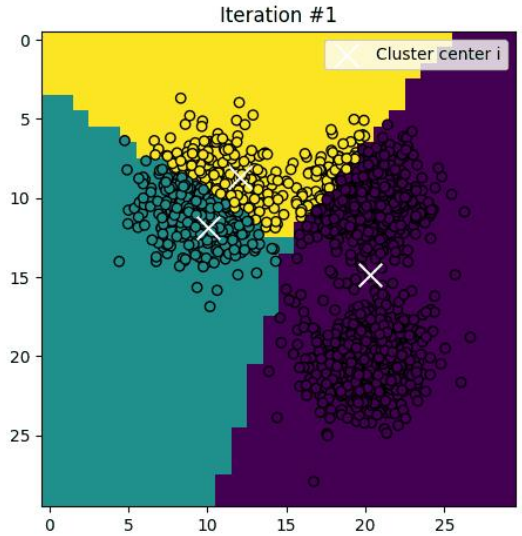
Starting point: Feature space or dimensionality reduction reveals “groups” in our data

Can we automatically identify these groups?



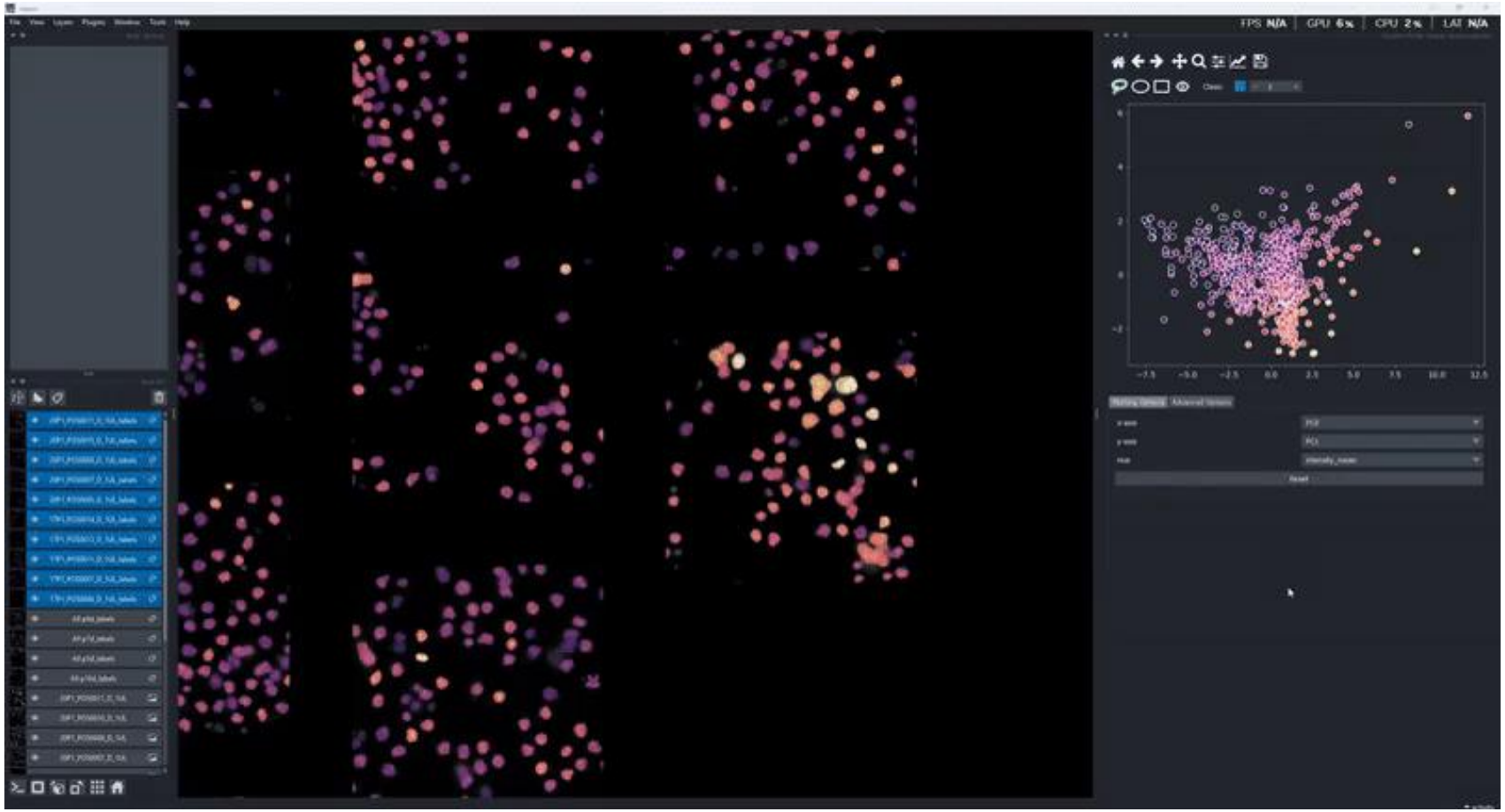
→ **Clustering** allows to stratify data into groups *without previous annotations*

K-means clustering



Data Exploration

... using interactive
feature visualization...




Laura Žigutytė
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Ryan Savill
@RyanSavill4

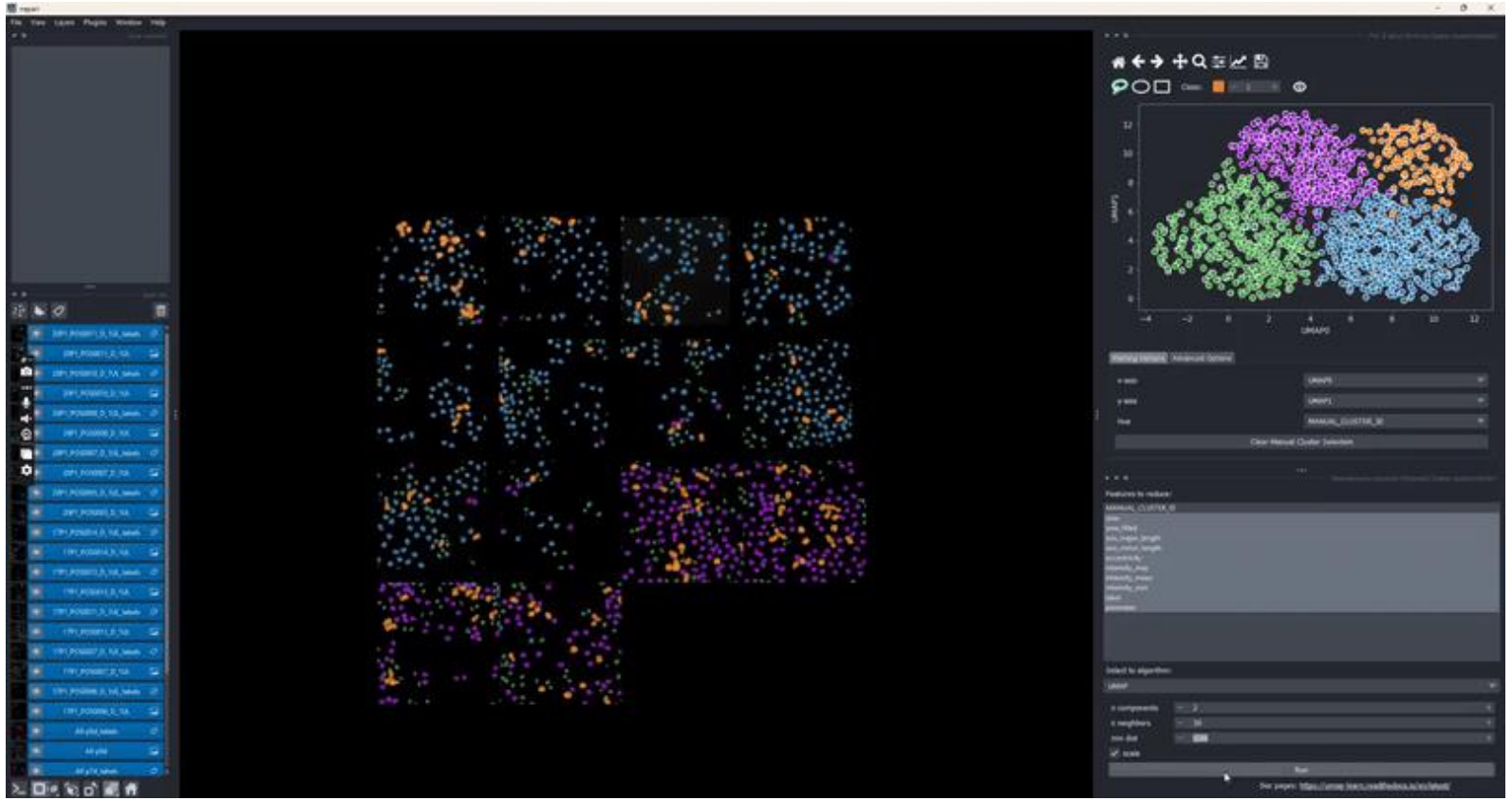

Johannes
Soltwedel


Marcelo
Zoccoler

https://github.com/BiAPoL/napari-clusters-plotter

Dimensionality Reduction and Clustering

… as well as interactive dimensionality reduction and clustering…



“How does this parameter affect the outcome of my analysis?”


Laura Žigutytė
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Ryan Savill
@RyanSavill4

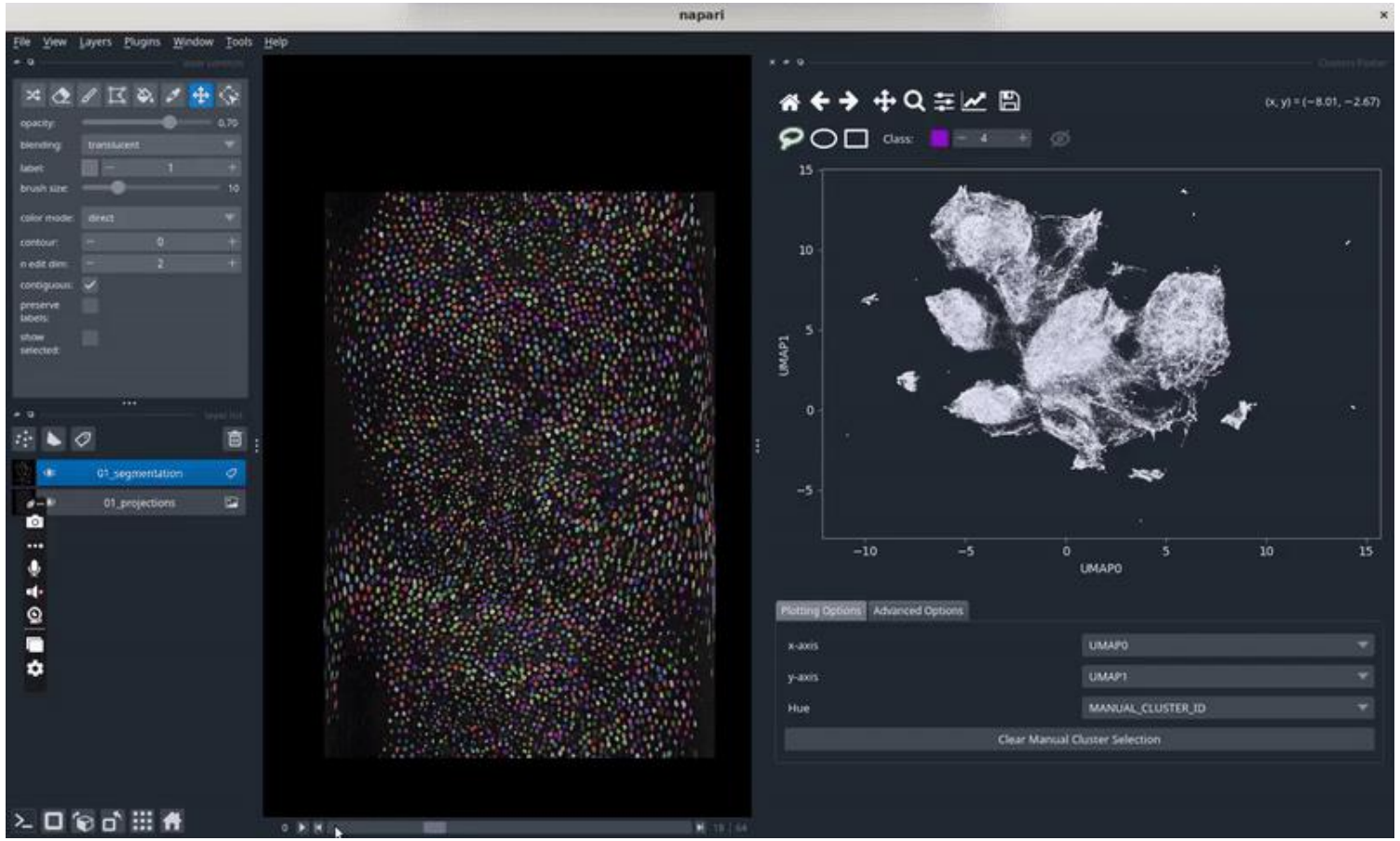

Johannes Soltwedel


Marcelo Zoccoler

https://github.com/BiAPoL/napari-clusters-plotter

Data Exploration

... and introspection!



"How are clusters in my data influenced by individual features?"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@zigutyte


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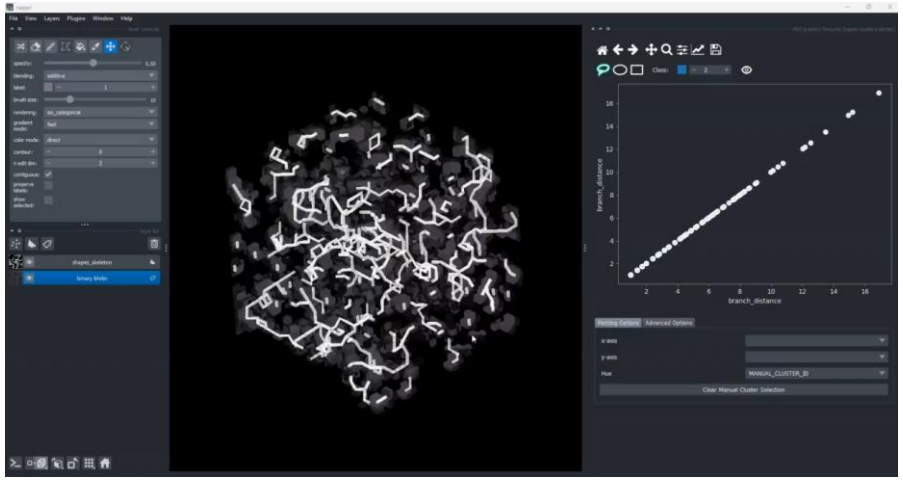
Johannes Soltwedel

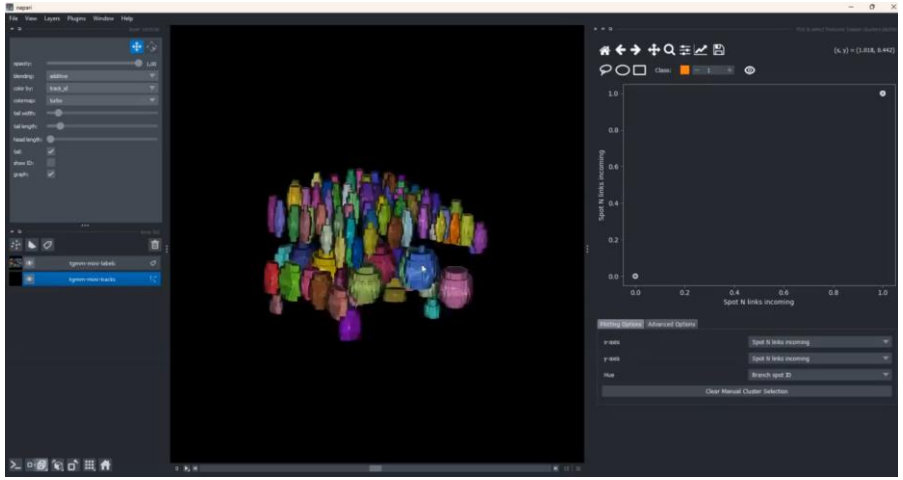

Marcelo Zoccoler

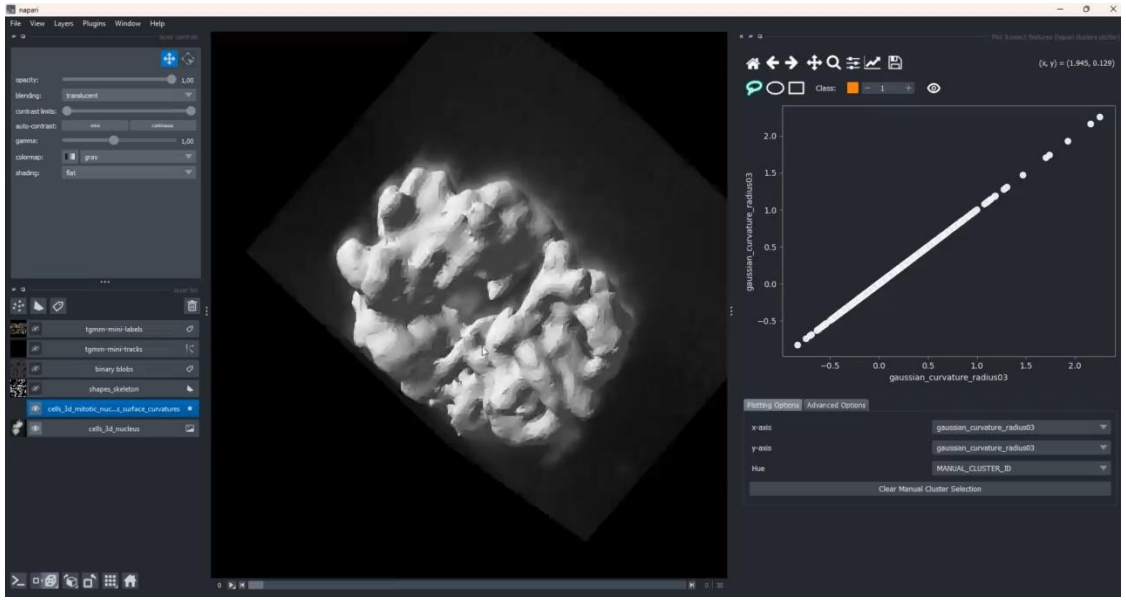
https://github.com/BiAPoL/napari-clusters-plotter

Data Exploration

Works for all sorts of layer types!





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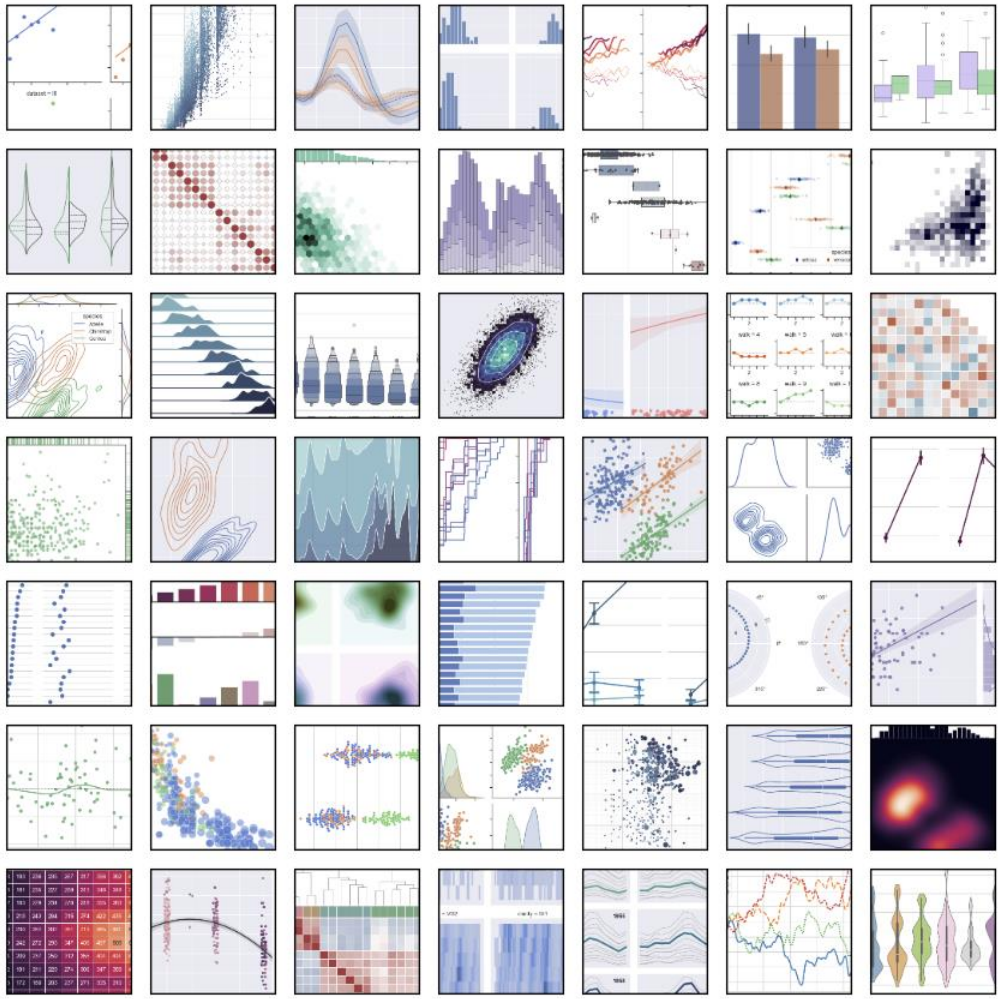
https://github.com/BiAPoL/napari-clusters-plotter

Scientific Plotting with Seaborn

- seaborn

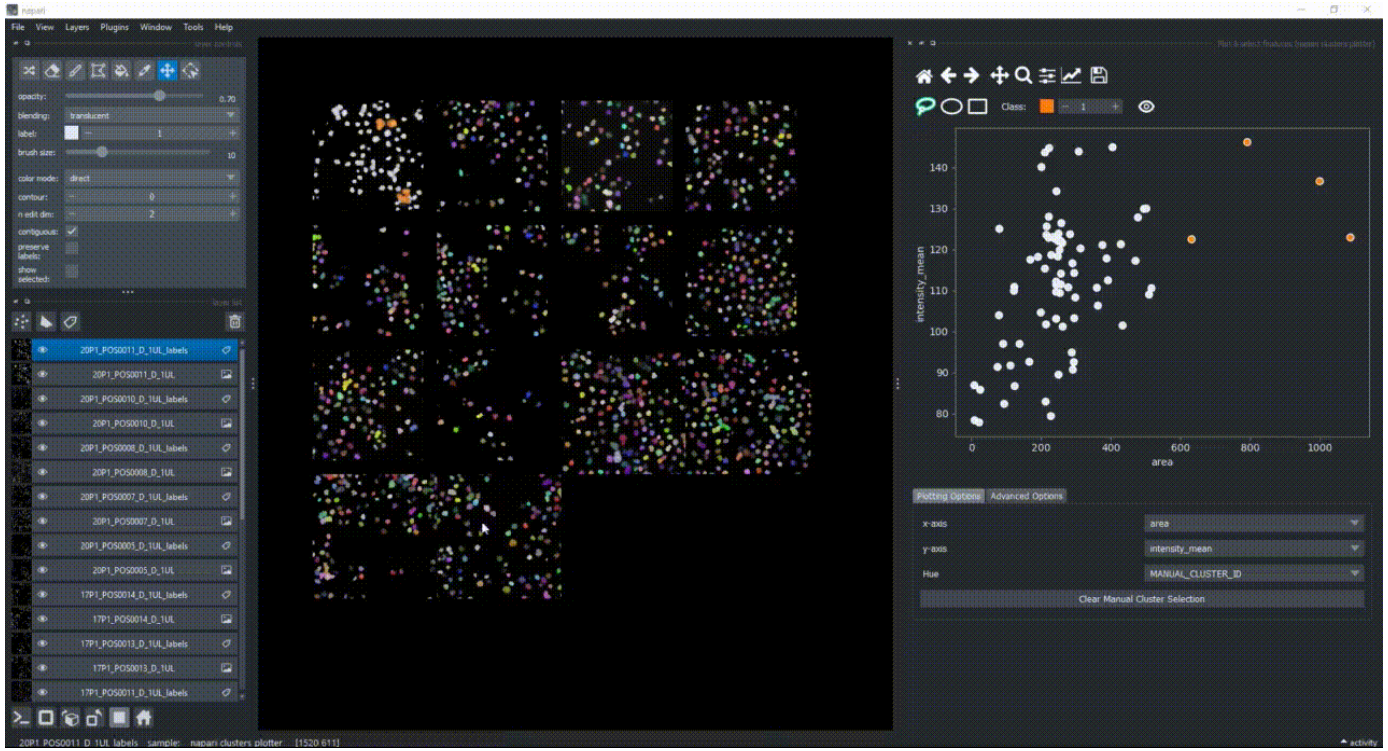
Seaborn gallery



https://seaborn.pydata.org/examples/index.html

Exercise: unsupervised object classification

Use Napari to classify objects (nuclei) without ground-truth



https://biapol.github.io/BioImage-Analysis-and-Data-Processing-Workshop-2025/interactive_unsupervised_object_classification/readme.html

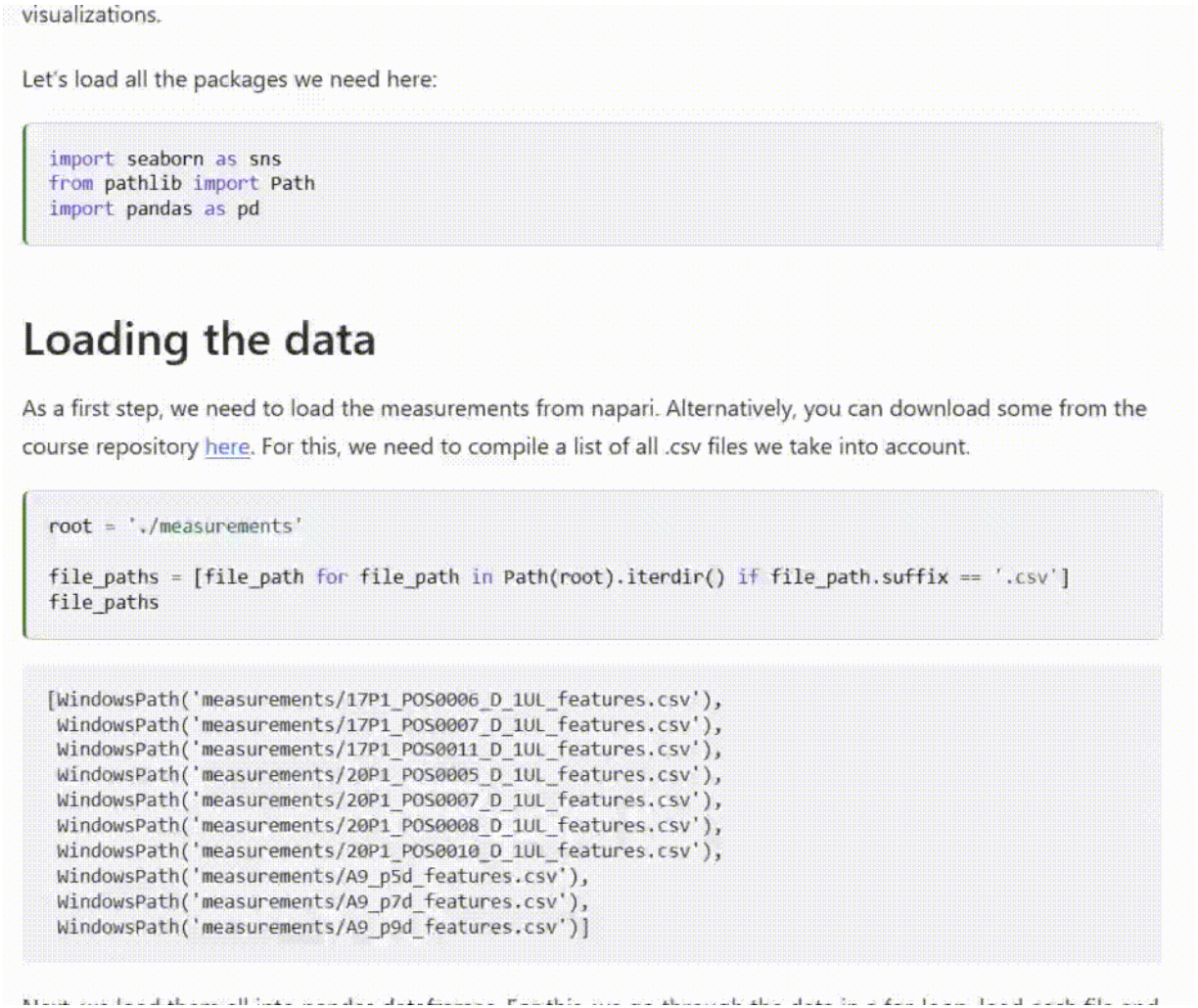
Exercise: scientific plotting with seaborn

1. Activate the environment
   `mamba activate napari25`

2. Navigate to your local repository
   `cd BioImage-Analysis-and-Data-Processing-Workshop-2025`

3. Start Jupyter Lab
   `jupyter lab`

4. Read and run the notebook, which can be found in docs > seaborn >seaborn_demo.ipynb



visualizations.

Let's load all the packages we need here:

```
import seaborn as sns
from pathlib import Path
import pandas as pd
```

Loading the data

As a first step, we need to load the measurements from napari. Alternatively, you can download some from the course repository here. For this, we need to compile a list of all .csv files we take into account.

```
root = './measurements'

file_paths = [file_path for file_path in Path(root).iterdir() if file_path.suffix == '.csv']
file_paths
```

```
[WindowsPath('measurements/17P1_POS0006_D_1UL_features.csv'),
 WindowsPath('measurements/17P1_POS0007_D_1UL_features.csv'),
 WindowsPath('measurements/17P1_POS0011_D_1UL_features.csv'),
 WindowsPath('measurements/20P1_POS0005_D_1UL_features.csv'),
 WindowsPath('measurements/20P1_POS0007_D_1UL_features.csv'),
 WindowsPath('measurements/20P1_POS0008_D_1UL_features.csv'),
 WindowsPath('measurements/20P1_POS0010_D_1UL_features.csv'),
 WindowsPath('measurements/A9_p5d_features.csv'),
 WindowsPath('measurements/A9_p7d_features.csv'),
 WindowsPath('measurements/A9_p9d_features.csv')]
```

Further resources:

Napari-hub:
www.napari-hub.org

BiaPoL image analysis course materials:

https://github.com/BiAPoL/Bio-image_Analysis_with_Python

Bio-image analysis materials:
https://bioimagebook.github.io/README.html

https://haesleinhuepf.github.io/BioImageAnalysisNotebooks/intro.html

Image Science Community Forum:
https://forum.image.sc/

Acknowledgements

PoL Physics of Life TU Dresden

BiAPoL team

- Marcelo Zoccoler
- Johannes Soltwedel
- Stefan Hahmann

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- Robert Haase
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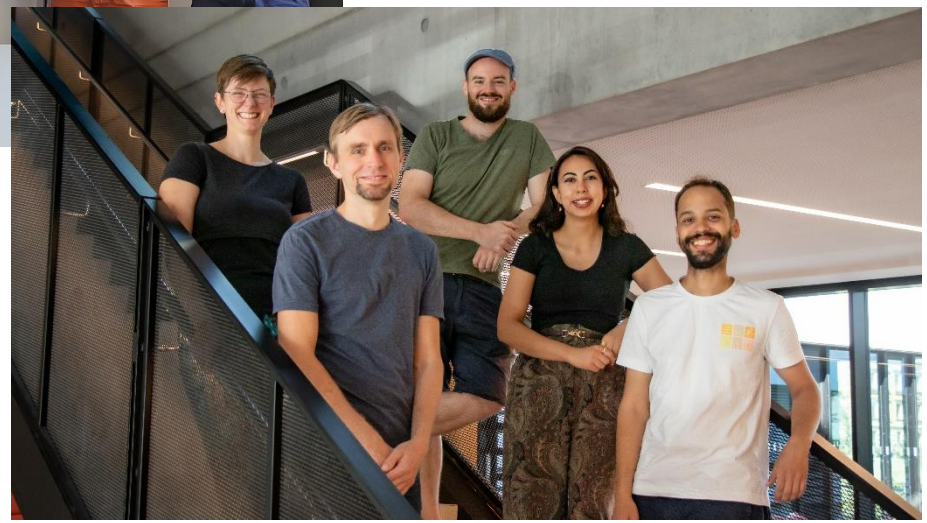
- Till Korten
- Mara Lampert
- Svetlana Iarovenko
- Ryan George Savill
- Laura Zigutyte
- Somashekhar Kulkarni
- Maleeha Hassan
- Tina Smejka



Networks

BioDIP Biopolis Dresden Imaging Platform

CENTER FOR SYSTEMS BIOLOGY DRESDEN

image.sc

NFDI4 BIOIMAGE

Funding

DFG public

@mazoc.bsky.social
@jo-soltwedel.bsky.social

https://physics-of-life.tu-dresden.de/bia

TECHNISCHE UNIVERSITÄT DRESDEN


Thank you!